

MANAGEMENT OF *Meloidogyne incognita* DAMAGING LETTUCE: POTENTIAL OF CHROMATOGRAPHIC FRACTIONS AND EXTRACTS FROM *Lawsonia inermis*

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ABSTRACT

Plant parasitic nematodes (PPNs) are acknowledged as a paramount factor which limits the production of staple crops and vegetables. Preferred standard control method had been the use of synthetic nematicides. However, owing to the undesirable consequences of the residual effect of nematicides in the environment, the need for alternative approaches becomes pertinent and this has prompted investigation into the nematicidal potential of extracts from *Lawsonia inermis* for practicable application on lettuce plants infected with *Meloidogyne incognita* in field experiments. The leaves of *L. inermis* were collected and divided into four parts for separate extraction and these were compared with the standard nematicide carbofuran individually. The essential oil (EO) was significantly more potent than the other extracts from *L. inermis*. There was no significant difference between plants treated with carbofuran and EO. The fractions were significantly better than crude methanol and ethanol extracts of *L. inermis*. Reproduction of *M. incognita* on lettuce plant roots was considerably reduced by utilization of *L. inermis* extracts. Notably higher vegetative growth was observed in treated lettuce plants. The IR, ¹H-NMR and ¹³C-NMR spectral data analysis confirmed the presence of sesquiterpenes in the chromatographic fraction. The GC-MS profile indicated phytol as the major constituent of the EO. The results obtained from this study indicates that extracts from *L. inermis* could be a viable option in the management of *M. incognita* damaging lettuce in dependable vegetable production.

Keywords: bio-pesticide, carbofuran, chromatography, essential oil, nematode, pollution, vegetable.

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INTRODUCTION

Plant parasitic nematodes (PPNs) infection bring about adverse consequences on nearly all crops (Fabiyi *et al.*, 2018; Fabiyi, 2020; Fabiyi *et al.*, 2020a). PPNS induce a sizeable damage in agriculture (El-Eslamboly *et al.*, 2019; Fabiyi *et al.*, 2020b; Fabiyi, 2021), worldwide losses of about US\$78 billion in a year have been reported (Similey, 2005). Root-knot nematodes (RKNs) *Meloidogyne* spp. are the principal PPNS which affects yield in several crops (Taniwiryonoc *et al.*, 2009; Fabiyi, 2019; Fabiyi and Olatunji, 2021a). Largely, amidst the specie *Meloidogyne incognita* is a foremost pest of vegetables (Fabiyi and Olatunji, 2021b). Lettuce (*Lactuca sativa*), a leafy vegetable in the family Asteraceae grown for its leaves which are used in salad preparations is prone to *M. incognita* infection (William, 2012; de Souza Alonso *et al.*, 2022; Fabiyi, 2022a). Productivity of lettuce is limited by several pests and diseases. Infection by the root knot nematode *M. incognita* is a key factor which limits yield extensively in lettuce plants (Oliveira *et al.*, 2015; Koleva and Mitova, 2021).

Considerably high economic losses are attributed to *M. incognita* in lettuce production. Several

lettuce cultivars are known to be highly susceptible to *M. incognita*, with production losses of about 100% (Widmer and Abawi, 2000; Wilcken *et al.*, 2005; Rodrigues *et al.*, 2012). Symptoms exhibited by infected crops include chlorosis, stunting, withered head and root galling (Siddiqi *et al.*, 2001; Pinheiro *et al.*, 2013; de Souza Alonso *et al.*, 2022). Conventionally, the control of nematode pest of lettuce relies mostly on the use of synthetic nematicide like carbofuran. This is, however, unsafe particularly in a country like Nigeria where there are no implementable stringent regulations on pesticide residue limits in crops at harvest and farmers hardly read pesticide labels (Jatto *et al.*, 2012). Hence the probability of applying below or above the manufacturers recommended dosage for crops is very high.

The concern for public health and the environment has spurred interest in safer alternative methods of *M. incognita* management (Atolani *et al.*, 2014a; Atolani *et al.*, 2014b; Fabiyi *et al.*, 2020). There are quite a number of researches on plant extracts and purified compounds regarding their potential in the control of various pests and diseases (Javaid *et al.*, 2018; Khan *et al.*, 2020; Fabiyi, 2021b; Fabiyi, 2022b). Reports by Viaene and Abawi (1998) specified that amending soil

with Sudan grass notably reduced *M. hapla* reproduction on lettuce resulting in increase of lettuce head weight at harvest. Countless leading-edge studies have shown that extracts of various plant species such as *Chenopodium quinoa* (Khan and Javaid, 2020), *Euphorbia prostrata* (Ferdosi *et al.*, 2021a), *Bergenia ciliata* (Ferdosi *et al.*, 2021b), *Eucalyptus officinalis* (Fabiyyi, 2021c), *Khaya senegalensis* (Fabiyyi, 2022c) and diverse purified active principles contain potent nematocidal compounds (Atolani and Fabiyyi, 2020).

Lawsonia inermis is widely known for its cosmetic uses, the anti-bacterial activity against clinical bacteria isolates was documented by Arun *et al.* (2010) and Gull *et al.* (2013), the anti-fungal activity was also reported by Sharma and Sharma, (2011). Reports on the application of organic extracts and isolates from *L. inermis* in the control of PPNs particularly *M. incognita* is however scanty in literature, hence the necessity for this research. Consequently, this research aimed at evaluating isolated compounds and different solvent extracts from *L. inermis* as a possible option in the management of *M. incognita* pest of lettuce.

MATERIALS AND METHODS

Collection and preparation of plant materials:

Lawsonia inermis leaves were collected in large quantity from Sentu village, which is about 8 km north east of Ilorin town and were air dried for three weeks at ambient temperature in the laboratory. The leave samples were identified at the University of Ilorin herbarium unit. The dry leaves were divided into four parts of 1 kg each and were milled into tiny pieces to increase the surface area. One kilogram of the dry leaves was hydro-distilled for three hours (3 hrs). The yellow volatile oil yielded was separated from the aqueous layer with dichloromethane, the solution was then dried with anhydrous sodium sulphate (Na₂SO₄). Two parts were packed separately in aspirator and extracted with ethanol and methanol, the last part was used directly as soil amendment.

Column chromatography: The ethanol and methanol extracts were decanted, filtered and concentrated using a rotary evaporator. A part of the crude extract was chromatographed over silica gel 100-120 mesh grade (Fabiyyi *et al.*, 2012) in a glass column with an initial mobile phase of 100% n-hexane. The polarity was increased to n-hexane/dichloromethane ratio 2:1, n-hexane/ dichloromethane ratio 1:1, 100% dichloromethane, then finally dichloromethane/ ethanol ratio 2:1. The 100% dichloromethane elution afforded 21 fractions of 200 ml each. The fractions each were spotted on Thin Layer Chromatographic Plates (Al₂O₃, G_F-254 0.2mm, Merck, Germany). The spots were visualized using UV light (254 and 366 nm). All fractions which

gave the same retention factor (R_f) on the thin layer chromatographic plate (TLC), were then combined, concentrated and subjected to IR, H¹-NMR and ¹³C-NMR analysis. The EO obtained from the hydro distillation procedure was analysed by Gas Chromatography Mass Spectroscopy (GC-MS).

