

## ANALYSIS OF LNCRNAs AND CIRC RNAs IN *GLYCINE MAX* UNDER DROUGHT AND SALINE-ALKALINE STRESSES

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### ABSTRACT

Long noncoding-RNAs (LncRNAs) and circular RNAs (circRNAs) are the major types of noncoding RNAs (ncRNAs) known to be involved during diverse biological processes in eukaryotes. However, less attention has been given to the identification, overall characteristics and expression analysis of LncRNAs and circRNAs under abiotic stresses in plants. In our study, strand-specific RNA-seq data revealed the identification of a total 2,020 LncRNAs, including 1,979 known and 41 novel ones, and 830 circRNAs in soybean. Overall, 118 and 47 LncRNAs showed significant changes in expression levels under drought stress and saline-alkaline stress, respectively. In addition, soybean LncRNAs indicated diverse expression patterns suggesting a positive correlation with many biological processes including photosynthesis during stress responses by regulating gene expression in a *cis/trans*-acting manner. Our findings further indicated that transcriptional biosynthesis pathways and RNA transcriptional regulation were positively regulated during stress conditions. Furthermore, the back-splicing sites of circRNAs were also confirmed. Collectively, our study identified a group of stress-responsive LncRNAs and circRNAs in soybean, highlighting their potential contributions to abiotic stress responses, which pave the way for future functional studies.

**Keywords:** Strand-specific RNA sequencing; Stress response; lncRNA; circRNA; Expression pattern;

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### INTRODUCTION

Among the abiotic stresses that negatively affect the growth and development of crop plants, environmental factors are the most common inhibitors of yield (Bouain *et al.* 2019). *Glycine max* (soybean) is an economically important crop that is sensitive to abiotic stresses, particularly drought and saline-alkaline stresses (Kunert *et al.* 2016). This sensitivity means that there has been only limited success in breeding stress-resistant soybean cultivars. To overcome these limitations and increase the productivity of soybean, the molecular mechanisms of stress tolerance and their interaction with environmental factors should be better understood (Deinlein *et al.* 2014).

Noncoding RNAs are now recognized as crucial components of the stress response signaling networks that regulate plant defense systems. Long noncoding RNAs (lncRNAs) are defined as a heterogeneous group of noncoding RNAs whose transcripts are longer than 200 nucleotides (nt) (Kapranov *et al.* 2007). They do not contain any evolutionarily conserved open reading frames (Rivas *et al.* 2017). Previous studies have shown that they are expressed in a tissue- or stage-specific manner and their expression levels change in response to various environmental factors (Di *et al.* 2014; Liu *et al.* 2015; Qi *et al.* 2013; Rymarquis *et al.* 2008). The lncRNAs are functionally significant because they regulate RNA

processing machinery in a *cis* or *trans* manner (Kung *et al.* 2013). Emerging evidence has indicated that some plant lncRNAs play multiple roles in the regulation of plant metabolism, development, stress responses, and other biological processes (Di *et al.* 2014; Shafiq *et al.* 2016; Wang *et al.* 2014). Numerous lncRNAs have been reported to participate in the responses to a wide variety of stresses in *Arabidopsis*, maize, medicago, and wheat (Liu *et al.* 2012; Wang *et al.* 2015b; Xin *et al.* 2011; Zhang *et al.* 2014). In addition, several stress-responsive lncRNAs such as COLDAIR, lncRNA16397, and Cdc28 have been functionally characterized in plants (Cui *et al.* 2016; Heo and Sung 2011; Nadal-Ribelles *et al.* 2014). However, we still have only a rudimentary understanding of the functions of most lncRNAs in response to stress.

Circular RNAs (circRNAs) are another category of noncoding RNAs. They are circular in structure and are widespread in viroids or eukaryotes (Capel *et al.* 1993; Cocquerelle *et al.* 1993). CircRNAs can arise from exons or introns but are distinct structures with an independent manner of regulation (Memczak *et al.* 2013). Similar to lncRNAs, circRNAs show low expression levels and cell-type-specific patterns in eukaryotic cells (Salzman *et al.* 2013; Zuo *et al.* 2016). CircRNAs were suggested to be functionless when they were first identified in junk DNA some two decades ago (Nigro *et al.* 1991). However, recent studies have shown that circRNAs potentially play a role in enhancing the

transcriptional activity of host genes (Hansen *et al.* 2013; Li *et al.* 2015; Memczak *et al.* 2013). Analyses of the expression patterns and regulatory functions of circRNAs have revealed that they are expressed in a range of eukaryotic organisms and have a regulatory pattern independent of, and different from, that of their cognate linear isoforms.

Stress-responsive lncRNAs and circRNAs have not been well studied in soybean. Little is known about how noncoding RNAs are expressed and responsible for gene expression regulation in soybean at the transcriptional or post-transcriptional levels during survival or acclimation responses. In this study, therefore, we conducted a whole-transcriptome analysis, identified comprehensive sets of soybean lncRNAs and circRNAs, assessed the expression profiles of lncRNAs, and analyzed the regulatory mechanisms of lncRNAs under drought and saline-alkaline stresses.

## MATERIALS AND METHODS

**Materials, stress treatments, and determination of physiological indexes:** Seeds of the cultivated soybean variety “JY72” (a landrace cultivated in northeastern China) were placed in a sterile culture container in 1× Hoagland’s solution (containing 4 mL/L Fe sequestrene, 6 mM K<sup>+</sup>, and 4 mM Ca<sup>2+</sup>). At the stage of four leaves, the plants were subjected to abiotic stress for 24 hours under continuous light (Liu *et al.* 2012). To impose the drought and saline-alkaline stresses, the seedlings were incubated in liquid Hoagland’s medium containing 8% PEG and 110 mM NaCl + 50 mM NaHCO<sub>3</sub>, respectively. Seedlings in liquid Hoagland’s medium served as the control. The leaves were harvested from three groups per treatment, frozen with liquid nitrogen, and then stored at -80°C. Leaves were collected at 0 h, 6 h, 12 h, and 24 h of the treatments to analyze superoxide dismutase (SOD) activity and malondialdehyde (MDA) content. A Tecan infinite M1000 Pro Microplate Reader (San Jose, CA, USA) was used to measure the absorbance of reaction solutions to determine SOD activity and calculate MDA content. The reaction solutions were prepared according to the instructions of a physiological assay kit (Nanjing Jiancheng Bioengineering Institute, Nanjing, China). All the processes were biologically and temporally repeated in three independent and parallel experiments.

**Library construction for RNA-seq and sequencing procedures:** mirVana™ miRNA Isolation Kit (Ambion, Foster City, CA, USA) was employed to isolate total RNAs. The TruSeq® Stranded RNA Sample Preparation reagent (Illumina, San Diego, CA, USA) was used to prepare Strand-specific libraries by following the manufacturer’s instructions. We used Ribo-Zero rRNA removal beads to get rid of Ribosomal RNA from total RNA. After purification, total RNA was break up in to

small pieces of fragments by using divalent cations at 95°C for 8 min. These RNA fragments were then synthesized into cDNA by using random primers. DNA Polymerase I was used to synthesis second-strand cDNA. Then, these cDNA fragments were precessed with end-repairment, including the addition of a single “A” base, and then ligated with adapters. Qubit® 2.0 Fluorometer (Life Technologies, Gaithersburg, MD, USA) was employed to quantify purified libraries and validated by Agilent 2100 bioanalyzer (Agilent Technologies, Palo Alto, CA, USA) to make sure the insert size and measure concentration. Clusters were generated by cBot with the library diluted to 10 pM and sequencing by Illumina HiSeq 2500 sequencer (Illumina, USA).

**Bioinformatics analysis:** After filtering rRNAs, adapters and low-quality reads, and short-fragments, FASTQC tools

(<http://www.bioinformatics.babraham.ac.uk/projects/fastqc/>) were adopted to process clean reads. We used TopHat v2.0.9 (Trapnell *et al.* 2009) to map the clean reads to the Glycine max v2.0 reference genome with two mismatches allowed. The transcriptomes of three libraries were assembled separately using Cufflinks and the whole genome sequence as a reference. by using Cuffcompare, when after comparing with the assembled transcript isoforms and the known protein coding transcripts, novel transcripts were gained. Putative lncRNAs were identified as novel transcripts, with the filters as followed: length more than 200 bp; exons more than 2; and ORF less than 300 bp (He *et al.* 2015; Wang *et al.* 2018; Wang *et al.* 2017); weak or non protein-coding ability (CPC score < 0 (Kong *et al.* 2007), CNCI score < 0 (Sun *et al.* 2013)), and no significant similarity with Pfam in a protein-coding database (Punta *et al.* 2012). We selected dynamically expressed lncRNAs used for predicting targets. The cis-acting target genes were considered as transcribed upstream or downstream of the lncRNAs within 10 kb distance. RNAplex software was employed to predicted trans-acting targets.

We used CIRI version 1.2 (Gao *et al.* 2015) to characterize circRNAs. We used CIRI V1.2 to detect junction sites in sequence reads with PCC signals and PEM signals that reflected circRNA candidates by SAM alignment. Preliminary filtering was implemented using GT-AG splicing signals for the junction reads. After junction clustering, CIRI V1.2 was used to scan the SAM alignment again to detect junction-related reads and conduct further filtering to eliminate false-positive candidates.

**qRT-PCR analyses:** cDNA first strand was synthesized by using Reverse Transcript reagent (Takara, Dalian, China) with total RNA as template. All qRT-PCR reactions were operated on a Stratagene Mx3000P equipment (Agilent, USA) with a reaction volume of 25 µl consisting of 12.5 µl 2× SYBR premix Ex taq™

(Takara, Otsu, Japan), 2  $\mu$ l cDNA and 10  $\mu$ M specific primers. The soybean tubulin gene was employed as an internal control.  $2^{-\Delta\Delta Ct}$  formula was used to calculate the relative gene expression.

