

## SEROPREVALENCE OF LEPTOSPIROSIS AND ITS ASSOCIATION WITH REPRODUCTIVE AND PRODUCTIVE PARAMETERS FROM BUFFALO POPULATION OF RAJANPUR AND MUZAFFARGARH DISTRICTS OF PAKISTAN

M. Ijaz, A. Ghaffar, A. Ali, S. H. Farooqi, Y. R. Khan and A. I. Aqib

Department of Clinical Medicine and Surgery, University of Veterinary and Animal Sciences, 54600-Lahore  
Corresponding author Email: mijaz@uvas.edu.pk

The paper was presented in International Buffalo Congress 2019, February 18-20, Lahore, Pakistan

### ABSTRACT

Leptospirosis is one of the important infectious diseases adversely affecting the fertility and reproductive parameters of domestic animals. The current study was aimed to investigate the sero-prevalence of leptospirosis and its association with reproductive and productive parameters from buffalo population of Rajanpur and Muzaffargarh districts of Punjab, Pakistan. A total of 386 buffalo serum samples were analyzed for anti-*Leptospira* antibodies through indirect ELISA using "Anti-*Leptospira* IgG kit". The information regarding disease determinants were captured in questionnaire. Univariable and multivariable logistic regression models were used to correlate the potential risk factors with the disease dynamics. The results revealed an overall 12.69% prevalence of leptospirosis in both of the study districts, with higher percentage reported in district Rajanpur followed by Muzaffargarh presenting 20.72 and 04.66% prevalence at significant difference ( $p \leq 0.05$ ), respectively. The univariable analysis found district, estrus repetition, body condition score and history of retained placenta as significant candidates ( $p \leq 0.05$ ) while, multivariable logistic regression presented district, gender of animal and estrus repetition as the key risk factors (Odds Ratio >1). The study concluded that leptospirosis is prevalent in study districts and there is association of assumed risk factors with disease dynamics, which demands early installment of control strategies before the disease become unleashed.

**Key words-** Leptospirosis, Sero-prevalence, Risk factors, Buffalos.

### INTRODUCTION

Leptospirosis is one of the important infectious diseases which adversely affect the fertility and reproductive parameters of domestic animals (Saglam *et al.*, 2008). It is emerging infectious disease of livestock with zoonotic significance (Koet *et al.*, 2009). The etiological agent responsible for this disease occurrence is a pathogenic bacterium of genus *Leptospira* (Fornazari *et al.*, 2012). This disease affects not only the milk production of animals but also causes several reproductive disorders like stillbirth, abortion and birth of weak calves leading to tremendous financial losses to small holder farming communities (Sanhueza *et al.*, 2013; Ndengu *et al.*, 2017). Due to availability of favorable environment for the transmission of *Leptospira*, this disease is prevalent in developing countries (Bharti *et al.*, 2003). In bovines, this disease leads to abortion after several weeks' infection which is usually not associated with an obvious ailment (Bahari *et al.*, 2011). This disease has an adverse influence on the reproductive tract of bovines leading to compromised production and ultimate economic losses (Bomfim and Koury, 2006). The exact data regarding the economic losses due to the abortions from leptospirosis is limited. It is very difficult to determine the exact extent of economic losses attributed to leptospirosis. An outbreak of this malady in

Argentina for a period of 1 year resulted in losses of up to US \$ 150,000 for deaths of 100 calves, treatment and vaccination of 1300 survivors (Draghi *et al.*, 2011). While in France, the estimated cost of losses due to this disease in cattle herds was from US \$97 to 2611 per aborted cow (Ayril, 2013). However, the cost of problems besides abortion which are common and frequent has not yet been estimated. The climatic conditions such as warm temperatures, soil pH and the high rainfall, are favorable for the existence of *Leptospira* and also for the long term preservation of its pathogenicity (Martins and Lilenbaum, 2015; Andre-Fontaine *et al.*, 2015).