Instruments: IR spectra were recorded on 8400s Fourier Transform Infrared (FTIR). Nuclear magnetic resonance (¹H-NMR and ¹³C-NMR) was evaluated with JEOL 400 MHz. The chemical shifts were documented in ppm relative to TMS, while the coupling constants are in Hz. Gas Chromatography-Mass Spectroscopy was conducted on GCMS-QP 2010 PLUS (Shimadzu Japan) attached to finigan. The column is an RTX5MS with a MAT ion trap detector, which was packed with dimethylpolysiloxane at 100% grade. The condition of the GC-MS is as follows. Initial column temperature was 60°C and was held for 5 min, the injection volume was 1L. Temperature was programmed to rise at 5°C per minute up to 250°C, for injection the temperature was set to 200°C, while maintaining detector (mass spectrophotometer) temperature at 250°C. The carrier gas was helium operating at 46.3 cm/s linear velocity and 100.2 kPa pressure. Electron impact (EI) was the ionization mode set at 70 eV voltage. The peak enrichment technique for reference compounds was used for component identification, which finally confirmed the peaks identified by GC-MS. NIST library mass spectra was compared with the spectral data obtained.

Field experiments: Field experiments were conducted in 2017 and 2018 cropping seasons at the Teaching and Research Farm University of Ilorin, Nigeria (Lat 8° 29' N of the Equator; Long: 4° 40' E of the Greenwich Meridian). The experimental field was 30 m x 25 m in size, this was harrowed and seventy-two beds of 1.5 m² raised to a height of 15 cm each were made. The experiment was a randomized complete block design (RCBD). There were four dosages of application, treatments were six in number. Each treatment had three replicates. *L. sativa* 'Mindelo' seedlings raised from seeds were transplanted to the field at eighteen days after emergence from the nursery. Spacing was 20 cm apart and 50 cm between the rows. Pure culture of *M. incognita* (Kofoid and White, 1919) Chitwood, 1949 raised on *Celosia argentea* was extracted using the method of Hussey and Baker (1973), the extracted eggs and juveniles served as source of inoculums. Approximately 1000 eggs and juveniles per ml of *M. incognita* were inoculated around the base of each lettuce plant (Fabiyyi *et al.*, 2019) at four days after transplanting. The crude extracts, fractions and essential oil were applied a week after inoculation at 200 ml in variants of 50, 75 and 100 mg/ml, while the dry powder leaves used as soil amendment was applied at 150, 175 and 200 g. The reference standard check was carbofuran (a synthetic

nematicide) applied at 0.5, 1.0 and 1.5 kg/ai/ha. Fertilizer was applied at 30 kg/ ha⁻¹ a month after transplanting. Head diameter and numbers of leaves were recorded on weekly basis. Head weight, gall index, nematode population in the roots and rhizosphere soil of lettuce plants were evaluated after harvest. The gall index was evaluated on a scale of 0-4 (Hussey and Janssen, 2002), where 0 = no galls (root healthy), 1 = 1-5 small galls (galls on 1-25% of root system), 2 = 2-15 small galls (galls on 26-50% of root system), 3 = 16-25 galls, many part of roots functioning (galls on 56-75% of root system), 4 = >26 galls (galls on >76% of root system). All data were subjected to Analysis of variance using GenStat 5.32. Means were separated were necessary at 5% level of probability using the new Duncan's multiple range test.

RESULTS

The infra-red spectrum of the dichloromethane fraction exhibited diagnostic bands at 2958 and 2950 cm⁻¹ which is attributed to C-H stretch of aliphatic compounds. The presence of an aldehyde was observed at 2850 cm⁻¹,

while 1734 cm⁻¹ depicted the C=O of an ester. C-H of aromatics in the finger print region was noted at 828 and 730 cm⁻¹. The GCMS result of the essential oil depicted 11 compounds (Fig.1) and eight were more than 5%, phytol (21.02 %) had the highest percentage composition, this was closely followed by trans pulegol (15.03%), sabinene (13.05%), terpinolene (12.14%), terpinen-4-ol (11.05%) and limonene (9.49%), α -humulene (8.01%), gamma-eudesmol (5.03%), beta bisabolene (2.07%), caryophyllene (2.01%) and δ -cadinene (1.10%). The ¹H-NMR (400MHz CDCl₃) results depicted three protons in the olefinic region δ 5.0-5.1, four tertiary methyl groups were observed at δ 1.60 - δ 1.72. A singlet at 1.60 is attributed to three methyl groups; a multiplex peak at δ 1.95-2.14 is associated to nine protons. From the ¹³C-NMR spectrum, olefinic carbons were observed from δ 123.2-137.8, another chemical shift was seen at δ 15.0-38.6 which is for the aliphatic group, and these data are in consonance with that expected for sesquiterpenes. The activity of extracts and dosages of utilization of *L. inermis* and carbofuran on leaf number of lettuce is depicted in Fig 2, lettuce plants