**Data analysis:** Data was subjected to a one-way ANOVA or Student's t test to determine the statistical significance. SPSS was used for statistical analysis. Differences among groups were considered statistically significant at  $P < 0.05$ .

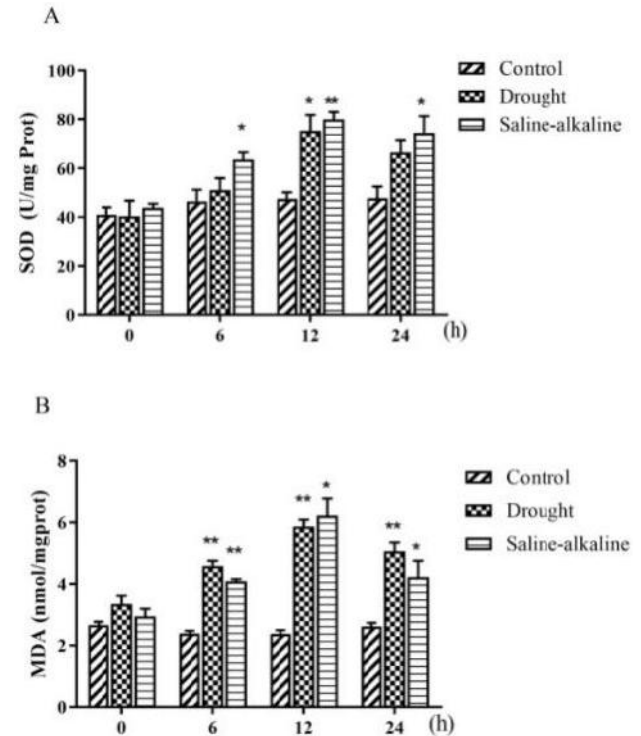
## RESULTS

### Identification of lncRNAs and circRNAs in soybean:

The SOD activity and MDA content were significantly increased by 12 h of the drought and saline-alkaline treatments (Figure 1), confirming that the physiological status of these plants was seriously affected under these conditions. Therefore, plants at 12 hours of these stress treatments were used as the source of RNAs for library construction and sequencing (Table S1). A total of 119,631 transcripts were acquired from the three libraries (Control, drought stress (8% PEG-treated for 12 h), and saline-alkaline stress (110 mM NaCl + 50 mM NaHCO<sub>3</sub>-treated for 12 h). The assembled transcripts showing alignment ( $P$  value  $< 1.0E^{-10}$ , identity more than 90%, coverage more than 80%) to known soybean lncRNA transcripts significantly were identified as known lncRNAs ( $n=1,979$ , 47.81%) (NCBI Glycine\_max\_v2.0). The remaining transcripts were processed by size selection to identify novel lncRNAs. A total of 155 transcripts with length more than two hundreds nucleotides and with an ORF length of fewer than one hundred amino acids was extracted. After filtering based on exon number, 103 transcripts with two or more exons were selected. Finally, 41 novel lncRNAs were identified (Table S2). Of these, four were intronic lncRNAs, 28 were intergenic lncRNAs, and the remaining nine were antisense lncRNAs. We have supplied an annotation file of novel lncRNAs (Table S2) listing distinct lncRNAs that could be investigated by other researchers. A total of 2,020 lncRNAs were selected for further analysis. We compared the transcript length between lncRNAs and coding genes (Figure 2A). The transcript lengths of most lncRNAs were shorter than mRNAs, and fewer lncRNAs than mRNAs contained two or three exons (Figure 2B). Our analyses uncovered a set of lncRNAs expressed during the stress response in soybean. These results highlighted genes that may play functional roles in the stress response of soybean.

We also identified circRNAs from our dataset of mapping reads. We identified 830 non-redundant circRNAs, comprising 327, 452, and 353 candidate circRNAs that were recognized from junction reads in the datasets of Control, drought-, and saline-alkaline treated samples, respectively (Table S3). Most of the circRNAs were located in exonic and intergenic regions (Figure

2C). The mapping of junction reads showed that they were uniformly distributed among all the chromosomes (Figure 2D). We detected circRNAs with different numbers of exons and from various intergenic regions. This may be attributed to their transcription sites in the genome or different regulatory functions.



**Figure 1.** Superoxide dismutase activity and malondialdehyde content in soybean under drought and saline-alkaline stresses. \*  $p < 0.05$ , \*\*  $p < 0.01$

**Expression profiles of lncRNA:** We determined the numbers of lncRNAs in the three libraries. In total, 1,765 (87.38%) of the lncRNAs were co-expressed among the control, drought, and saline-alkaline treatments. Interestingly, few lncRNAs were commonly expressed between control and the drought treatment, between control and the saline-alkaline treatment, or between the drought and saline-alkaline treatments. Only a few lncRNAs were explicitly expressed in the drought and saline-alkaline treatments (Figure 3A), implying that these lncRNAs were conserved regulators of plant development or the stress response.

To identify drought and saline-alkaline stress-responsive lncRNAs, the normalized expression FPKM (fragments per kilobase of transcript, per million fragments sequenced) number of lncRNAs were compared among the three libraries. Among the 2,020 lncRNAs, 143 were identified as differentially expressed. Of them, 118 exhibited changes in transcript levels under drought stress and 47 did so under saline-alkaline

stress(Figure 3B). In the drought treatment, 109 (71.19%) lncRNAs were significantly highly expressed and 9 (28.81%) lncRNAs were down-regulated, as compared with control. In the saline-alkaline treatment, 25 (53.19%) lncRNAs were up expressed and 22 (46.81%) lncRNAs

were down-regulated, as compared with control. In addition, 10 co-expressed lncRNAs were up-regulated under both stresses, and 3 co-expressed lncRNAs were down-regulated under both stresses.

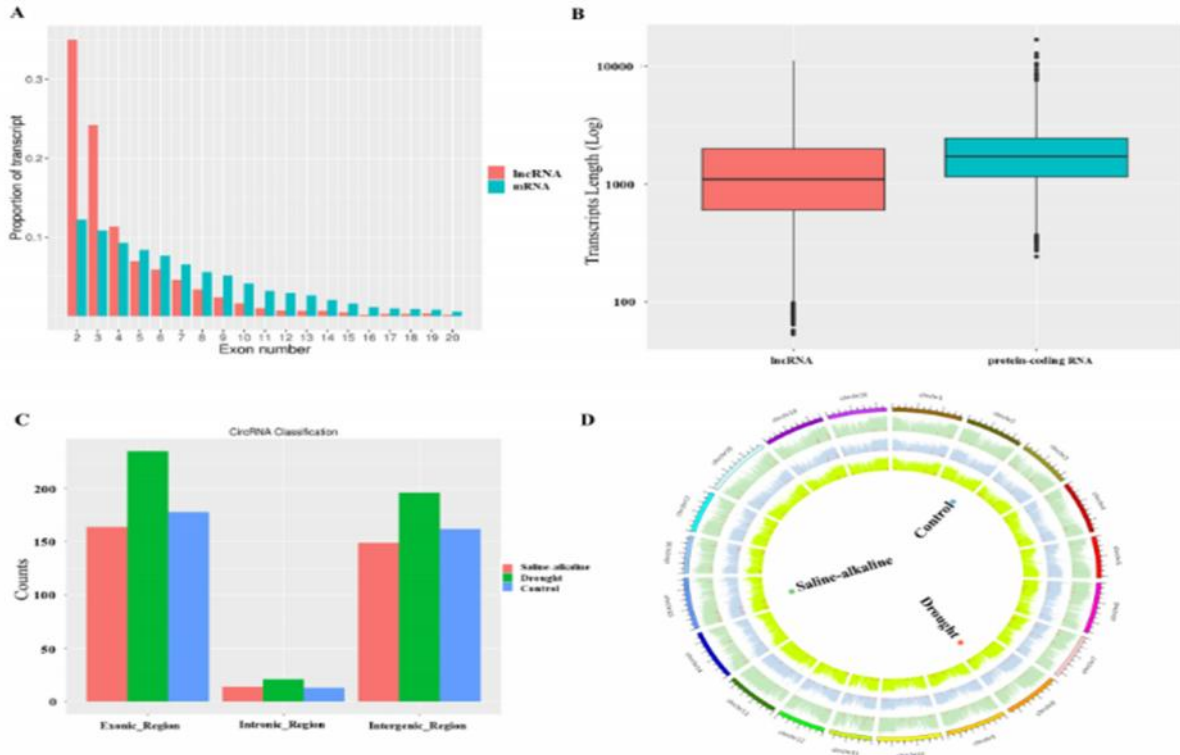


Figure 2. Identification of lncRNAs and circRNAs. a Number of exons in lncRNAs and coding RNAs. b Transcript levels of lncRNAs and coding RNAs. c Relative amount of each type of circRNAs. d Junction reads representing circRNAs (red) and mapping reads across treatment libraries mapped to genome.