The definite diagnosis and identification of leptospirosis is by isolation technique but the detection by anti-leptospiral antibodies is also practiced. The isolation of bacteria is time consuming and it also demands livability of pathogen along with the ability of bacteria to grow on culture media, thus serological method is obliged for the detection purpose worldwide (Brandao *et al.*, 1998). Reported prevalence of leptospirosis in various animal species across different countries of world ranges between 2.00-46.00% (Leal-Castellanos *et al.*, 2003; Faria *et al.*, 2007). This wide variation might be related to several factors such as climate, animal species, time of the year, and method of investigation. Due to the presence of promising transmission circumstances,

leptospirosis infection is emerging health malady in developing countries (Ahmad, 2000). The transmission of infection has been reported by direct contact with uterine discharges, urine, placenta, and sexual contact with affected animals but indirect way of transmission of disease that involves the contact with contaminated environment (Bharti *et al.*, 2003).

In this context the current study was aimed to investigate the sero-prevalence of leptospirosis and its association with reproductive and productive parameters from buffalo population of Rajanpur and Muzaffargarh districts of Pakistan.

## MATERIALS AND METHODS

**Study design:** The study area of current project comprised of two flood hit districts—Muzaffargarh and Rajanpur (Figure 1) – of south Punjab province of Pakistan. The study districts are livestock rich and animals are the main source for livelihood of more than 2/3<sup>rd</sup> population. The district Muzaffargarh is having four tehsils, namely Muzaffargarh, Kotaddu, Alipur and Jatoi, whereas, district Rajanpur has three tehsils, namely Jampur, Rajanpur and Rojhan.

A total of 386 blood samples (193 from each study district) were collected aseptically from buffalos which were not having vaccination history against leptospirosis, by convenient sampling technique. From Muzaffargarh district sampling was conducted in such a way that 6 samples were collected from each village; from on tehsil four union councils were selected and from each union council two villages were further selected for sampling which comprised 192 blood samples, one additional sample was taken from kotaddu tehsil. Similarly from Rajanpur district 8 blood samples were collected from each village, from one tehsil four union councils were selected and from each union council two villages were further selected for sampling which comprised 192 blood samples, an additional sample was taken from Rojhan tehsil. The data regarding the risk factor analysis was collected in questionnaire.

**Blood sampling:** All the blood samples were collected aseptically from the jugular veins of buffalos individually and transferred to non-EDTA coated vacutainers separately. The collected blood samples were then placed undisturbed at room temperature for 30 minutes to clot formation. The samples were centrifuged within 2 hours of collection in order to prevent hemolysis and serum was decanted into sterile Eppendorf tubes by sterile technique. The collected serum samples were shipped to Medicine Laboratory Department of Clinical Medicine and Surgery, University of Veterinary and Animal Sciences (UVAS), Lahore maintaining the cold chain. The samples were kept at 4 °C till further processing.

**Processing of samples by serology:** The serum samples were processed by indirect (Enzyme linked immunosorbent assay) ELISA using (Alpha Diagnostic, international, USA) “RecomBac Bovine Anti-Leptospira LipL32 IgG kit” following the manufacturer instructions.

**Statistical analysis:** The data regarding the summary of risk factors was analyzed by logistic regression analysis. Firstly, all the study variables were analyzed by univariable analysis, the variables which produced  $P \leq 0.2$  were further analyzed by logistic regression analysis to find out the potential risk factors. The analysis was conducted on SPSS Version 20.00.

## RESULTS

The study has detected 12.69 % of the buffalo's positive for anti-leptospirosis antibodies in both of the study districts. The buffaloes of district Rajanpur presented higher prevalence (20.72%) as compared to district Muzaffargarh's (04.66%). Results have presented study districts to be statistically associated ( $p \leq .05$ ) with leptospirosis sero-prevalence, the buffaloes of Muzaffargarh district are at higher risk (OR = 8.464; CI = 3.307–21.665) towards *Leptospira* exposure than buffaloes of Rajanpur district. Gender of the animals was also found to be a key risk factor, having significant ( $p \leq .05$ ) impact on *Leptospira* sero-prevalence. The female buffaloes are more than two times as likely (OR = 2.009; CI = 0.470–8.584) to be seropositive than those to male buffaloes. Data analysis revealed that the buffaloes having history of estrus repetition are at higher risk (OR = 8.244; CI = 1.4849–46.899) as compared to those who are not having history of estrus repetition towards anti-leptospirosis antibodies.