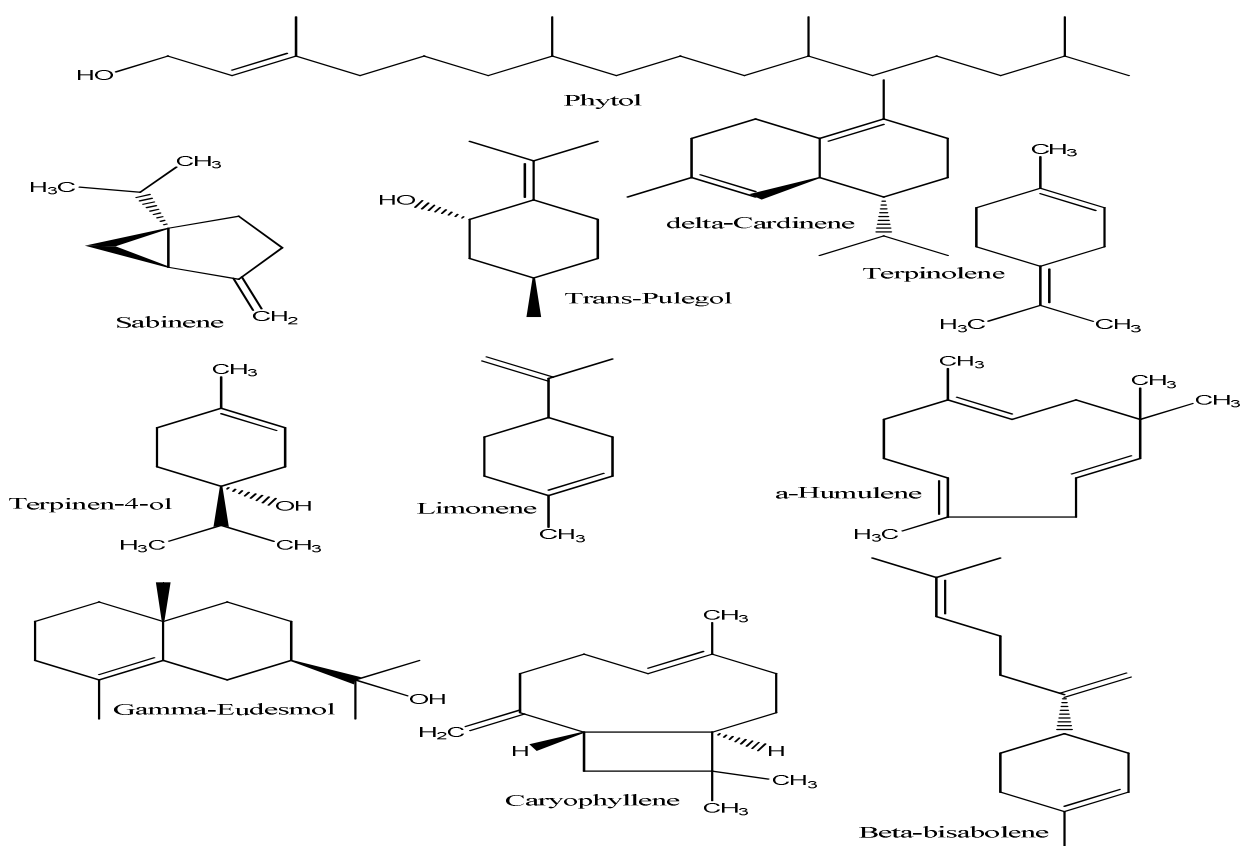


Figure 1: Structures of compounds obtained from the essential oil of *Lawsonia inermis*.

treated with essential oil and carbofuran (LSNI/EO and CBFN) had significantly ($p < 0.05$) more numbers of leaf,

leaf diameters were equally significantly wider with a corresponding heavier head weight at harvest (Figs. 3 &

4). Plants treated with powdered materials as soil amendments and crude extracts from ethanol and methanol extracts (LSNI/ODR, LSNI/EtOH and LSNI/MeOH) produced significantly fewer numbers of leaves, smaller head diameter and lighter head weight as opposed to the essential oil and carbofuran treated plants (Figs. 2, 3 & 4). The untreated control plants were not as healthy as the treated plants. Lettuce plants in beds amended with plant materials (LSNI/ODR) were significantly better in vegetative growth relative to the untreated control (0 mg/ml) plants. Nematode count in root and rhizosphere soil of untreated control plants were

significantly ($p < 0.05$) more compared to all treated plants (Figs. 5 & 6). A higher gall index was also recorded in the untreated control plants (Fig. 7). The variance in the quantity of extracts applied brought about a remarkable effect on the parameters evaluated. Vegetative growth of lettuce was directly proportional to the quantity of extracts and fractions applied. The highest (100 mg/ml) dosage of treatment produced more leaves with a wider diameter and heavier head weight, similarly nematode population was lower in soil and roots of plants treated with 100 mg/ml as opposed to the untreated control plants (Figs. 2, 3, 4, 5, 6 & 7).

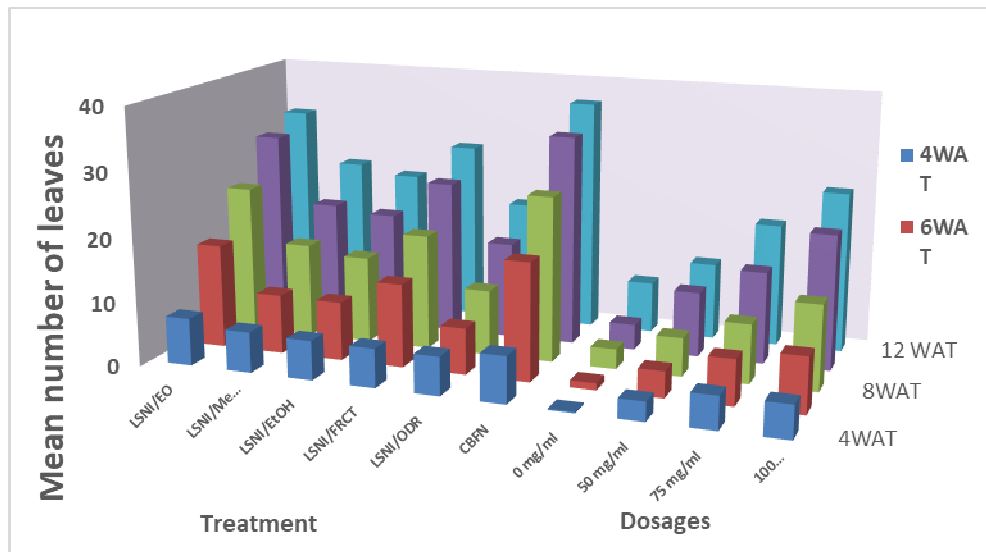


Figure 2: Effect of different treatments and dosages of *Lawsonia inermis* extracts on mean leaf number of lettuce plants

Key: LSNI/EO (*Lawsonia inermis* essential oil), LSNI/EtOH (*Lawsonia inermis* ethanol extract), LSNI/MeOH (*Lawsonia inermis* methanol extract), LSNI/FRCT (*Lawsonia inermis* Fraction) and CBFN (carbofuran).

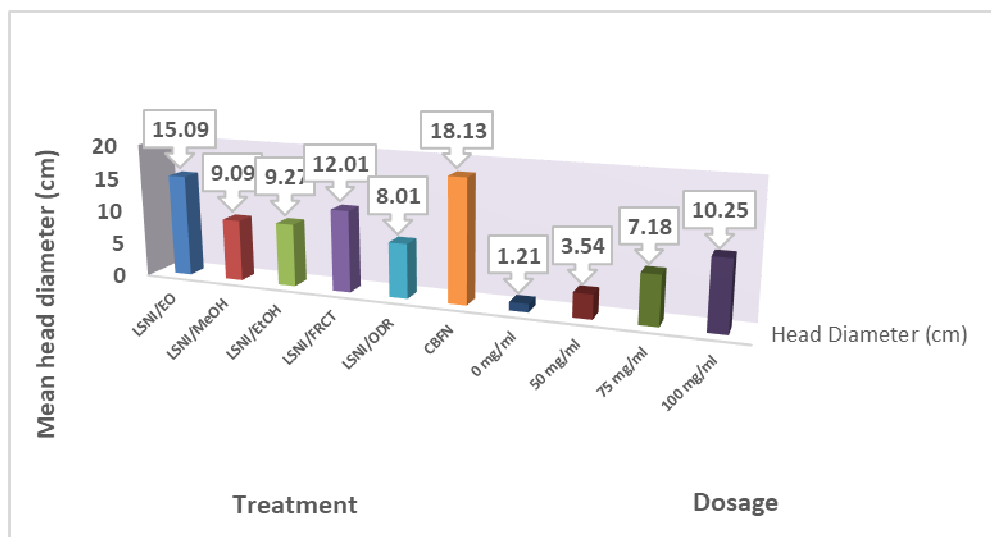


Figure 3: Effect of different treatments and dosages of *Lawsonia inermis* extracts on mean head diameter of lettuce plants.