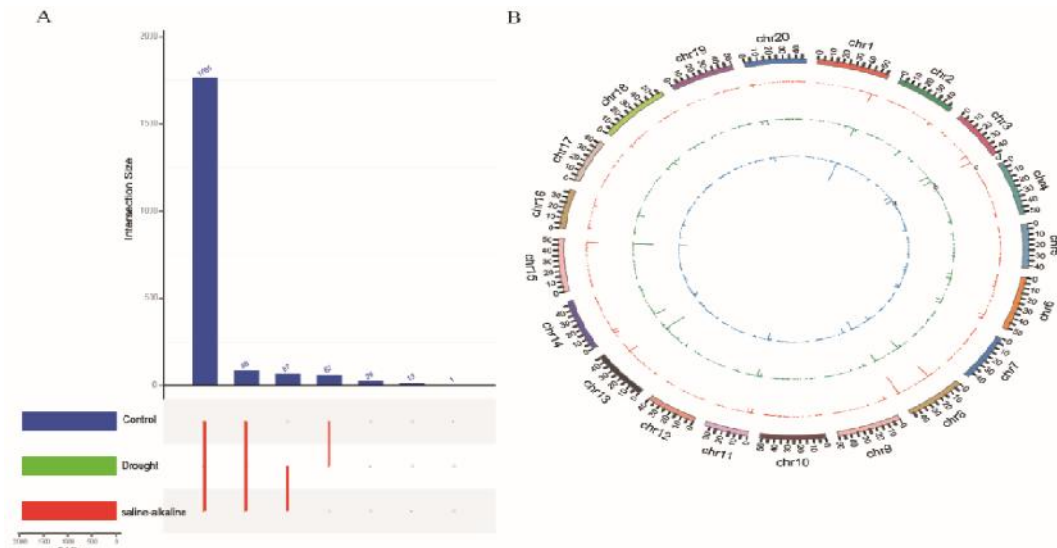


Figure 3. lncRNA expression profiles. a Categories of lncRNAs expressed under control, drought, and saline-alkaline conditions. b Relative abundance of lncRNA transcript levels under control (a, blue), drought (b, green), and saline-alkaline conditions (c, red).

**Analyses of functional roles of potential targets of lncRNAs:** The lncRNAs can take part in transcription process both in cis and in trans. Therefore, to predict the biological functions of genes affected by the differentially expressed lncRNAs under drought and saline-alkaline stresses, we identified the mRNAs potentially regulated by the 143 differentially expressed lncRNAs in cis or in trans. In this way, we predicted 5,575 mRNA target genes of lncRNAs.

First, we analyzed the genes whose transcription could be regulated by lncRNAs in cis. We conducted Gene Ontology (GO) analyses of the target genes to assign functional categories. Comparing the drought treatment with control, the targets of the dynamically expressed lncRNAs were significantly enriched in the categories of nucleoplasm (p-value = 5.60e-04) and transcription from RNA polymerase II promoter (p-value = 1.31e-03). Comparing the saline-alkaline treatment with control, the targets of the 47 dynamically expressed lncRNAs were significantly enriched in the categories of transcription factor activity (p-value = 9.55e-03), and signal transduction (p-value = 1.36e-02) (Table S4).

We conducted the same analyses for the genes whose transcription could be regulated by lncRNAs in trans. The trans targets of the lncRNAs differentially expressed under drought were significantly enriched in

biological processes, including ATPase activity (p-value = 1.47e-03), r pyrophosphatase activity (p-value = 1.29e-03), and acting on acid anhydrides (p-value = 1.35e-03). In the saline-alkaline treatment, the trans targets of the differentially expressed lncRNAs were significantly enriched in molecular functions, including hydrolyzing O-glycosyl compounds (p-value = 1.85e-04), acting on glycosyl bonds (p-value = 3.00e-04), and carbohydrate metabolic process (p-value = 2.47e-03). These results indicated that specific genes and pathways were regulated by differentially expressed lncRNAs during the plant stress response.

Together, these results showed that the lncRNAs whose expression levels changed upon exposure to drought and saline-alkaline stresses were functionally significant in pathways via their regulation of RNA processing in a cis- or trans-acting manner.

**Expression validation of lncRNAs and targets:** We selected 8 pairs of lncRNAs and the potential target mRNAs to validate their expression pattern under stresses by qRT-PCR. Excepted two pairs of lncRNA and potential targets (XR\_001386720.1/LOC100792149 and XR\_419416.2/LOC100815558), the others showed consistency with sequencing results (Figure 4).

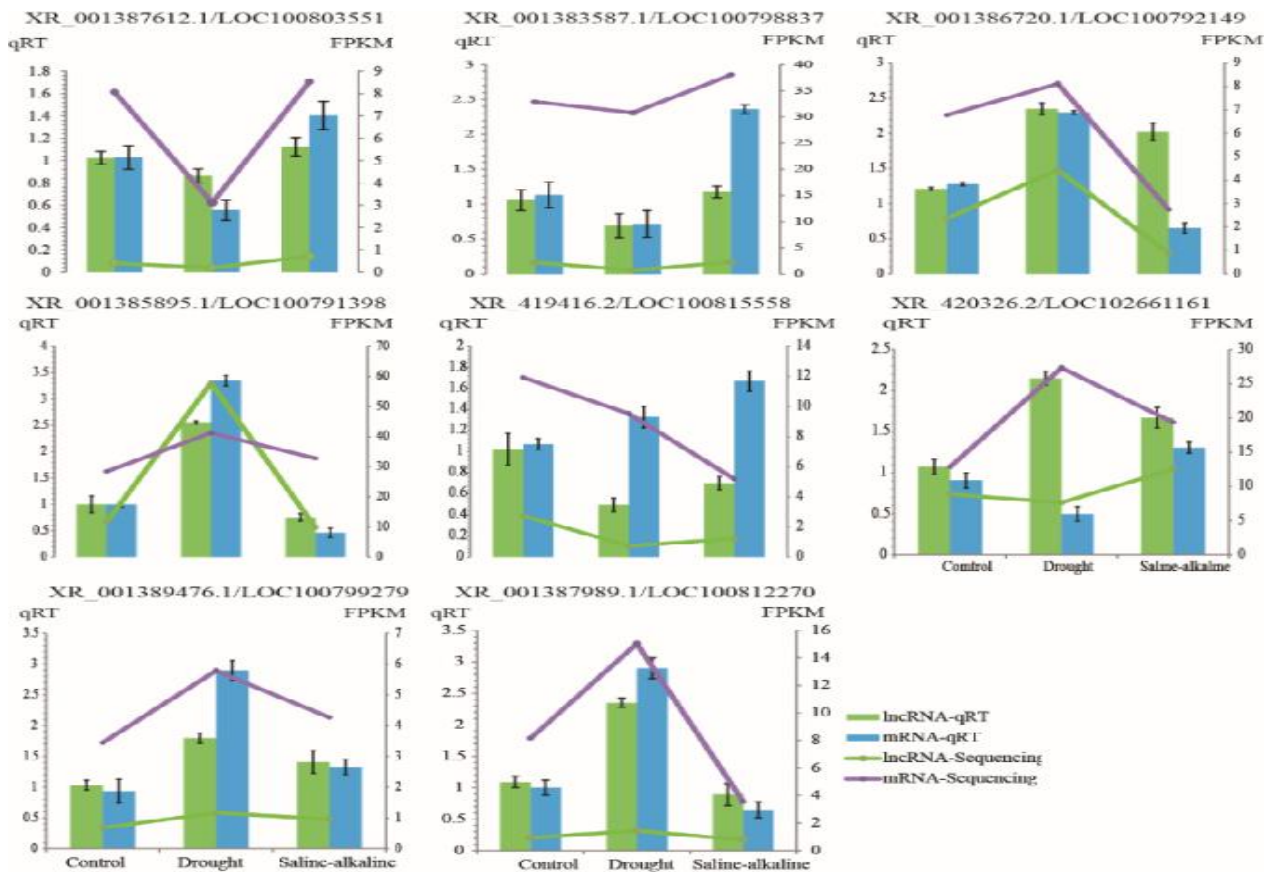
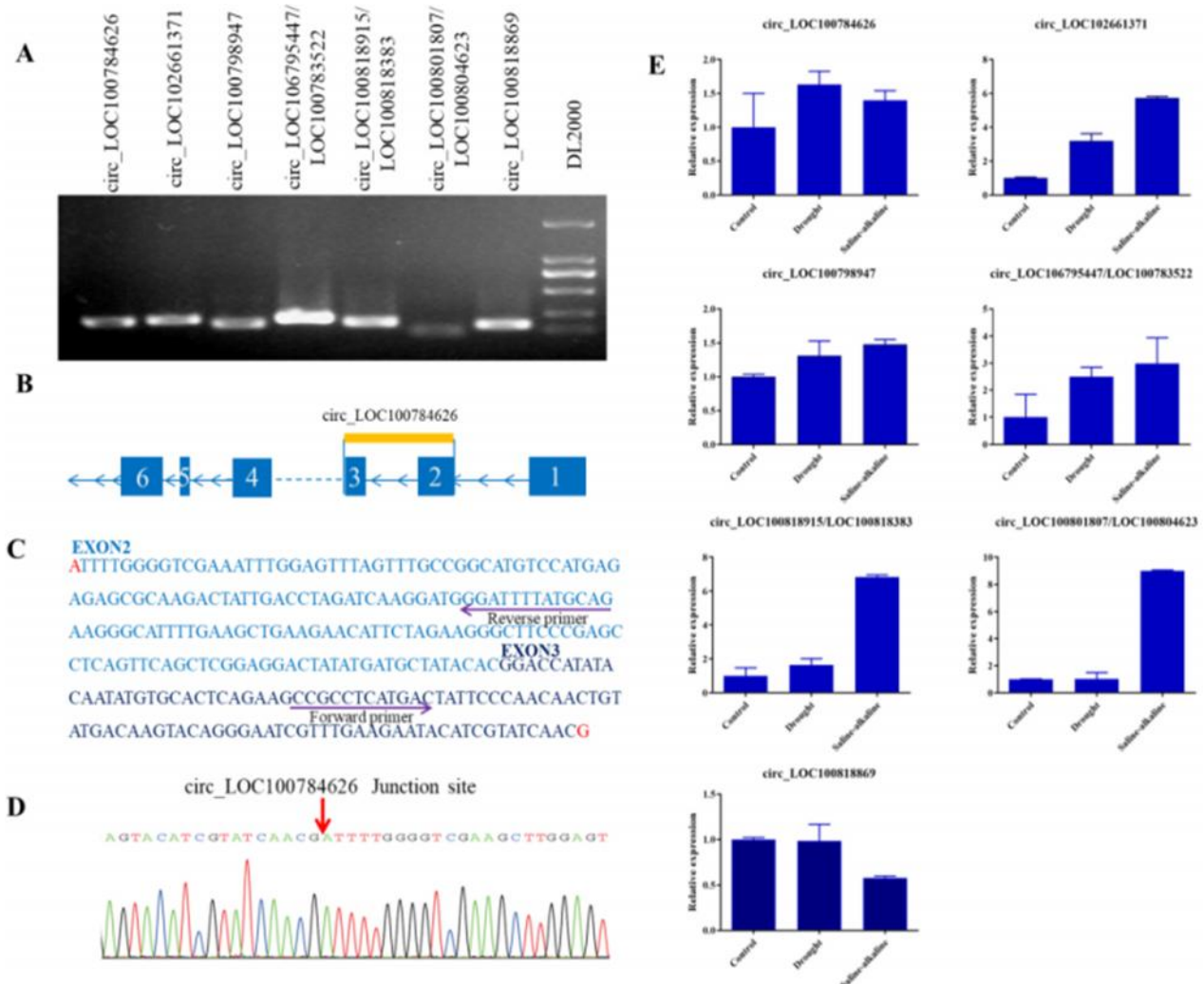


Figure 4. Expression validation of candidate lncRNAs and potential targets.