The assumed risk factors like study district from which sampling was conducted, gender of animals, mastitis history, history regarding repetition of estrus, body condition score, history of retained placenta, previous history of any treatment given to animals, history of blood in milk and reduction in milk production, physiological and immune status of animal and abortion history were statistically analyzed (Table 1). To find out the association of the key risk factors with the seropositivity, all the risk factors were analyzed through univariable and multivariable logistic regression analysis. The gender of animal, history of mastitis, reduction in milk production, previous history of treatment given to the animals, status of animals, history of blood in milk and abortion history were not proved to be significant candidates towards anti-leptospirosis antibodies.

Initially, twelve variables were included in statistical analysis (Table 1), the variables which produced  $p \leq 0.2$  by univariable analysis were further analyzed by multivariable analysis to find out the potential risk factors, so six variables were included in

final multivariable analysis model (Table 2), from which three were found to be the potential risk factors towards leptospiral sero-positivity. So, multivariable logistic regression presented buffaloes from Muzaffargarh district, female and animals having history of repeat breeding to be associated with the occurrence of disease

dynamics. The history of retained placenta and immune status of the animals were proved to be significant candidates on the basis of univariable analysis but non-significant on the basis of multivariable logistic regression model.

**Table 1. Univariable analysis association and descriptive statistics in Buffaloes.**

Variable	Categories	Positive (%)	Negative	p-Value
District	Muzaffargarh	09 (04.66)	184	0.001*
	Rajanpur	40 (20.72)	153	
Gender	Female	44 (12.08)	320	0.180
	Male	05 (22.73)	17	
Mastitis	Yes	06 (08.00)	69	0.358
	No	10 (13.16)	66	
Estrus repetition	NA	33 (14.04)	202	0.001*
	Yes	08 (05.13)	148	
Body Condition Score	No	09 (09.47)	86	0.001*
	NA	32 (23.70)	103	
Reduction in milk production	Normal	41 (18.06)	186	0.001*
	Thin	07 (05.26)	126	
Previous history of treatment given	Emaciated	01 (03.85)	25	0.273
	Yes	27 (12.27)	193	
Animal status	No	14 (17.50)	66	0.600
	NA	08 (09.30)	78	
Immune status against major diseases	Yes	31 (13.42)	200	0.913
	No	18 (11.61)	137	
History of blood in milk	Pregnant	31 (13.25)	203	0.057
	Non pregnant	07 (12.28)	50	
Abortion history	NA	11 (11.58)	84	0.717
	Non Vaccinated	22 (09.91)	200	
History of retained placenta	Vaccinated	27 (16.46)	137	0.530
	Yes	06 (12.00)	44	
History of retained placenta	No	38 (12.33)	270	0.001*
	NA	05 (17.85)	23	
History of retained placenta	Yes	43 (12.11)	312	0.001*
	No	04 (18.18)	18	
History of retained placenta	NA	02 (22.22)	7	0.001*
	Yes	02 (15.38)	11	
History of retained placenta	No	13 (06.37)	191	0.001*
	NA	34 (20.12)	135	

\*  $p \leq 0.05$  = significant difference

**Table 2. Final Model of Multivariable logistic regression identifying the potential risk factors in the studied predictors.**

Variable	Category	S. E**	Wald	P-value	Odds Ratio	Lower C.I (95%)	Upper C.I (95%)
District	Muzaffargarh	.480	19.840	.000	8.464	3.307	21.665
	Rajanpur						
Gender	Female	.741	.887	.346	2.009	.470	8.584
	Male						
Estrus repetition	Yes	.887	5.656	.017	8.244	1.449	46.899
	No	.834	1.296	.255	2.585	.504	13.263
	NA						