Key: LSNI/EO (*Lawsonia inermis* essential oil), LSNI/EtOH (*Lawsonia inermis* ethanol extract), LSNI/MeOH (*Lawsonia inermis* methanol extract), LSNI/FRCT (*Lawsonia inermis* Fraction) and CBFN (carbofuran).

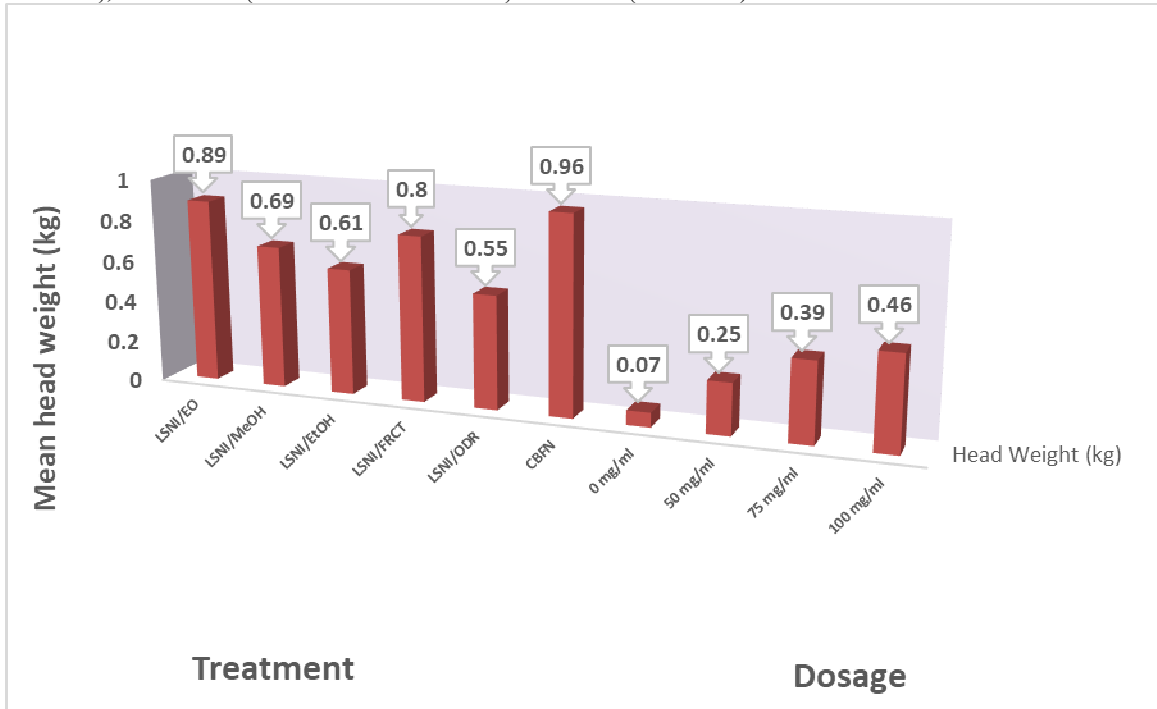


Figure 4: Effect of different treatments and dosages of *Lawsonia inermis* extracts on mean head weight of lettuce plants.

Key: LSNI/EO (*Lawsonia inermis* essential oil), LSNI/EtOH (*Lawsonia inermis* ethanol extract), LSNI/MeOH (*Lawsonia inermis* methanol extract), LSNI/FRCT (*Lawsonia inermis* Fraction) and CBFN (carbofuran).

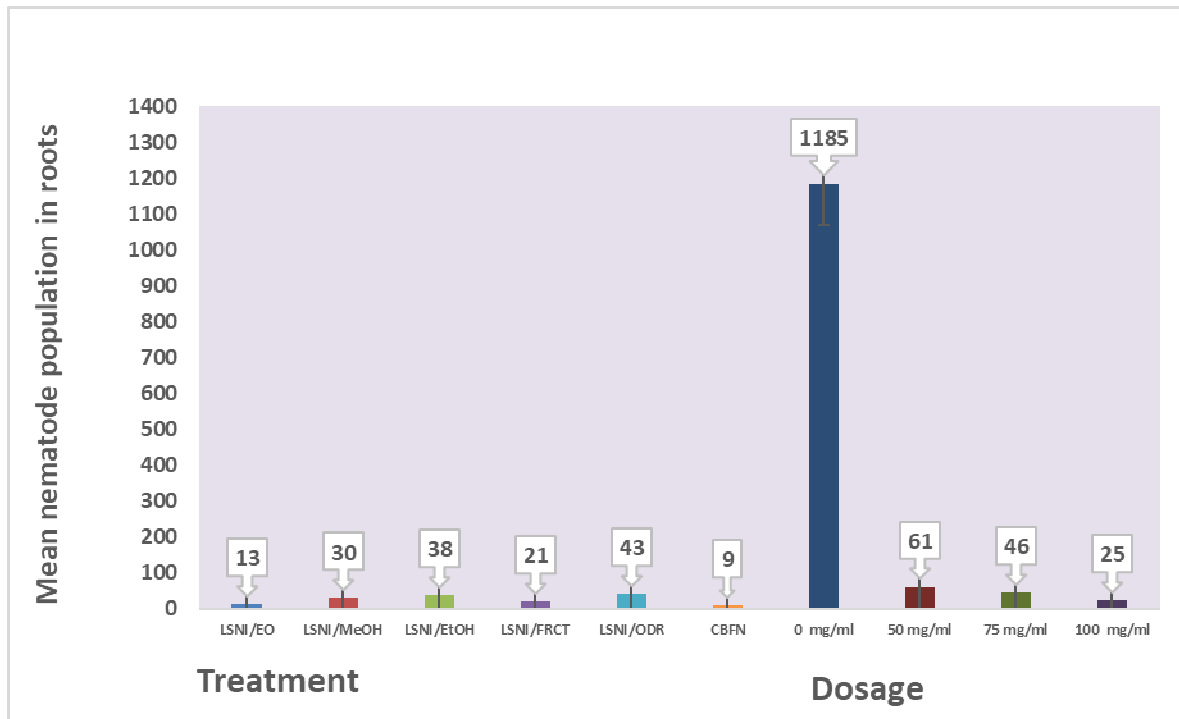


Figure 5: Effect of different treatments and dosages of *Lawsonia inermis* extracts on mean nematode population in roots of lettuce plants

Key: LSNI/EO (*Lawsonia inermis* essential oil), LSNI/EtOH (*Lawsonia inermis* ethanol extract), LSNI/MeOH (*Lawsonia inermis* methanol extract), LSNI/FRCT (*Lawsonia inermis* Fraction) and CBFN (carbofuran).