**Characterization and validation of circRNAs:** Soybean circRNAs generated from exonic, intergenic, and intronic regions were identified according to the transcriptomic data. Among the identified circRNAs, 123 were generated from exons of a single protein-coding gene and were classified as exonic circRNAs, and 223 were generated from intergenic regions. There was a relatively low rate of circRNAs from the intron-flanking regions. Splicing junction sites are one of the most important factors delineating the origin of the circRNAs and in

differentiating linear splicing from back-splicing. We found that seven of 12 circRNAs (four exonic circRNAs, two intronic circRNA, and four intergenic circRNAs) had junction sites (Figure 6a and Figure S1). Primers were specifically designed for circRNA segment amplification (Figure 6b, c, d). The expression patterns of some circRNAs were verified by qRT-PCR (Figure 6e). Only circ\_LOC100818869 was expressed at low levels under stress conditions. All the other circRNAs were expressed at high levels under stress conditions.



**Figure 5.** Characterization and validation of circRNAs. **a** PCR product amplified with lncRNA-specific primers from soybean cDNA. **b** Distribution of circ\_LOC100784626 in cullin-1 exon. **c** Primers designed to amplify circ\_LOC100784626. **d** Sequencing results showing junction sites in circ\_LOC100784626. **e** Expression of circRNA validated by qRT-PCR. \*  $p < 0.05$ , \*\*  $p < 0.01$ .

## DISCUSSION

Noncoding RNAs such as lncRNAs have been detected out or identified by many different techniques, such as strand-specific microarrays, conventional expression arrays, next generation RNA sequencing, or

real-time experiments (Herzog *et al.* 2014; Lu *et al.* 2012). Strand-specific next generation RNA sequencing permitted the recovery of antisense transcription activities and led to novel lncRNAs recognition. At present, plant biologists utilized this method to analyze transcriptomes and characterize a wide range of

noncoding RNAs in many plant varieties (Cruz de Carvalho *et al.* 2016; Joshi *et al.* 2016; Wang *et al.* 2015a). Building on this work, we performed genome-wide identification of noncoding RNAs by analyzing strand-specific paired-end RNA sequencing data. A total of 2,020 lncRNAs and 830 circRNAs were identified from the strand-specific RNA sequencing data. We found far fewer lncRNAs than were found in another recent study (Li *et al.* 2014), possibly because we used more rigorous filtering criteria to identify lncRNAs.

The expression profiles of circRNAs were not considered in this study. After discarding low-mapping-quality reads and back-spliced junction sequences with homology to linear exon junctions, most of the identified circRNAs showed very low expression levels and high false-positive rates. Although new methods such as CIRI-full have been proposed to evaluate transcript levels of circRNAs (Zheng *et al.* 2019), a comprehensive analysis of circRNA expression is still lacking.

We focused on the expression patterns and features of lncRNAs in this study. To understand the tolerance mechanisms of soybean, we first profiled the drought and saline-alkaline induced changes in the expression profiles of lncRNAs. We found 118 and 47 differentially expressed lncRNAs in response to dehydration or saline-alkaline conditions at the transcriptional level. According to recent studies, lncRNAs primarily regulate transcription in a *cis*- or *trans*- manner to positively or negatively control the expression of coding genes (Bumgarner *et al.* 2009; Guil and Esteller 2012). In our study, we found that lncRNAs targeted 5,575 mRNAs in *acis*- or *trans*- manner, indicating that there are widespread regulatory interactions between lncRNAs and coding RNAs in soybean. In the drought and saline-alkaline treatments, the regulation of cell signaling was affected by 6 and 8 differentially expressed lncRNAs, respectively. The putative targets of the dynamically expressed lncRNAs with *cis*-acting regulation included genes encoding auxin response factor 3 (ARF3), IAA, and ras-related protein (RABA1). These findings suggested that lncRNAs are participated in regulating the expression of RNAs involved in signal transduction at the transcriptional level. This conclusion is consistent with the recent finding that lncRNAs function as regulators of cell signaling under stress (Saeedi Borujeni *et al.* 2018). Previous analyses have revealed that certain lncRNAs may improve plants' resistance to stress (Liu and Zhu 2014). Here, we identified differentially expressed lncRNAs under drought and saline-alkaline stresses. As an intergenic lncRNA, XR\_001389476.1 regulates the expression of a gene encoding an F-box protein in *acis*-acting manner in soybean. Drought caused a rapid change in the XR\_001389476.1 expression level. Previous studies have shown that the F-box gene expression level increases under oxidative stress and

affects the levels of signaling molecules such as reactive oxygen species (Kong *et al.* 2016). Further research is needed to elucidate the molecular mechanism of lncRNA in soybean.

CircRNAs are produced by head-to-tail back-splicing events and are participated in the RNA expression regulation at the translational level or the post-translational level (Darbani *et al.* 2016). CircRNAs were recently reported to be novel regulators of gene expression in plants (Zuo *et al.* 2016). Using strand-specific RNA-seq methods, we identified 830 circRNAs originating from diverse genomic regions under our stress conditions. Despite the low abundance of circRNAs, we validated the back-splicing sites to confirm the existence of circRNAs generated from exonic, intronic, and intergenic regions (Siegel *et al.* 2014). Taken together, our research indicated that circRNAs may have additional functions in plants and/or undiscovered functions in stress responses.

**Conclusion:** In summary, with the aid of strand-specific transcriptomic analysis, we identified 2,020 lncRNAs, including 1,979 known and 41 novel ones, in soybean. In addition, 830 circRNAs were identified. The results of GO functional enrichment analyses indicated that the dynamically expressed lncRNAs were participated in the regulation of certain biological processes. Putative *trans/cis* targets of the dynamically expressed lncRNAs were predicted. Most circRNAs originated from exonic, intronic, and intergenic sequences of *G. max* and back-spliced junction sites of predicted circRNAs were validated through Sanger sequencing. These results increase our knowledge of the lncRNAs and circRNAs that are involved in soybean stress responses.

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## REFERENCES

- Bouain, N., G. Krouk, B. Lacombe and H. Rouached. (2019). Getting to the Root of Plant Mineral Nutrition: Combinatorial Nutrient Stresses Reveal Emergent Properties. *Trends in plant science* 24(6). 542-552.
- Bumgarner, S.L., R.D. Dowell, P. Grisafi, D.K. Gifford and G.R. Fink. (2009). Toggle involving *cis*-interfering noncoding RNAs controls variegated gene expression in yeast. *Proceedings of the National Academy of Sciences* 106(43). 18321-18326.