Body Condition Score	Normal	4.730	.030	.094	.011	.792	1.086
	Thin	.455	.500	.470	.053	4.210	1.118
	Emaciated						
Immune status	Non Vaccinated	.516	2.379	.123	.451	.164	1.241
	Vaccinated						
History of retained placenta	No	.968	.692	.406	.447	.067	2.980
	Yes	1.168	3.157	.076	.126	.013	1.239
	NA						

\*\*Standard error

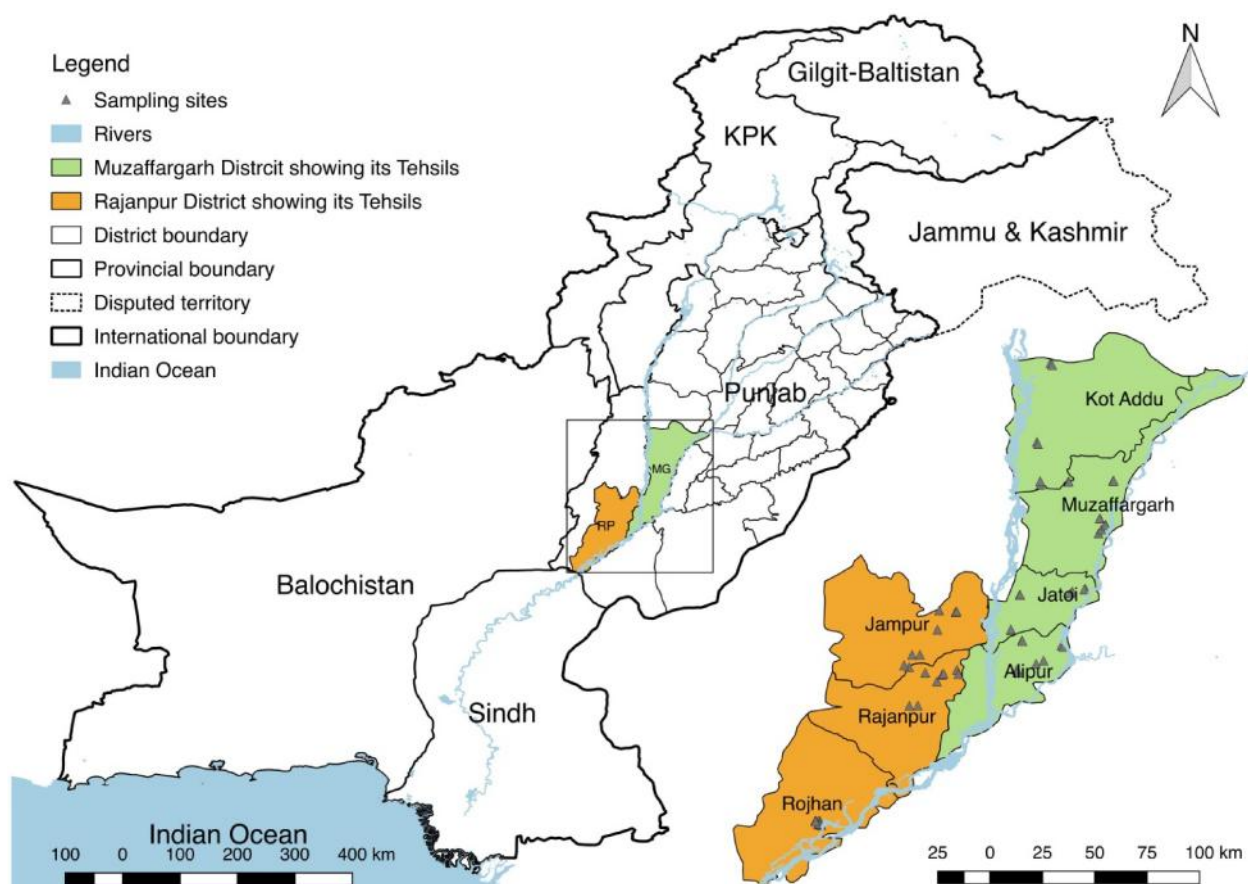


Figure 1. Map showing the study districts.