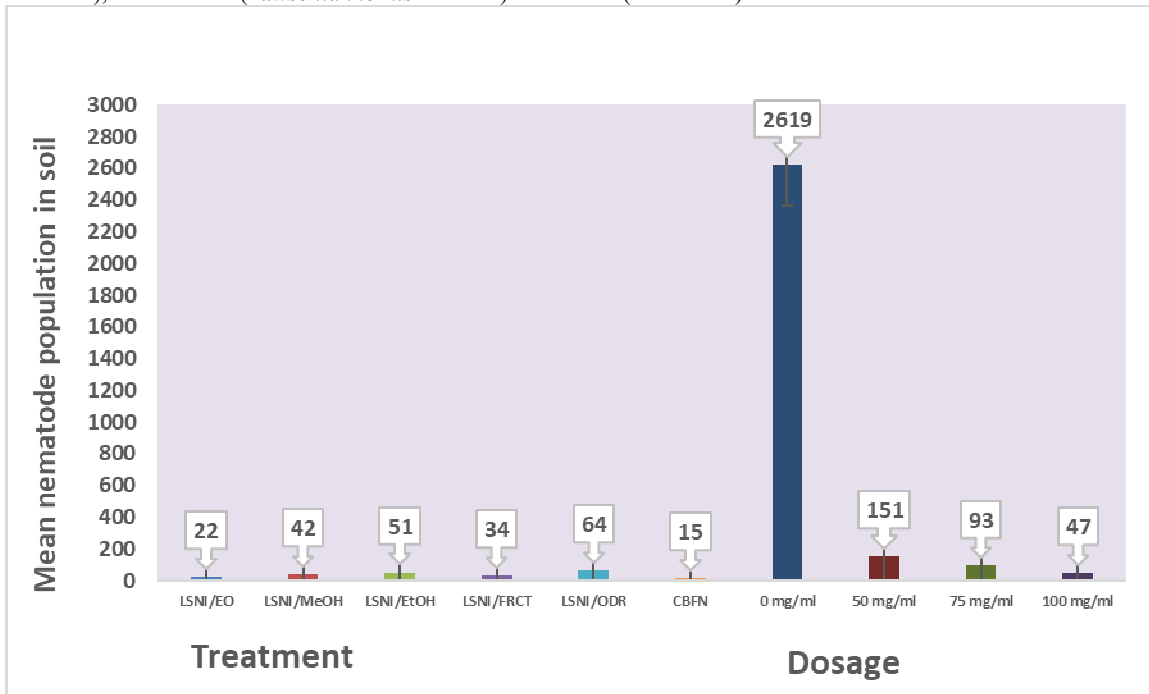


Figure 6: Effect of different treatments and dosages of *Lawsonia inermis* extracts on nematode population in rhizosphere soil of lettuce plants

Key: LSNI/EO (*Lawsonia inermis* essential oil), LSNI/EtOH (*Lawsonia inermis* ethanol extract), LSNI/MeOH (*Lawsonia inermis* methanol extract), LSNI/FRCT (*Lawsonia inermis* Fraction) and CBFN (carbofuran).

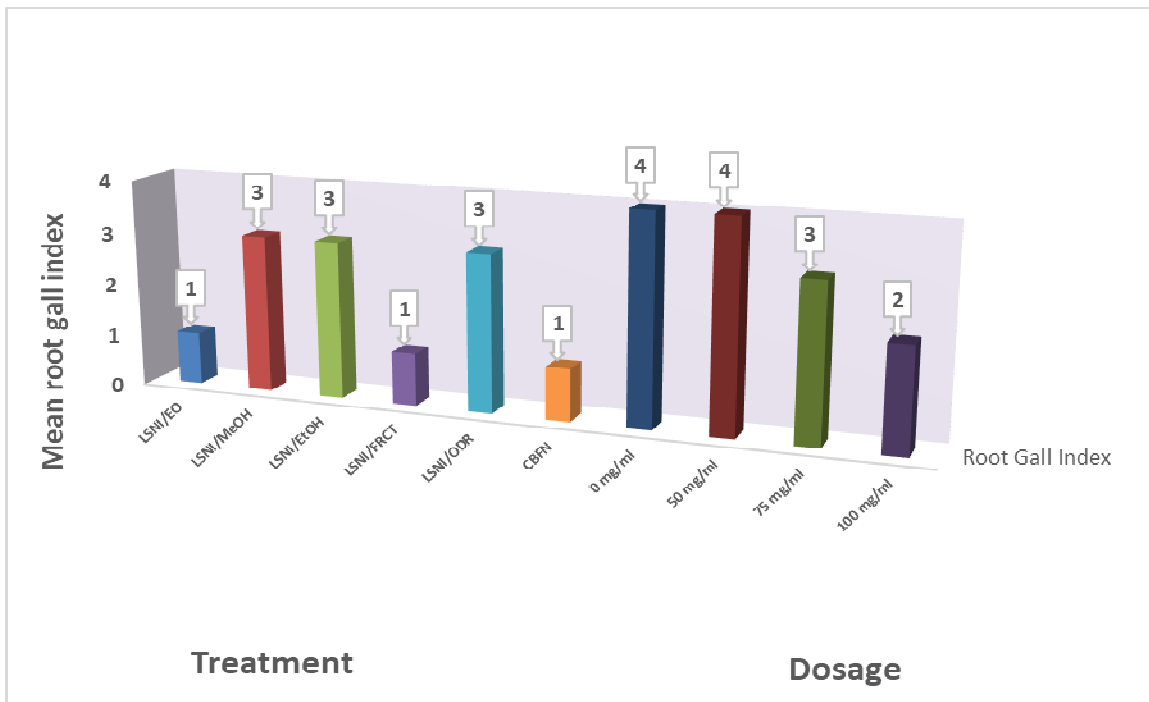


Figure 7: Effect of different treatments and dosages of *Lawsonia inermis* extracts on root gall index of lettuce plants

Key: LSNI/EO (*Lawsonia inermis* essential oil), LSNI/EtOH (*Lawsonia inermis* ethanol extract), LSNI/MeOH (*Lawsonia inermis* methanol extract), LSNI/FRCT (*Lawsonia inermis* Fraction) and CBFN (carbofuran).