- Capel, B., A. Swain, S. Nicolis, A. Hacker, M. Walter, P. Koopman, P. Goodfellow and R. Lovell-Badge. (1993). Circular transcripts of the testis-determining gene *Sry* in adult mouse testis. *Cell* 73(5). 1019-1030.
- Cocquerelle C., B. Mascrez, D. Hetuin and B. Bailleul. (1993). Mis-splicing yields circular RNA molecules. *FASEB J* 7(1).155-160.
- Cruz de Carvalho, M.H., H.X. Sun, C. Bowler and N.H. Chua. (2016). Noncoding and coding transcriptome responses of a marine diatom to phosphate fluctuations. *New Phytol* 210(2). 497-510.
- Cui, J., Y. Luan, N. Jiang, H. Bao and J. Meng . (2016). Comparative transcriptome analysis between resistant and susceptible tomato allows the identification of *lncRNA16397* conferring resistance to *Phytophthora infestans* by co-expressing glutaredoxin. *Plant J* 89(3). 577-589.
- Darbani, B., S. Noeparvar and S. Borg. (2016). Identification of Circular RNAs from the Parental Genes Involved in Multiple Aspects of Cellular Metabolism in Barley. *Frontiers in plant science* 7. 776.
- Deinlein, U., A.B. Stephan, T. Horie, W. Luo, G. Xu and J.I. Schroeder. (2014). Plant salt-tolerance mechanisms. *Trends in plant science* 19(6).371-9.
- Di, C., J. Yuan, Y. Wu, J. Li, H. Lin, L. Hu, T. Zhang, Y. Qi, M.B. Gerstein, Y. Guo and Z.J. Lu. (2014). Characterization of stress-responsive *lncRNAs* in *Arabidopsis thaliana* by integrating expression, epigenetic and structural features. *Plant J* 80(5).848-61.
- Gao, Y., J. Wang and F. Zhao. (2015). CIRI: an efficient and unbiased algorithm for de novo circular RNA identification. *Genome Biol* 16(1).4.
- Guil, S. and M. Esteller. (2012). Cis-acting noncoding RNAs: friends and foes. *Nat Struct Mol Biol* 19(11). 1068-1075.
- Hansen, T.B., T.I. Jensen, B.H. Clausen, J.B. Bramsen, B. Finsen, C.K. Damgaard and J. Kjems. (2013). Natural RNA circles function as efficient microRNA sponges. *Nature* 495(7441).384-388.
- He, F., Q. Liu, L. Zheng, Y. Cui, Z. Shen and L. Zheng. (2015). RNA-Seq Analysis of Rice Roots Reveals the Involvement of Post-Transcriptional Regulation in Response to Cadmium Stress. *Frontiers in plant science* 6. 1136.
- Heo, J.B. and S. Sung. (2011). Vernalization-mediated epigenetic silencing by a long intronic noncoding RNA. *Science* 331(6013).76-9.
- Herzog, V.A., A. Lempradl, J. Trupke, H. Okulski, C. Altmutter, F. Ruge, B. Boidol, S. Kubicek, G. Schmauss, K. Aumayr, M. Ruf, A. Pospisilik, A. Dimond, H.B. Senergin, M.L. Vargas, J.A. Simon and L. Ringrose. (2014). A strand-specific switch in noncoding transcription switches the function of a Polycomb/Trithorax response element. *Nat Genet* 46(9).973-981.
- Joshi, R.K., S. Megha, U. Basu, M.H. Rahman and N.N. Kav. (2016). Genome Wide Identification and Functional Prediction of Long Non-Coding RNAs Responsive to *Sclerotinia sclerotiorum* Infection in *Brassica napus*. *PLoS One* 11(7). e0158784.
- Kapranov, P., J. Cheng, S. Dike, D.A. Nix, R. Duttagupta, A.T. Willingham, P.F. Stadler, J. Hertel, J. Hackermuller, I.L. Hofacker, I. Bell, E. Cheung, J. Drenkow, E. Dumais, S. Patel, G. Helt, M. Ganesh, S. Ghosh, A. Piccolboni, V. Sementchenko, H. Tammana and T.R. Gingeras. (2007). RNA maps reveal new RNA classes and a possible function for pervasive transcription. *Science* 316(5830).1484-1488.
- Kong, L., Y. Zhang, Z.Q. Ye, X.Q. Liu, S.Q. Zhao, L. Wei and G. Gao. (2007). CPC: assess the protein-coding potential of transcripts using sequence features and support vector machine. *Nucleic Acids Res* 35. W345-349.
- Kong, X., S. Zhou, S. Yin, Z. Zhao, Y. Han and W. Wang. (2016). Stress-Inducible Expression of an F-box Gene *TaFBA1* from Wheat Enhanced the Drought Tolerance in Transgenic Tobacco Plants without Impacting Growth and Development. *Frontiers in plant science* 7. 1295.
- Kunert, K.J., B.J. Vorster, B.A. Fenta, T. Kibido, G. Dionisio and C.H. Foyer. (2016). Drought Stress Responses in Soybean Roots and Nodules. *Frontiers in plant science* 7. 1015.
- Kung, J.T., Colognori D and Lee JT. (2013). Long noncoding RNAs: past, present, and future. *Genetics* 193. 651-669.
- Li, J., B. Wu, J. Xu and C. Liu. (2014). Genome-wide identification and characterization of long intergenic non-coding RNAs in *Ganoderma lucidum*. *PLoS One* 9. e99442.
- Li, Z., C. Huang, C. Bao, L. Chen, M. Lin, M. Wang M, G. Zhong, B. Yu, W. Hu, L. Dai, P. Zhu, Z. Chang, Q. Wu, Y. Zhao, Y. Jia, P. Xu, H. Liu and G. Shan. (2015). Exon-intron circular RNAs regulate transcription in the nucleus. *Nat Struct Mol Biol* 22(3). 256-264.
- Liu, J., C. Jung, Xu J, H. Wang, S. Deng, L. Bernad, C. Arenas-Huertero and N.H. Chua. (2012). Genome-wide analysis uncovers regulation of long intergenic noncoding RNAs in *Arabidopsis*. *Plant Cell* 24(11). 4333-4345.
- Liu, R. and J-K. Zhu. (2014). Non-coding RNAs as potent tools for crop improvement. *National Science Review* 1. 186-189.



- Liu, X., Hao L, Li D, Zhu L and Hu S. (2015). Long non-coding RNAs and their biological roles in plants. *Genomics Proteomics Bioinformatics* 13(3). 137-147.
- Lu, T., C. Zhu, G. Lu, Y. Guo, Y. Zhou, Z. Zhang, Y. Zhao, W. Li, Y. Lu, W. Tang, Q. Feng and B. Han. (2012). Strand-specific RNA-seq reveals widespread occurrence of novel cis-natural antisense transcripts in rice. *BMC genomics* 13. 721.
- Memczak, S., M. Jens, A. Elefsinioti, F. Torti, J. Krueger, A. Rybak, L. Maier, S.D. Mackowiak, L.H. Gregersen, M. Munschauer, A. Loewer, U. Ziebold, M. Landthaler, C. Kocks, F. le Noble and N. Rajewsky. (2013). Circular RNAs are a large class of animal RNAs with regulatory potency. *Nature* 495(7441). 333-338.
- Nadal-Ribelles, M., C. Sole, Z. Xu, L.M. Steinmetz, E. de Nadal and F. Posas. (2014). Control of Cdc28 CDK1 by a stress-induced lncRNA. *Mol Cell* 53(4). 549-561.
- Nigro J.M., K.R. Cho, E.R. Fearon, S.E. Kern, J.M. Ruppert, J.D. Oliner, K.W. Kinzler and B. Vogelstein. (1991). Scrambled exons. *Cell* 64(3). 607-613.
- Punta, M., P.C. Coghill, R.Y. Eberhardt, J. Mistry, J. Tate, C. Boursnell, N. Pang, K. Forslund, G. Ceric, J. Clements, A. Heger, L. Holm, E.L. Sonnhammer, S.R. Eddy, A. Bateman and R.D. Finn. (2012). The Pfam protein families database. *Nucleic Acids Res* 40. D290-301.
- Qi, X., S. Xie, Y. Liu, F. Yi and J. Yu. (2013). Genome-wide annotation of genes and noncoding RNAs of foxtail millet in response to simulated drought stress by deep sequencing. *Plant Mol Biol* 83(4-5). 459-473.
- Rivas E., J. Clements and S.R. Eddy. (2017). A statistical test for conserved RNA structure shows lack of evidence for structure in lncRNAs. *Nat Methods* 14(1). 45-48.
- Rymarquis, L.A., J.P. Kastenmayer, A.G. Huttenhofer and P.J. Green. (2008). Diamonds in the rough: mRNA-like non-coding RNAs. *Trends in plant science* 13(7). 329-334.
- Saeedi Borujeni, M.J., E. Esfandiary, A. Baradaran, A. Valiani, M. Ghanadian, P. Codoner-Franch, R. Basirat, E. Alonso-Iglesias, H. Mirzaei and A. Yazdani. (2018). Molecular aspects of pancreatic beta-cell dysfunction: Oxidative stress, microRNA, and long noncoding RNA. *J. Cellular Physiology*. 234(6).8411-8425.
- Salzman, J., R.E. Chen, M.N. Olsen, P.L. Wang and P.O. Brown. (2013). Cell-type specific features of circular RNA expression. *PLoS Genet* 9(9). e1003777.
- Shafiq, S., J.Li and Q. Sun. (2016). Functions of plants long non-coding RNAs. *Biochim Biophys Acta* 1859(1). 155-162.
- Siegel, T.N., C.C. Hon, Q. Zhang, J.J. Lopez-Rubio, C. Scheidig-Benatar, R.M. Martins, O. Sismeiro, J.Y. Coppee and A. Scherf. (2014). Strand-specific RNA-Seq reveals widespread and developmentally regulated transcription of natural antisense transcripts in *Plasmodium falciparum*. *BMC genomics* 15(1). 150.
- Sun, L., H. Luo, D. Bu, G. Zhao, K. Yu, C. Zhang, Y. Liu, R. Chen and Y. Zhao. (2013). Utilizing sequence intrinsic composition to classify protein-coding and long non-coding transcripts. *Nucleic Acids Res* 41(17). e166.
- Trapnell, C., L. Pachter and S.L. Salzberg. (2009). TopHat: discovering splice junctions with RNA-Seq. *Bioinformatics* 25(9). 1105-1111.
- Wang, J., J. Lin, J. Kan, H. Wang, X. Li, Q. Yang, H. Li and Y. Chang. (2018). Genome-Wide Identification and Functional Prediction of Novel Drought-Responsive lncRNAs in *Pyrus betulifolia*. *Genes* 9(6).311
- Wang, J., W. Yu, Y. Yang, X. Li, T. Chen, T. Liu, N. Ma, X. Yang, R. Liu and B. Zhang. (2015a). Genome-wide analysis of tomato long non-coding RNAs and identification as endogenous target mimic for microRNA in response to TYLCV infection. *Sci Rep* 5. 16946.
- Wang, S., W. Ge, Z. Luo, Y. Guo, B. Jiao, L. Qu, Z. Zhang and X. Wang. (2017). Integrated analysis of coding genes and non-coding RNAs during hair follicle cycle of cashmere goat (*Capra hircus*). *BMC genomics* 18(1). 767.
- Wang, T.Z., M. Liu, M.G. Zhao, R. Chen and W.H. Zhang. (2015b). Identification and characterization of long non-coding RNAs involved in osmotic and salt stress in *Medicago truncatula* using genome-wide high-throughput sequencing. *BMC Plant Biol* 15. 131.
- Wang, Y., X. Wang, W. Deng, X. Fan, T.T. Liu, G. He, R. Chen, W. Terzaghi, D. Zhu and X.W. Deng. (2014). Genomic features and regulatory roles of intermediate-sized non-coding RNAs in *Arabidopsis*. *Mol Plant* 7. 514-527.
- Xin, M., Y. Wang, Y. Yao, N. Song, Z. Hu, D. Qin, C. Xie, H. Peng, Z. Ni, and Sun Q. (2011). Identification and characterization of wheat long non-protein coding RNAs responsive to powdery mildew infection and heat stress by using microarray analysis and SBS sequencing. *BMC Plant Biol* 11. 61.
- Zhang, W., Z. Han, Q. Guo, Y. Liu, Y. Zheng, F. Wu and Jin W. (2014). Identification of maize long non-coding RNAs responsive to drought stress. *PLoS One* 9(6). e98958.