## DISCUSSION

Most of the infectious diseases are proved to be serious threat to the livestock (Yilmaz *et al.*, 2016; Elhaig *et al.*, 2016; Qayyum *et al.*, 2016). *Leptospira* infection frequently remains clinically silent but is found positive on serological assays (Hassanpour *et al.*, 2011).

*Leptospira*'s sero-prevalence has been reported in human as well as in domestic animals of Pakistan by rapid microscopic slide agglutination test (Ahmad, 1987). There are also few studies which have reported this pathogen in dogs (Saleem *et al.*, 2013a), horses (Sohail *et al.*, 2016), bovine (Ijaz *et al.*, 2018a, b; Islam *et al.*, 2019) and human population (Saleem *et al.*, 2013b; Sohail *et al.*, 2018) of Pakistan. This study shows the association

of *leptospira* sero positivity with reproductive parameters which is supported by Boqvist *et al.* (2002). This study was conducted in buffaloes of flood hit zones of Pakistan, and flood water is supposed to be positively associated with the occurrence of disease. These findings are justified by Bandino *et al.* (2015) and Sohail *et al.* (2016) as they proposed floods and rain fall both strongly associated with sero-prevalence of *Leptospira*.

The current study revealed 12.69% of the buffalo's positive for anti-leptospiral antibodies. The reported sero-prevalence of *leptospira* in buffaloes of Uganda (Atherstone *et al.*, 2014) and Malaysia (Bahaman *et al.*, 1987) is 42.39% and 31%, respectively which is quite higher than that of India in which prevalence was reported 2.8% (Schoonman and Swai, 2010). The

variation in prevalence might be due to distinct geographies, different management and husbandry practices and sampling strategies.

A significant difference was observed between the *Leptospira* sero-prevalence and study districts. In this study buffaloes of district Rajanpur presented higher prevalence of antileptospiral antibodies than the animals from Muzaffargarh district. These types of results are in agreement with the findings of Sohail *et al.* (2016) who also have reported significant difference of *Leptospira* sero-prevalence and study area in horses of Pakistan. The difference in prevalence in both of the study districts might be due to different production and rearing systems. Prevalence of leptospirosis was higher in buffaloes having history of abnormally repeating estrus. *Leptospira* infection is found to be strongly associated with estrus repetition in cattle (Libonati *et al.*, 2018). Female were found to be at higher risk of getting the disease as compared to males. These findings are also in line with findings of Sohail *et al.* (2016), who has reported higher sero-prevalence of *Leptospira* in female horses of Pakistan. It might be due to pregnancy stress or lactation stress in females.

Although non-significant in this study, but *Leptospira* infection has been found to be associated with abortions in animals (Prescott *et al.*, 1988). If reproductive system's infection is involved it could result in a "storm of abortions" causing considerable losses in form of meat and milk yield reductions (Tooloei *et al.*, 2008). Infertility, shown as delayed calving intervals and increased services per conception, is associated with this infection (Guitian *et al.*, 1999; Dhaliwal *et al.*, 1996). The exact pathogenesis is not clearly understood, however apparently the evidence for the presence of this pathogen in the oviduct and uterus of infected animals might interfere with the implantation of embryo or some other early pregnancy events (Grooms, 2006). The body condition score of animals was proved to be significant candidate on the basis of univariable analysis but was not found to be risk factor on the basis of final logistic regression model which resembled with the findings of Parvez *et al.* (2015).

The study concluded that the buffalo population of Rajanpur and Muzaffargarh districts is having considerable amount of anti-leptospiral antibodies and leptospirosis affects the animals silently. The study demands the scheduled vaccination program to control this zoonotic malady.