DISCUSSION

The issue of environmental pollution brought about by pesticide use calls for caution in agricultural pest management. Several plant materials have been indicated as alternatives in the control option of plant parasitic nematodes (Chitwood, 2002; Fabiyi, 2021c) with positive outcomes. In this research the extracts from *L. inermis* has proven effective in the control of *M. incognita* damaging lettuce. The nematicidal action observed in this study could be attributed to the secondary metabolites contained in *L. inermis*. The constituents identified in this research aligns with the reports of Asmah *et al.* (2006) who also reported phytol as one of the constituents of EO from *L. inermis*, while the presence of sabinene, terpinolene and terpinen-4-ol was established by Najar and Pistelli (2017), as major composition of *L. inermis* EO. The nematicidal activity displayed by *L. inermis* essential oil (EO) was comparable to that of a synthetic nematicide. Results from ¹H-NMR and ¹³C-NMR indicates that the fractions contain sesquiterpenes. The accounts of El-Habashy *et al.* (2020) confirmed the efficacy of sesquiterpenes in *M. javanica* management in laboratory and greenhouse tests. Strong nematicidal activity was displayed with remarkable reduction in number of second stage juveniles of *M. javanica* in soil, egg masses per plant and number of galls at 500 mg/l. The vegetative parameters of eggplant were also increased in parallel with oxamyl a reference standard. In the laboratory, notable reduction in egg hatch of *M. javanica* was observed simultaneously with high juvenile mortality. *L. inermis* is known to be a highly potent plant and has been documented to be antiviral, antibacterial and antifungal (Najar and Pistelli, 2017). The tuberculostatic activity of *L. inermis* was reported by Sharma (1990), 6 µg/ml of extract inhibited H₃₇Rv *in vitro* and *L. inermis* extract at 5 mg/kg body weight of mice led to significant cure of mice infected with *M. tuberculosis* H₃₇Rv. Owing to the potency of *L. inermis* extracts, it has been indicated in the treatment of trypanosomiasis in livestock (Atawodi *et al.*, 2002). Identically, Okpekon (2004), reported that *L. inermis* was particularly effective among all plants evaluated for antihelminthic and trypanocidal activity. The ability of *L. inermis* to control *Tinea pedis* in humans was demonstrated by Sherifa *et al.* (2015). Fabiyi and Atolani (2011), reported the *in vitro* and screenhouse evaluation of aqueous extract and powdered material of *L. inermis*. Significant reduction in egg hatch of *M. incognita* was achieved with 15% crude extract concentration. Increased vegetative growth of *Corchorus olitorius* and nematode population reduction was also observed in the screenhouse at 15% concentration. The EO from *L. inermis* was exceptionally effective in comparison with

other extracts of *L. inermis*. A handful of study have confirmed the potency of essential oils in the management of *Meloidogyne* sp. Ozdemir and Gozel (2018) substantiated the findings in this study, they found essential oil from different plants to reduce the number of eggs masses and gall formation on tomato plant roots at 3 and 5% level of application. Comparably, Jardim *et al.* (2020) corroborated the potency of EO on *M. incognita* juveniles and eggs. They found 63 µg/ml of garlic essential oil significantly more active than 173 µg/ml of carbofuran on eggs and second stage juveniles of *M. incognita*. The reproduction and infectivity of *M. incognita* on tomato plants was remarkably reduced with application of 0.2 ml of garlic EO per litre of substrate. The effectiveness of garlic EO was statistically the same as 0.25 g of dazomet. Likewise, Pardavella *et al.* (2020a) evaluated essential oil extracted from *Cuminum cyminum* (cumin) seeds on survival, hatching and motility of second stage juveniles and eggs of *M. incognita* and *javanica*. At 62.5 µl/l concentration, the J2s were paralyzed, while hatching of eggs decreased with increase in EO concentration. In the same vein, Pardavella *et al.* (2020b) recorded significant reduction in nematode population in infested soil and roots of tomato plants after application of EO from *Satureja hellenica* at 4000µl/l, while 100% paralysis of *M. incognita* and *javanica* J2s was noted after 96 hours of exposure. They confirmed that the EO of *S. hellenica* contains 4-terpineol (3.65%), γ-terpinene (4.63%), carvacrol methylether (6.77%), carvacrol (23.25%), borneol (6.79%) and p-cymene (27.46%) as active principles. The use of EO from *L. inermis* promoted vegetative growth in lettuce; hence it can be employed in the control of *M. incognita* in lettuce, thus reducing the unnecessary use of pesticides in lettuce production.

Conclusion: In this study, the leaves of *L. inermis* was extracted in different ways whereby, the EO was found to be significantly more potent than other extractives. The active chemical compounds in the EO could be isolated individually and considered for proper formulation into commercial use to minimise the challenges associated with *M. incognita* management. Dry leaves of *L. inermis* can also be incorporated into the soil to bring down *M. incognita* population in lettuce cultivation.

REFERENCES

- Atolani, O., O. A. Fabiyi and G.A. Olatunji (2014a). Isovitexin from *Kigelia pinnata*, A Potential Eco-friendly Nematicidal Agent. Trop. Agric. 91(2): 67-74.
- Atolani, O., O. A. Fabiyi and G.A. Olatunji (2014b). Nematicidal Isochromane Glycoside from *Kigelia pinnata* leaves. Acta Agric. Slov. 104 (1): 25-31.