Zheng, Y., Ji P, S. Chen, L. Hou and Zhao F. (2019). Reconstruction of full-length circular RNAs enables isoform-level quantification. *Genome medicine* 11(1). 2.

Zuo, J., Q. Wang, B. Zhu, Y. Luo and Gao L. (2016). Deciphering the roles of circRNAs on chilling injury in tomato. *Biochem Biophys Res Commun* 479(2). 132-138.

**Table S1: Sequencing read counts statistics.**

	CK	Drought treated	Saline-alkaline treated
Raw reads	74,950,632	65,770,632	71,420,564
Clean reads	58,022,071	47,403,083	55,733,811
rRNA trimmed reads	57,961,918	47,321,763	55,680,454
rRNA ratio	0.1%	0.2%	0.1%
Mapped reads	53,696,524	42,793,029	51,610,967
Mapped Unique reads	52,066,800	41,315,747	50,140,391
Mapped ratio	92.6%	90.4%	92.7%

**Table S2: Identification of novel lncRNAs under drought and Saline-alkaline in soybean.**

	lncRNA ID	class_code	lncRNA class	Cpc score	enci_score	Pfam
1	TCONS_00012324	U	Intergenic LncRNA	-0.9221	-0.00783	no-significantly
2	TCONS_00023602	U	Intergenic LncRNA	-0.90843	-0.01005	no-significantly
3	TCONS_00025836	X	Antisense LncRNA	-0.93126	-0.01252	no-significantly
4	TCONS_00036595	U	Intergenic LncRNA	-1.26151	-0.09042	no-significantly
5	TCONS_00033253	U	Intergenic LncRNA	-1.22851	-0.00768	no-significantly
6	TCONS_00046989	X	Antisense LncRNA	-1.09754	-0.02199	no-significantly
7	TCONS_00046988	x	Antisense LncRNA	-1.06345	-0.00957	no-significantly
8	TCONS_00045400	x	Antisense LncRNA	-1.22812	-0.00239	no-significantly
9	TCONS_00052681	u	Intergenic LncRNA	-1.12832	-0.00969	no-significantly
10	TCONS_00057019	i	Intron LncRNA	-1.2064	-0.00518	no-significantly
11	TCONS_00059481	u	Intergenic LncRNA	-1.31747	-0.00761	no-significantly
12	TCONS_00057716	x	Antisense LncRNA	-0.98229	-0.01628	no-significantly
13	TCONS_00059767	u	Intergenic LncRNA	-1.29368	-0.00041	no-significantly
14	TCONS_00061791	x	Antisense LncRNA	-1.15556	-0.01043	no-significantly
15	TCONS_00004367	x	Antisense LncRNA	-1.24674	-0.00505	no-significantly
16	TCONS_00076768	u	Intergenic LncRNA	-0.23619	-0.00041	no-significantly
17	TCONS_00074610	u	Intergenic LncRNA	-0.90069	-0.07398	no-significantly
18	TCONS_00075173	u	Intergenic LncRNA	-1.0945	-0.0313	no-significantly
19	TCONS_00075171	u	Intergenic LncRNA	-1.21018	-0.00113	no-significantly
20	TCONS_00077280	u	Intergenic LncRNA	-1.1109	-0.00885	no-significantly
21	TCONS_00075496	u	Intergenic LncRNA	-1.28975	-0.02906	no-significantly
22	TCONS_00082373	i	Intron LncRNA	-0.80982	-0.01808	no-significantly
23	TCONS_00079757	i	Intron LncRNA	-0.67632	-0.01233	no-significantly
24	TCONS_00082403	u	Intergenic LncRNA	-1.01098	-0.00041	no-significantly
25	TCONS_00080978	i	Intron LncRNA	-1.30921	-0.00041	no-significantly
26	TCONS_00081627	u	Intergenic LncRNA	-1.2581	-0.00384	no-significantly
27	TCONS_00087583	u	Intergenic LncRNA	-1.12123	-0.00957	no-significantly
28	TCONS_00093860	u	Intergenic LncRNA	-1.02239	-0.0069	no-significantly
29	TCONS_00092205	x	Antisense LncRNA	-0.88372	-0.02035	no-significantly
30	TCONS_00092233	u	Intergenic LncRNA	-1.00057	-0.00656	no-significantly
31	TCONS_00093840	u	Intergenic LncRNA	-1.32728	-0.02142	no-significantly
32	TCONS_00101224	u	Intergenic LncRNA	-0.846	-0.01057	no-significantly
33	TCONS_00113917	u	Intergenic LncRNA	-0.25127	-0.00241	no-significantly
34	TCONS_00114666	x	Antisense LncRNA	-1.03661	-0.00041	no-significantly
35	TCONS_00114109	u	Intergenic LncRNA	-0.25629	-0.00187	no-significantly
36	TCONS_00114110	u	Intergenic LncRNA	-0.19471	-0.00452	no-significantly
37	TCONS_00114886	u	Intergenic LncRNA	-1.21534	-0.00696	no-significantly

38	TCONS_00113534	u	Intergenic LncRNA	-1.29174	-0.00221	no-significantly
39	TCONS_00117108	u	Intergenic LncRNA	-1.18054	-0.01071	no-significantly
40	TCONS_00120669	u	Intergenic LncRNA	-1.34841	-0.00848	no-significantly
41	TCONS_00119739	u	Intergenic LncRNA	-1.25763	-0.00946	no-significantly

**Table S3: Identification of circRNA under Drought and Saline-alkaline in soybean.**

circRNA id	locus	circRNA type	gene id	Gene symbol
circRNA1	chr18:1884830 1885872(+)	Exon	100788781	LOC100788781
circRNA2	NW_014573484.1:15615 24814(-)	Intron	100499660	LOC100499660
circRNA3	chr19:49657767 49661626(+)	Exon	100775991	LOC100775991
circRNA4	chr3:28532292 28532473(+)	exon	100809668	LOC100809668
circRNA5	chr20:46722693 46727982(-)	exon	100790062	LOC100790062
circRNA6	chr8:47160960 47164126(-)	exon	100799467	LOC100799467
circRNA7	chr2:47568773 47583823(-)	intergenic region	n/a	-
circRNA8	chr17:24450022 24466703(+)	exon	100775929	LOC100775929
circRNA9	chr15:15637548 15656589(+)	intergenic region	n/a	-
circRNA10	chr20:46116509 46120220(-)	exon	100817872	LOC100817872
circRNA11	chr1:56815648 56816260(-)	exon	100780905	LOC100780905
circRNA12	chr12:40193 43348(+)	exon	100799920	LOC100799920
circRNA13	chr9:5772046 5774924(+)	exon	100820121	LOC100820121
circRNA14	chr20:2508603 2551745(-)	intergenic region	n/a	-
circRNA15	chrPltd:335 5514(-)	intergenic region	n/a	-
circRNA16	chrPltd:5368 6725(-)	exon	3989271	rbcl
circRNA17	chr7:7658602 7672377(-)	intergenic region	n/a	-
circRNA18	chr2:11494849 11616531(-)	intergenic region	n/a	-
circRNA19	chr12:32880 43348(+)	exon	100799920	LOC100799920
circRNA20	chr11:8391413 8392550(-)	exon	100813590	LOC100813590
circRNA21	chr14:1314546 1335196(-)	intergenic region	n/a	-
circRNA22	chr17:36354902 36355447(-)	exon	547792	LOC547792
circRNA23	chr3:9655011 9783089(-)	intergenic region	n/a	-
circRNA24	chr2:15872048 15873057(+)	exon	100800938	LOC100800938
circRNA25	chr3:6122686 6276422(-)	intergenic region	n/a	-
circRNA26	chr10:1959804 1970140(-)	intergenic region	n/a	-
circRNA27	chr18:39078403 39079643(+)	exon	102660444	LOC102660444
circRNA28	chr2:46960851 46961461(+)	exon	100789468	LOC100789468
circRNA29	chr17:40831152 40871234(-)	intergenic region	n/a	-
circRNA30	chr18:49836569 49849800(+)	intergenic region	n/a	-
circRNA31	chr8:44683812 44705494(+)	intergenic region	n/a	-
circRNA32	chr18:57506054 57507716(-)	exon	100811073	LOC100811073
circRNA33	chr3:27913096 27915318(-)	exon	100813936	LOC100813936
circRNA34	chr5:40299361 40304875(+)	exon	100791130	LOC100791130
circRNA35	chr11:27859973 27860542(-)	exon	100798309	LOC100798309
circRNA36	chr15:7218238 7222390(+)	intergenic region	n/a	-
circRNA37	chr13:30568820 30569013(-)	exon	100777805	LOC100777805
circRNA38	chr8:40889591 40891415(-)	exon	100787473	LOC100787473
circRNA39	chr12:16512230 16736082(+)	intergenic region	n/a	-
circRNA40	chr13:36343547 36345126(+)	exon	100816142	LOC100816142
circRNA41	chr14:12305639 12321322(-)	exon	100500679	LOC100500679
circRNA42	chr10:30293099 30293740(+)	exon	100798991	LOC100798991
circRNA43	chr2:32745356 32746444(+)	exon	100794561	LOC100794561
circRNA44	chr12:5573114 5573957(-)	exon	100805061	LOC100805061
circRNA45	chr10:43763701 43765098(+)	exon	100803596	LOC100803596
circRNA46	chr18:9120644 9144957(+)	intergenic region	n/a	-
circRNA47	chr2:14648729 14667584(+)	intergenic region	n/a	-
circRNA48	chr19:778328 778923(+)	exon	100781868	LOC100781868