**Acknowledgements:** The authors acknowledge the Higher Education Commission (HEC) of Pakistan for funding the current project no. 20-4310/NRPU/R & D/HEC/14/593. The authors are thankful to Medicine Laboratory and Department of Clinical Medicine and Surgery, University of Veterinary and Animal Sciences,

Lahore for the provision of Laboratory facilities for smooth run of this project

**Conflict of interest:** Authors declare no conflict of interest.

## REFERENCES

- Ahmad, I (2000). Correlation of leptospiral, antibodies to the non-functional ovaries & biometrical observations in the slaughtered buffaloes & cattle. M.Sc Dissertation, Department of Theriogenology, College of Veterinary and Animal Sciences, Lahore. <http://agris.fao.org>. p96
- Ahmed, I.P (1987). Serological studies on leptospirosis in Pakistan. J. Pakistan Med. Assoc., 37(9): 233-236.
- Andre-Fontaine, G., F. Aviat and C. Thorin (2015). Waterborne leptospirosis: survival and preservation of the virulence of pathogenic *Leptospira* spp. in fresh water. Curr. Microbiol., 71: 136-142
- Anwar.K., N. Khan and M. Mujtabullah.(2013). Sero prevalence of leptospirosis in aborted dairy cattle in Peshawar district suburb, Khyber Pakhtunkhwa Pakistan. Int. J. Curr. Microbiol. Appl. Sci., 2(8): 73-78.
- Atherstone, C., K. Picozzi and G. Kalema-Zikusoka (2014). Sero-prevalence of *Leptospira* hardjo in cattle and African buffalos in southwestern Uganda. Am. J. Trop. Med. Hyg., 90(2): 288-290
- Ayral, F (2013). La leptospirose dans les cheptels bovins laitiers en France: impact économique de l'infection. Bulletin GTV 69: 61-67.
- Bahaman, A.R., A.L. Ibrahim and H. Adam (1987). Serological prevalence of leptospiral infection in domestic animals in West Malaysia. Epidemiol. Infect., 99(2): 379-392.
- Bahari, A., G. Abdollahpour, A. Sadeghi-Nasab, S. Sattari-Tabrizi, M. Yavari and B Dadmehr (2011). A serological survey on leptospirosis in aborted dairy cattle in industrial farms of Hamedan suburb, Iran. Iran J. Vet. Res., 12: 337-339.
- Bandino, J.P., A. Hang and S.A. Norton (2015). The infectious and noninfectious dermatological consequences of flooding: a field manual for the responding provider. Am. J. Clin. Dermatol., 16 (5): 399-424.
- Bharti, A.R., J.E. Nally, J.N. Ricaldi, M.A. Matthias, M.M. Diaz, M.A. Lovett, P.N. Levett, R.H. Gilman, M.R. Willig, E. Gotuzzo and J.M. Vinetz (2003). Peru-United States Leptospirosis Consortium. Leptospirosis: a zoonotic disease of