- Atolani, O. and O. A. Fabiyi (2020). Plant Parasitic Nematodes Management Through Natural Products: Current Progress and Challenges. In Management of Phytonematodes: Recent Advances and Future Challenges. Ansari, R. A., R. Rizvi and Mahmood, I. (Eds). 297-315. Singapore. https://doi.org/10.1007/978-981-15-4087-5_13
- Arun, P., K. G. Purushotham, J. Johnsy, K. Vasantha and D. Chamundeeswari (2010). Screening antibacterial activity of various extracts of *Lawsonia inermis*. Res. J. Pharm Phytochem. 2(3):103-8
- Asmah, R., S., Edrini, P., Ismail, T.Y. Yun Hin, and M.F. Abu bakar (2006). Chemical constituents, antioxidant activity and cytotoxic effects of essential oil from *Strobilanthes crispus* and *Lawsonia inermis*. J. Bio. Sci. 6(6):1005-1010. DOI: 10.3923/jbs.2006.1005.1010
- Atawodi, S.E., D.A., Ameh, S., Ibrahim, J.N., Andrew H.C., Nzelibe, E.O., Onyike, K.M., Anigo, E.A., Abu, D.B., James, G.C., Njoku, and A.B. Sallau (2002). Indigenous knowledge system for treatment of trypanosomiasis in kaduna state of Nigeria. J. Ethnopharm. 79(2):279-82. doi: 10.1016/s0378-8741(01)00351-8
- CABI Crop Protection Compendium. (2008). *Lactuca sativa* (lettuce) datasheet. Available at: <http://www.cabi.org/cpc/datasheet/29609>. [Accessed 8 February 17 2022]. <https://doi.org/10.1079/cabicompendium.29609>
- Chitwood, D.J. (2002). Phytochemical based strategies for nematode control. Annual Review of Phytopathology. 40:221-249 doi: 10.1146/annurev.phyto.40.032602.130045.
- de Souza Alonso, T.A., D.L., da Silva, R., de Mello Prado, P.L Martins Soares, L.F.L, Tenesaca, and R.J. Ferreira (2022). Silicon promotes the control of *Meloidogyne incognita* in lettuce by increasing ascorbic acid and phenolic compounds. J. Pest. Sci .95, 1453-1466. <https://doi.org/10.1007/s10340-021-01470->
- El-Eslamboly, A. A.S.A. Mona M. Abd El-Wanis1 and A. W. Amin (2019). Algal application as a biological control method of root-knot nematode *Meloidogyne incognita* on cucumber under protected culture conditions and its impact on yield and fruit quality. Egypt J. Biol. Pest Control. 29: 18. <https://doi.org/10.1186/s41938-019-0122-z>
- El-Habashy, D.E., M.A., Abdel Rasoul and S.A.M. Abdelgaleil (2020). Nematicidal activity of phytochemicals and their potential use for the control of *Meloidogyne javanica* infected eggplant in the greenhouse. Eur. J Plant Pathol. 158: 381-390. <https://doi.org/10.1007/s10658-020-02079-6>.
- Fabiyi, O. A. and O. Atolani (2011). *Lawsonia inermis* in the control of *Meloidogyne* spp. on *Corchorus olitorus*. Electronic J. Environ Agric. Food Chem. 10(3):2000-2006.
- Fabiyi, O.A., G.A. Olatunji and A.O. Saadu (2018). Suppression of *Heterodera sacchari* in rice with agricultural waste-silver nano particles. J. Solid Waste Technol. Manag. 44(2): 87-91. DOI: <https://doi.org/10.5276/JSWTM.2018.87>
- Fabiyi, O.A. (2019). Management of Groundnut (*Arachis hypogea*) Root-knot nematode (*Meloidogyne incognita*): Effect of *Prosopis africana* Pods. Indian J. Nematol. 49(2): 214-216.
- Fabiyi, O.A., G.A., Olatunji and I.O. Daodu (2019). Nematicidal effect of organic extract metal complex on *Meloidogyne incognita* infecting groundnuts (*Arachis hypogea*). Sci. Agric. Bohem. 50(3): 191-196. DOI: 10.2478/sab-2019-0026
- Fabiyi O. G., Olatunji, O., Atolani, and O. Olawuyi (2020a). Preparation of bio-nematicidal nanoparticles of *Eucalyptus officinalis* for the control of cyst nematode (*Heterodera sacchari*). J. Anim. Plant Sci. 30(5).1172-1177.
- Fabiyi, O. A. O., Atolani, and G.A. Olatunji (2020b). Toxicity Effect of *Eucalyptus globulus* on *Pratylenchus* spp of *Zea mays*. Sarhad J. Agric. 36(4): 1244-1253.
- Fabiyi, O.A., O.D., Saliu, A.O., Claudius-Cole, I.O., Olaniyi, O.V., Oguntebi, O.V and G.A. Olatunji (2020c). Porous starch citrate biopolymer for controlled release of carbofuran in the management of root knot nematode *Meloidogyne incognita*. Biotechnology Reports. 25(e00428): 1-9. <https://doi.org/10.1016/j.btre.2020.e00428>.
- Fabiyi, O. A. (2020). Growth and yield response of groundnut *Arachis hypogaea* (Linn.) under *Meloidogyne incognita* infection to furfural synthesized from agro-cellulosic materials. J. Trop. Agric. 58(2):241-245.
- Fabiyi, O.A. (2021a). Application of furfural in sugarcane nematode pest management. Pakistan J. Nematol. 39(2): 151-155. doi.org/10.17582/journal. PjN /2021.39.2.151.155. DOI: 10.17582/journal.pjn/2021.39.2.151.155
- Fabiyi, O.A. (2021b). Evaluation of Nematicidal activity of *Terminalia glaucescens* fractions against *Meloidogyne incognita* on *Capsicum chinense*. J Hort. Res. 29(1): 67-74. DOI: 10.2478/johr-2021-0006

- Fabiyyi, O.A. (2021c). Evaluation of plant materials as root-knot nematode (*Meloidogyne incognita*) suppressant in okro (*Abelmoscous esculentus*). Agric. Conspec. Sci. 86(1):51-56.
- Fabiyyi, O. A. and G.A. Olatunji (2021a). Toxicity of derivatized citrulline and extracts of water melon rind (*Citrullus lanatus*) on root-knot nematode (*Meloidogyne incognita*). Trop. Agric. 98(4): 347-355.
- Fabiyyi, O. A. and G.A. Olatunji (2021b). Environmental Sustainability: Bioactivity of *Leucaena leucocephala* Leaves and Pesticide Residue analysis in Tomato Fruits. Acta Univ. Agric. Silv. Mend. Brunensis. 69 (4):473-480. DOI: 10.11118/actaun.2021.042
- Fabiyyi, O.A. (2022a). Evaluation of weeds against root-knot nematode (*Meloidogyne incognita*) in vegetables. Sarhad J. Agric. 38(4): 1289-1299. DOI: 10.17582/journal.sja/2022/38.4.1289.1299
- Fabiyyi, O.A. (2022b). Fractions from *Mangifera indica* as an Alternative in *Meloidogyne incognita* Management. Pakistan J. Nematol. 40(1): 65-74. DOI:10.17582/journal.pjn/2022/40.1.65.74
- Fabiyyi, O.A. (2022c). Cytotoxicity and Nematicidal Potential of Leaf Extracts of *Adansonia digitata* and *Khaya senegalensis* on Root Knot Nematode (*Meloidogyne incognita*) Associated with Cabbage (*Brassica oleracea*). J. Agric. Sci. -Sri Lanka. 17(3):425-436. DOI: 10.4038/jas.v17i3.9922
- Ferdosi, M.F.H, I.H., Khan A., Javaid, M., Nadeem. and A. Munir (2021a). Natural pesticidal compounds of *Euphorbia prostrata*. Pakistan J. Phytopathol. 33(2): 349-355.
- Ferdosi, M.F.H, I.H., Khan, A., Javaid, H.M., Saeed, I., Butt, and A. Munir (2021). GC-MS analysis and bioactive components of flowers of *Bergenia ciliata*, a weed of rock crevices in Pakistan. Pakistan J. Weed Sci. Res. 27(4): 527-535. DOI:10.28941/pjwsr.v27i4.1012
- Gull Iram, Maria Sohail, Muhammad Shahbaz Aslam and Muhammad Amin Athar. (2013). Phytochemical, toxicological and antimicrobial evaluation of *Lawsonia inermis* extracts against clinical isolates of pathogenic bacteria. Ann. Clin. Microbiol. Antimicrobials. 12:36. DOI: 10.1186/1476-0711-12-36
- Hussey, R.S. and K. Baker (1973). A comparison of methods of collecting inocula for *Meloidogyne* spp. including a new technique. Plant Disease Reporter. 57:1025-1028.
- Hussey, R.S. and G.J.W. Janssen (2002). Root-knot nematodes: *Meloidogyne* species. pp 43-70. In: J.L. Starr, R. Cook, and J. Bridge, (eds.). Plant resistance to parasitic nematodes. CAB International, London, England.
- Jardim, I.N., D.F., Oliveira, V.P., Campos, G.H., Silva, and P.E. Souza (2020). Garlic essential oil reduces the population of *Meloidogyne incognita* in tomato plants. Eur. J. Plant Pathol. 157, 197-209 (2020). <https://doi.org/10.1007/s10658-020-02000-1>
- Jatto, N. A, M.A., Maikasuwa, A., Audu, and A. Alkali (2012). Assessment of farmers' understanding of the information displayed on pesticide product labels in Ilorin metropolis of Kwara state. Agrosearch. 12(1):107-116. DOI:10.4314/agrosh.v12i1.10
- Javaid A, I.H., Khan, A. Shoaib (2018). Management of charcoal rot of mungbean by two *Trichoderma* species and dry biomass of *Coronopus didymus*. Planta Daninha 36: article e018182795. DOI: 10.1590/s0100-83582018360100124
- Khan, I.H and A. Javaid (2020). Anticancer, antimicrobial and antioxidant compounds of quinoa inflorescence. Adv. Life Sci. 8(1): 68-72. Khan, I.H, Javaid, A., Al-Taie, A.H. and D. Ahmed (2020). Use of neem leaves as soil amendment for the control of collar rot disease of chickpea. Egypt. J. Biol. Pest Control 30: Article 98. DOI: 10.1186/s41938-020-00299-w
- Kidanemariam.T. K, Tesema, K. T, Asressu, K.H. and A.D. Boru (2013). Chemical investigation of *Lawsonia inermis* L. Leaves from a far region, Ethiopia. Oriental J. Chem. 29(3). 1129-1134.
- Koleva, L. and I. Mitova (2021). Effect of Mixed Crops on Soil Plant Parasitic Nematodes in Lettuce Cultivation (*Lactuca sativa* L.). Indian J. Agric. Res. 55 756-760. DOI: 10.18805/IJARE.A-622
- Najar, B. and L. Pistelli (2017). Essential oil composition of *Lawsonia inermis* leaves from Tunisia. American J. Essential oil Nat. Products. 5(3):7-11
- Okpekon T, Yolou S, Gleye C, Roblot F, Loiseau P, Bories C, Grellier P, Frappier F, Laurens A, and R. Hocquemiller (2004). Antiparasitic activities of medicinal plants used in Ivory Coast. J. Ethnopharm. 90(1):91-7. <https://doi.org/10.1016/j.jep.2003.09.029>
- Oliveira, G.H.F., Santana, S.R.A., Fonseca, R.C.N., de Lima, L.E., Gomes, L.A.A. and de J.L.S. Carvalho Filh (2015). *Meloidogyne incognita* resistant strains of leaf lettuce. African J. Agric. Res. 10(51):4660-4667.
- Ozdemir, E. and U. Gozel (2018). Nematicidal activities of essential oils against *Meloidogyne incognita* on tomato plant. Fresenius Environmental Bulletin. 27(6):4511-4517.
- Pardavella, I., Daferera, D., Tselios, T., Skiada, P. and I. Giannako (2020). The use of essential oil and