circRNA49	chr12:39177164 39206976(-)	intergenic region	n/a	-
circRNA50	chr13:14415558 14436083(+)	intergenic region	n/a	-
circRNA51	chr15:1655878 1689424(+)	intergenic region	n/a	-
circRNA52	NW_014573484.1:15243 33868(-)	intergenic region	n/a	-
circRNA53	chr4:2706439 2714687(+)	intergenic region	n/a	-
circRNA54	chr20:1005647 1021407(+)	intergenic region	n/a	-
circRNA55	chr12:1286106 1292115(+)	intergenic region	n/a	-
circRNA56	chrPltd:6292 6408(-)	exon	3989271	rbcL
circRNA57	chr8:42660284 42660951(-)	exon	100782497	LOC100782497
circRNA58	chr18:50313127 50313492(+)	exon	100796148	LOC100796148
circRNA59	chr5:35568183 35578395(-)	intergenic region	n/a	-
circRNA60	chr20:16899360 16922585(-)	exon	102665764	LOC102665764
circRNA61	chr9:48932795 48934044(-)	exon	100811378	LOC100811378
circRNA62	chr9:24716008 24737521(+)	intergenic region	n/a	-
circRNA63	chr16:33084437 33251415(-)	intergenic region	n/a	-
circRNA64	chr13:29047135 29060169(-)	exon	100814539	LOC100814539
circRNA65	chr4:13572434 13573325(+)	exon	100789878	LOC100789878
circRNA66	chr13:14430407 14448409(+)	intergenic region	n/a	-
circRNA67	chr13:36343727 36345126(+)	exon	100816142	LOC100816142
circRNA68	chrPltd:1211 6911(-)	intergenic region	n/a	-
circRNA69	chr8:6307674 6310633(+)	exon	100807979	LOC100807979
circRNA70	chr19:38082939 38083918(+)	exon	100782594	LOC100782594
circRNA71	chr5:32741577 32742040(+)	exon	100802419	LOC100802419
circRNA72	chr18:9300525 9301248(+)	exon	106797140	LOC106797140
circRNA73	chr3:5477689 5523589(+)	intergenic region	n/a	-
circRNA74	chr3:235414 236481(-)	exon	100779711	LOC100779711
circRNA75	chr12:39171698 39201930(-)	intergenic region	n/a	-
circRNA76	chr12:35935071 35948579(-)	intron	100807714	LOC100807714
circRNA77	chrPltd:877 979(-)	exon	3989259	psbA
circRNA78	chr19:37245872 37251761(-)	exon	100785430	LOC100785430
circRNA79	chr13:30370619 30439972(-)	intergenic region	n/a	-
circRNA80	chr12:791508 805053(+)	intergenic region	n/a	-
circRNA81	chr1:3493520 3504862(+)	intergenic region	n/a	-
circRNA82	chr14:1457027 1471866(+)	intergenic region	n/a	-
circRNA83	chr7:5652654 5726099(-)	intron	100780862	LOC100780862
circRNA84	chr20:47534333 47534877(+)	exon	100811473	LOC100811473
circRNA85	chr7:15811552 15811976(-)	exon	100795391	LOC100795391
circRNA86	chr8:45413699 45428606(-)	intergenic region	n/a	-
circRNA87	chr10:30309271 30310511(+)	exon	100798991	LOC100798991
circRNA88	chr16:387312 388102(-)	exon	100804966	LOC100804966
circRNA89	chr7:6617619 6640644(+)	intergenic region	n/a	-
circRNA90	chrPltd:5383 6900(-)	intergenic region	n/a	-
circRNA91	chr8:44900765 44917039(-)	intergenic region	n/a	-
circRNA92	chr10:19655305 19665468(+)	intergenic region	n/a	-
circRNA93	NW_014573526.1:2705 17929(-)	intergenic region	n/a	-
circRNA94	chr12:16511175 16733760(+)	intergenic region	n/a	-
circRNA95	chr19:7190007 7253858(+)	intergenic region	n/a	-
circRNA96	chrPltd:132706 132974(-)	intergenic region	n/a	-
circRNA97	chr10:13923926 13925267(-)	exon	100814107	LOC100814107
circRNA98	chr2:14380600 14381817(+)	exon	547616	MAPK2
circRNA99	chr12:26068628 26068918(-)	exon	100796730	LOC100796730
circRNA100	chr16:20371089 20371938(-)	exon	100792777	LOC100792777
circRNA101	chr15:12786696 12787221(+)	exon	100784626	LOC100784626
circRNA102	chr6:44787569 44831144(-)	intergenic region	n/a	-
circRNA103	chr20:40308573 40316905(+)	intergenic region	n/a	-
circRNA104	chr19:5539449 5646201(+)	intergenic region	n/a	-

circRNA105	chr13:2011919 2012373(-)	exon	100783198	LOC100783198
circRNA106	chrPltd:292 532(-)	intergenic region	n/a	-
circRNA107	chr6:10377283 10378900(+)	exon	100815774	LOC100815774
circRNA108	chr7:35234977 35235530(+)	exon	100798214	LOC100798214
circRNA109	chrPltd:542 1151(-)	exon	3989259	psbA
circRNA110	chr3:44106649 44107117(-)	exon	100790207	LOC100790207
circRNA111	chr9:18439107 18505640(-)	intergenic region	n/a	-
circRNA112	chr15:23326207 23327192(-)	exon	100798669	LOC100798669
circRNA113	chr11:10552718 10554646(-)	exon	100797436	LOC100797436
circRNA114	chr1:40769039 40810598(+)	intergenic region	n/a	-
circRNA115	chr6:22933744 22935605(-)	exon	100783185	LOC100783185
circRNA116	chr9:191797 195693(-)	exon	100787334	LOC100787334
circRNA117	chr11:794904 795841(-)	exon	100776523	LOC100776523
circRNA118	chr4:41849024 42290025(+)	intergenic region	n/a	-
circRNA119	chr8:14980418 14982102(+)	exon	100795772	LOC100795772
circRNA120	chr20:40309646 40317081(+)	intergenic region	n/a	-
circRNA121	chr10:31033852 31120951(-)	intergenic region	n/a	-
circRNA122	chr18:2172273 2181785(-)	intergenic region	n/a	-
circRNA123	chr5:31230154 31230868(+)	exon	100810940	LOC100810940
circRNA124	chr1:55927718 56185545(+)	intergenic region	n/a	-
circRNA125	chr17:3261202 3263694(+)	exon	100782540	LOC100782540
circRNA126	chr13:37975092 37985968(-)	intergenic region	n/a	-
circRNA127	chr9:64393 81608(+)	intergenic region	n/a	-
circRNA128	chr9:10053318 10055815(-)	exon	100777704	LOC100777704
circRNA129	chr4:3174493 3189631(-)	intergenic region	n/a	-
circRNA130	chr13:14428824 14448409(+)	intergenic region	n/a	-
circRNA131	chr7:17921799 17935529(-)	intergenic region	n/a	-
circRNA132	chr15:9813151 9818094(+)	intergenic region	n/a	-
circRNA133	chr7:5657042 5757743(-)	intron	100780862	LOC100780862
circRNA134	chr5:41257574 41258431(+)	exon	100819489	LOC100819489
circRNA135	chr19:37738765 37740055(-)	exon	100794420	LOC100794420
circRNA136	chrPltd:1334 6331(-)	intergenic region	n/a	-
circRNA137	chr3:28917416 28918382(+)	exon	100786632	LOC100786632
circRNA138	chr11:5927662 5935738(+)	intergenic region	n/a	-
circRNA139	chr16:31135686 31161545(-)	intergenic region	n/a	-
circRNA140	chr12:39171381 39201509(-)	intergenic region	n/a	-
circRNA141	chr19:39072981 39073308(+)	exon	100809299	LOC100809299
circRNA142	chr9:27017380 27026534(-)	intergenic region	n/a	-
circRNA143	chr18:47705623 47726024(-)	intergenic region	n/a	-
circRNA144	chr9:33972255 33996509(+)	intergenic region	n/a	-
circRNA145	chrPltd:9507 9631(+)	intergenic region	n/a	-
circRNA146	chr20:3772851 37802721(-)	intergenic region	n/a	-
circRNA147	chr16:31516460 31524228(-)	intergenic region	n/a	-
circRNA148	chr19:45711045 45731347(+)	intergenic region	n/a	-
circRNA149	chr3:4545901 4597563(+)	intergenic region	n/a	-
circRNA150	chr13:10973964 10974580(+)	exon	100786712	LOC100786712
circRNA151	chr16:31124428 31159320(-)	intergenic region	n/a	-
circRNA152	chr4:3176618 3194504(-)	intergenic region	n/a	-
circRNA153	chr4:19122731 19124097(-)	exon	100781333	LOC100781333
circRNA154	chr13:30370619 30439929(-)	intergenic region	n/a	-
circRNA155	chrPltd:626 132481(-)	intergenic region	n/a	-
circRNA156	chr3:40132578 40132842(-)	exon	100812152	LOC100812152
circRNA157	chr8:20205875 20206460(-)	exon	100819005	LOC100819005
circRNA158	chr20:19887772 19902316(-)	intergenic region	n/a	-
circRNA159	chr10:20673681 20682531(-)	exon	100798812	LOC100798812
circRNA160	chr3:1326887 1339399(+)	intergenic region	n/a	-