- global importance. *Lancet Infect. Dis.*, 3: 757-771
- Bomfim, M.R.Q and M.C. Koury (2006). Evaluation of LSSP-PCR for identification of *Leptospira* spp. In urine samples of cattle with clinical suspicion of leptospirosis. *Vet. Microbiol.*, 118: 278-288.
- Boqvist, S., H.T.V. Thu, I. Vågsholm and U. Magnusson (2002). The impact of *Leptospira* sero-positivity on reproductive performance in sows in southern Viet Nam. *Theriogenology.*, 58(7): 1327-1335.
- Brandão, A.P., E.D. Camargo, E.D. da Silva, M.V. Silva and R.V. Abrão (1998). Macroscopic agglutination test for rapid diagnosis of human leptospirosis. *J. Clin. Microbiol.*, 36(11): 3138-3142
- Dhaliwal, G.S., R.D. Murray, H. Dobson, J. Montgomery and W.A. Ellis (1996). Reduced conception rates in dairy cattle associated with serological evidence of *Leptospira interrogans* serovar hardjo infection. *Vet. Rec.*, 139:110–114
- Draghi, M.G., B. Brihuega, D. Benítez, J.M. Sala, G.M. Biotti, M. Pereyra, A. Homse and L. Guariniello (2011). Leptospirosis outbreak in calves from Corrientes Province, Argentina. *Rev. Argent. Microbiol.*, 43: 42–44.
- Elhaig, M.M., A. Selim, M.M. Mahmoud and E.K. El-Gayar (2016). Molecular confirmation of *Trypanosoma evansi* and *Babesia bigemina* in cattle from lower Egypt. *Pakistan Vet. J.* 36: 409–414.
- Faria, M.T., D.A. Athanazio, E.A.G. Ramos, E.F. Silva, M.G. Reis and A.I. Ko (2007). Morphological alterations in the kidney of rats with natural and experimental *Leptospira* infection. *J. Comp. Pathol.*, 137: 231-238.
- Fornazari, F., R.C. Da Silva, V.B. Richini-Pereira, H.E. Beserra, M.C. Luvizotto and H. Langoni (2012). Comparison of conventional PCR, quantitative PCR, bacteriological culture and the warthin starry technique to detect *Leptospira* spp. in kidney and liver samples from naturally infected sheep from Brazil. *J. Microbiol. Methods.*, 90: 321-326 .
- Grooms, D.L (2006). Reproductive losses caused by bovine viral diarrhoea virus and leptospirosis. *Theriogenology.*, 66(3): 624-628.
- Guitian, J., M.C. Thurmond and S.K. Hietala (1999). Infertility and abortion among first-lactation dairy cows seropositive or sero-negative for *Leptospira interrogans* serovar hardjo. *J. Am. Vet. Med. Assoc.*, 215: 515–518.
- Hassanpour, A., M. Imandar, G.R. Abdollahpour and M. Mahsayekhi (2011). Sero-prevalence of Leptospiral Infection in Ewes in Khoy, Iran. *Adv. Environ. Biol.*, 5: 2033-2038
- Ijaz, M., S. N. Abbas, S. H. Farooqi, A. I. Aqib, G.A. Anwar, A. Rehman, M.M. Ali, K. Mehmood and A. Khan (2018b). Sero-epidemiology and hemato-biochemical study of bovine leptospirosis in flood affected zone of Pakistan. *Acta.trop.*, 177: 51-57.
- Ijaz, M., S. N. Abbas, S. H. Farooqi, A. I. Aqib, P. Bakht, A. Ali, A. Ghaffar and S. Saleem (2018a). Sero-Epidemiology of Bovine Leptospirosis and Associated Risk Factors in a Flood Affected Zone of Pakistan. *Pakistan Vet. J.*, 38(2): 179-183.
- Islam, S., W. Shehzad, A. A. Bajwa, M. Imran, M. Y. Zahoor, M. Abdullah, M. I. Rashid, K. Ashraf, Y. F. Chang, A. Nadeem, M. Younas, S. A. M. Bukhari, M. M. Hassan, Z. I. Qureshi and R. Akhtar (2019). Molecular Detection of Brucellosis, Leptospirosis and Campylobacteriosis by Multiplex PCR and Screening by ELISA Assays in Buffalo Breeding Bulls. *Pakistan Vet. J.*, <http://dx.doi.org/10.29261/pakvetj/2019.092>
- Ko, A.I., C. Goarant and M. Picardeau (2009). *Leptospira*: the dawn of the molecular genetics era for an emerging zoonotic pathogen. *Nat. Rev. Microbiol.*, 7: 736-747.
- Leal-Castellanos, C.B., R. Carcia-Suarez, E. Gonzalez-Figueroa, J.L. Fuentes-Allen and J. Escobedo-De La penal (2003). Risk factors and the prevalence of leptospirosis infection in a rural community of Chiapas, Mexico. *Epidemiol. Infect.*, 131: 1149-1156
- Libonati, H.A., G.B. Santos, G.N. Souza, F.Z. Brandao and W. Lilenbaum (2018). Leptospirosis is strongly associated to estrus repetition on cattle. *Trop. Anim. Health and Prod.*, 50(7): 1625-1629.
- Martins, G and W. Lilenbaum, 2015. Comments of environmental conditions for the maintenance of *Leptospira* in tropical scenarios. *Curr. Microbiol.*, 71: 624–625
- Ndengu, M., M. Garine-Wichatitsky, D.M. Pfukenyi, M. Tivapasi, B. Mukamuri and G. Matope (2017). Assessment of community awareness and risk perceptions of zoonotic causes of abortion in cattle at three selected livestock-wildlife interface areas of Zimbabwe. *Epidemiol. Infect.*, 6: 1–16.
- Parvez, M.A., M.A.M., Prodhan, M.A., Rahman and M.R. Faruque (2015). Seroprevalence and associated risk factors of *Leptospira interrogans* serovar hardjo in dairy cattle of Chittagong, Bangladesh. *Pakistan Vet. J.*, 35(3): 350-354.
- Prescott, J. F., R.B. Miller, V.M. Nicholson, S.W. Martin and T. Lesnick (1988). Seroprevalence and