- hydrosol extracted from *Cuminum cyminum* seeds for the control of *Meloidogyne incognita* and *Meloidogyne javanica*. *Plants* (Basel, Switzerland). 10(1):E46. DOI: 10.3390/plants10010046.
- Pardavella, I., Nasiou, E., Daferera, D., Trigas, P. and I. Giannakou (2020). The use of essential oil and hydrosol extracted from *Satureja hellenica* for the control of *Meloidogyne incognita* and *M. javanica*. *Plants*. 9: 856. doi:10.3390/plants9070856.
- Pinheiro, J.B, Pereira, R.B, Carvalho, A.D.F, Rodrigues, C.S. and F.A. Suinaga (2013). Manejo de nematoides na cultura da alface. Brasília: Embrapa Hortaliças, Circular Técnica. 8p.
- Rodrigues, C.S, Pinheiro, J.B, Suinaga, F.A, Pereira, R.B, and A.D.F. Carvalho (2012). Seleção preliminar de cultivares de alface para resistência ao nematoide-das-galhas. *Horticultura Brasileira*. 30: S2048-S2054.
- Sharma, A. and K. Sharma (2011). Assay of antifungal activity of *Lawsonia inermis* (L) and *Eucalyptus citriodora* Hook. *J. Pharm. Res.* 4(5):1313-1314.
- Sharma, V.K. (1990). Tuberculostatic activity of henna (*Lawsonia inermis* Linn.). *Tubercle*. doi.org/10.1016/0041-3879 (90)90044-9. 71(4):293-295.
- Sherifa Mostafa M. Sabra, Luluah Mohammed R. Al-Masoudi, Hala Abd ElMageed E. Hasan Samar Ahamed H. Al-Gehani and A. A. O. Abu-Harbah. (2015). The Importance of the chemical composition of henna tree leaves (*Lawsonia inermis*) and its ability to eliminate *Tinea pedis*, with reference to the extent of usage and storage in the Saudi Society. *J. Pharm. Biol. Sci.* 10(4):23-29
- Siddiqui, Z. A, Iqbal, A. and I. Mahmood (2001). Effects of *Pseudomonas fluorescens* and fertilizers on the reproduction of *Meloidogyne incognita* and growth of tomato. *Appl. Soil Ecol.* 16:179-85. [https://doi.org/10.1016/S0929-1393\(00\)00083-4](https://doi.org/10.1016/S0929-1393(00)00083-4)
- Smiley, R. (2005). Plant-parasitic nematodes affecting wheat yield in the Pacific Northwest. Oregon State University extension publication. EM 8887. 4 pp. available at <http://extension.oregonstate.edu/catalog/pdf/em/em8887.pdf>.
- Taniwiryonoc Wiratno D, Van Den Berg H, Riksen, J.A.G, Rietjens, I.M.C.M, Djiwanti, S.R. and J.E. Kammenga (2009). Nematicidal activity of plant extracts against the root-knot nematode, *Meloidogyne incognita*. *Open Natural Product Journal*. 2:77-85. <https://doi.org/10.2174/1874848100902010077>
- Viaene, N. M, and G.S. Abawi (1998). Management of *Meloidogyne hapla* on lettuce in organic soil with Sudan grass as a cover crop. *Plant Disease*. 82: 945-952. doi: 10.1094/PDIS.1998.82.8.945.
- Widmer, T. L. and G. S. Abawi. (2000). Mechanism of suppression of *Meloidogyne hapla* and its damage by a green manure of Sudan grass. *Plant Disease*. 84: 562-568. doi: 10.1094/PDIS.2000.84.5.562
- Wilcken, S.R.S, Garcia, M.J.M, and N. Silva (2005). Resistência de alface do tipo Americana a *Meloidogyne incognita* raça 2. *Nematologia Brasileira*. 29: 267-271.
- Williams, M. (2012). Organic lettuce and leafy greens. University of Kentucky Cooperative Extension service. Available at: [http://www.uky.edu/Ag/New Crops/intro sheets/organic_lettuce.pdf](http://www.uky.edu/Ag/New_Crops/intro_sheets/organic_lettuce.pdf). [Accessed 10 February 17 2022].