- association with abortion of leptospirosis in cattle in Ontario. *Can. J. Vet. Res.*, 52(2): 210
- Qayyum, A., J. A. Khan, R. Hussain, M. Awais, N. Ahmad and M.S. Khan (2016). Investigation of milk and blood serum biochemical profile as an indicator of subclinical mastitis in Cholistani cattle. *Pakistan Vet. J.*, 36: 275–279.
- Saglam, Y.S., Z. Yener, A. Tenur and E. Yalcin (2008). Immunohistochemical detection of leptospiral antigens in cases of naturally occurring abortions in sheep. *Small Rum. Res.*, 74: 119-122.
- Saleem, M. H., M.S. Khan, A.Z. Durrani, A. Hassan, M. Ijaz and M. M. Ali (2013b). Leptospirosis: An emerging zoonosis in Pakistan. *Pakistan J. Zool.*, 45(4)
- Saleem, M. H., M.S. Khan, M. A. Khan, M. A. Khan, M. Ijaz, A. Sardar and K. Mehmood (2013a). Sero surveillance of canine leptospirosis under different climatic conditions in and around Lahore, Pakistan. *Pakistan Vet. J.*, 33: 241-243.
- Sanhueza, J.M., C. Heuer, D. West (2013). Contribution of *Leptospira*, Neosporacanthium and bovine viral diarrhoea virus to fetal loss of beef cattle in New Zealand. *Prev. Vet. Med.* 112: 90 –98.
- Schoonman, L., and Swai, E. S. (2010). Herd-and animal-level risk factors for bovine leptospirosis in Tanga region of Tanzania. *Trop. Anim. Health and Prod.*, 42(7): 1565-1572.
- Sohail, M. L., M.S. Khan, M. Ijaz, O. Naseer, Z. Fatima, A.S. Ahmad and W. Ahmad (2018). Seroprevalence and risk factor analysis of human leptospirosis in distinct climatic regions of Pakistan. *Acta trop.*, 181: 79-83.
- Sohail, M.L., M.S. Khan, M. Avais, M.Y. Zahoor, M. Ijaz, A. Ullah, Z. Fatima, O. Naseer, I. Khattak and S. Ali (2016). Sero-prevalence of *Leptospira* spp. in Horses of Distinct Climatic Regions of Punjab, Pakistan. *J. Equine Vet. Sci.*, 44: 82-89.
- Tooloei, M., G. Abdollahpour, H. Karimi and A. Hasanpor (2008). Prevalence of serum antibodies against six *leptospira* serovars in sheep in Tabriz, North-western Iran. *J. Anim. Vet. Adv.*, 7: 450-455
- Yilmaz, R., I.T. Cangul, K. Onat, A. Akkoc, M.O. Ozyigit and E. Akdesir (2016). Histopathological, immunohistochemical and bacteriological characterization of *Mycoplasma bovis* pneumonia in cattle. *Pakistan Vet. J.*, 36: 316–321.