circRNA161	chr16:14844159 14848870(+)	intergenic region	n/a	-
circRNA162	chr11:27859973 27860888(-)	exon	100798309	LOC100798309
circRNA163	chr8:8369331 8516096(-)	intergenic region	n/a	-
circRNA164	chrPltd:1334 5484(-)	intergenic region	n/a	-
circRNA165	chr20:44394506 44395149(+)	exon	100802781	LOC100802781
circRNA166	chr6:50840970 50844495(-)	exon	100776945	LOC100776945
circRNA167	chr8:30760157 30768879(-)	exon	100820096	LOC100820096
circRNA168	chr14:29788489 30062860(-)	intergenic region	n/a	-
circRNA169	chr20:1005654 1021407(+)	intergenic region	n/a	-
circRNA170	chr19:37242977 37251761(-)	exon	100785430	LOC100785430
circRNA171	chr20:34004446 34029487(+)	intergenic region	n/a	-
circRNA172	chr9:64104 81299(+)	intergenic region	n/a	-
circRNA173	chr7:17922785 17935680(-)	intergenic region	n/a	-
circRNA174	chr3:14468483 14468812(+)	exon	102668539	LOC102668539
circRNA175	chr11:7552065 7559088(-)	intergenic region	n/a	-
circRNA176	chrPltd:5461 5622(-)	exon	3989271	rbcL
circRNA177	chr1:40786535 40827205(+)	intergenic region	n/a	-
circRNA178	chr20:2506759 2549623(-)	intergenic region	n/a	-
circRNA179	chrPltd:133206 133620(-)	intron	3989373	trnI-UAG
circRNA180	chr6:4483790 4494067(-)	intron	100785124	LOC100785124
circRNA181	chr12:32096391 32100150(+)	exon	100800447	LOC100800447
circRNA182	chr14:40928724 40956103(-)	intergenic region	n/a	-
circRNA183	chr18:46934714 46936264(-)	exon	100305446	LOC100305446
circRNA184	chr3:1328507 1341190(+)	intergenic region	n/a	-
circRNA185	chr3:28917416 28918379(+)	exon	100786632	LOC100786632
circRNA186	chr3:32868274 32911501(+)	intergenic region	n/a	-
circRNA187	NW_014574060.1:521 3875(+)	intergenic region	n/a	-
circRNA188	chr4:13791756 13793984(-)	exon	100794972	LOC100794972
circRNA189	chr16:4913081 4926458(+)	intergenic region	n/a	-
circRNA190	chr2:47565637 47580782(-)	intergenic region	n/a	-
circRNA191	chr2:46152749 46154820(+)	exon	100820132	LOC100820132
circRNA192	chr8:44911922 44917185(-)	intergenic region	n/a	-
circRNA193	chr5:36906191 36906573(-)	exon	100801361	LOC100801361
circRNA194	chr7:15549263 15553072(+)	exon	100803526	LOC100803526
circRNA195	chr15:47836263 47843921(-)	intergenic region	n/a	-
circRNA196	chr15:3669200 3670897(-)	exon	100815107	LOC100815107
circRNA197	chr16:2978592 3003970(+)	intergenic region	n/a	-
circRNA198	chr20:47237921 47239194(-)	exon	100803121	LOC100803121
circRNA199	chrPltd:430 998(-)	exon	3989259	psbA
circRNA200	chr2:11466195 11523591(-)	intergenic region	n/a	-
circRNA201	chr10:863419 864940(-)	exon	100820160	LOC100820160
circRNA202	chr18:9735298 9750657(+)	intergenic region	n/a	-
circRNA203	chr11:30285324 30314856(-)	intergenic region	n/a	-
circRNA204	chr17:32909354 32910593(-)	exon	100788741	LOC100788741
circRNA205	NW_014573410.1:47527 56640(+)	intergenic region	n/a	-
circRNA206	chr4:102297 103240(+)	exon	100819848	LOC100819848
circRNA207	chr11:10376551 10397935(-)	intergenic region	n/a	-
circRNA208	chr7:2201472 2213268(+)	intergenic region	n/a	-
circRNA209	chr12:4359910 4387150(-)	intergenic region	n/a	-
circRNA210	chr4:7315218 7324979(+)	intergenic region	n/a	-
circRNA211	chr2:11468224 11557778(-)	intergenic region	n/a	-
circRNA212	chr6:14547681 14549278(+)	exon	100807766	LOC100807766
circRNA213	chrPltd:5246 5927(-)	intergenic region	n/a	-
circRNA214	chr1:52131183 52131984(+)	exon	100792600	LOC100792600
circRNA215	chr15:9813383 9819778(+)	intergenic region	n/a	-
circRNA216	chr16:2971678 2992343(+)	intergenic region	n/a	-

circRNA217	chr2:21066335 21066881(-)	exon	100785390	LOC100785390
circRNA218	chr2:32249856 32251696(+)	exon	100788765	LOC100788765
circRNA219	chr10:22028105 22029156(-)	exon	100793515	LOC100793515
circRNA220	chr12:36484497 36489389(+)	exon	100819307	LOC100819307
circRNA221	chr12:39176011 39206326(-)	intergenic region	n/a	-
circRNA222	chr20:34004505 34029487(+)	intergenic region	n/a	-
circRNA223	chr9:31173293 31173778(-)	exon	100816030	LOC100816030
circRNA224	chr1:53842717 53844884(+)	exon	100807112	LOC100807112
circRNA225	chr13:13639664 13681722(-)	intergenic region	n/a	-
circRNA226	chr8:15951432 16038560(-)	intergenic region	n/a	-
circRNA227	chr20:385688 391272(+)	exon	100789712	LOC100789712
circRNA228	chr1:3461288 3492739(+)	intergenic region	n/a	-
circRNA229	chr7:5583072 5614007(-)	exon	100808315	LOC100808315
circRNA230	chrPltd:1179 5347(-)	intergenic region	n/a	-
circRNA231	chr15:12790117 12790338(+)	intron	100784626	LOC100784626
circRNA232	NW_014573484.1:28490 37523(-)	intergenic region	n/a	-
circRNA233	chrPltd:633 5964(-)	intergenic region	n/a	-
circRNA234	chr7:5656878 5731898(-)	intron	100780862	LOC100780862
circRNA235	chr17:18178233 18186666(+)	exon	100800186	ZEP
circRNA236	chr3:23066214 23066760(+)	exon	100780970	LOC100780970
circRNA237	chrPltd:9840 10263(+)	intron	3989275	trnV-UAC
circRNA238	chr2:11526908 11556538(-)	intergenic region	n/a	-
circRNA239	chr10:45606873 45608241(-)	exon	100812684	LOC100812684
circRNA240	chr1:40785159 40810914(+)	intergenic region	n/a	-
circRNA241	chr14:21814581 21829377(-)	exon	100779257	LOC100779257
circRNA242	NW_014573413.1:165611 166172(+)	exon	100811292	LOC100811292
circRNA243	chr10:51526197 51526934(-)	exon	100798296	LOC100798296
circRNA244	chr9:49712857 49713414(-)	exon	102664796	LOC102664796
circRNA245	chr9:2391817 2392758(+)	exon	778173	LOC778173
circRNA246	chr15:5590004 5600707(+)	intergenic region	n/a	-
circRNA247	chr14:16760177 16762546(-)	exon	100798031	LOC100798031
circRNA248	chr17:8194399 8197820(-)	exon	100780025	LOC100780025
circRNA249	chr19:777173 777327(+)	exon	100781868	LOC100781868
circRNA250	chr13:9580102 9580490(+)	intergenic region	n/a	-
circRNA251	chr11:33098778 33099370(+)	exon	100813586	LOC100813586
circRNA252	chr4:9878470 9886594(+)	intergenic region	n/a	-
circRNA253	chr19:49181759 49191822(-)	intergenic region	n/a	-
circRNA254	chr8:818630 818893(-)	exon	100816702	LOC100816702
circRNA255	chr18:49532142 49539485(+)	intergenic region	n/a	-
circRNA256	chr2:11526158 11588303(-)	intergenic region	n/a	-
circRNA257	chr15:13671345 13671512(-)	intergenic region	n/a	-
circRNA258	chr20:47873733 47874480(-)	exon	100779579	LOC100779579
circRNA259	chr10:7879254 7881864(+)	exon	100793363	LOC100793363
circRNA260	chr8:13601186 13612458(+)	intergenic region	n/a	-
circRNA261	chr1:5506067 5508778(+)	exon	100807287	LOC100807287
circRNA262	chr18:9300508 9301248(+)	exon	106797140	LOC106797140
circRNA263	chr16:35732103 35740925(+)	exon	100810667	LOC100810667
circRNA264	chr1:53935062 53936075(+)	exon	100790851	LOC100790851
circRNA265	chrPltd:5419 5598(-)	exon	3989271	rbcL
circRNA266	chr7:1333801 1345817(-)	intergenic region	n/a	-
circRNA267	chr4:19464519 19465559(+)	exon	100782956	LOC100782956
circRNA268	chr6:7553758 7554264(-)	exon	100796777	LOC100796777
circRNA269	chrPltd:572 5598(-)	intergenic region	n/a	-
circRNA270	chrPltd:1211 5927(-)	intergenic region	n/a	-
circRNA271	chr15:35365065 35367804(-)	exon	100797874	LOC100797874
circRNA272	chr10:22028105 22029151(-)	exon	100793515	LOC100793515

circRNA273	chr6:10308428 10308900(+)	exon	100814187	LOC100814187
circRNA274	chrPltd:992 5418(-)	intergenic region	n/a	-
circRNA275	chr14:8964282 8986373(-)	intergenic region	n/a	-
circRNA276	chr3:27913096 27921314(-)	exon	100813936	LOC100813936
circRNA277	chr10:3959982 3961351(+)	exon	100794242	LOC100794242
circRNA278	chr2:6424549 6436781(-)	intergenic region	n/a	-
circRNA279	chr15:3178780 3183716(+)	intergenic region	n/a	-
circRNA280	chr3:34288577 34293474(+)	exon	548016	LOC548016
circRNA281	chr7:1337588 1349472(-)	intergenic region	n/a	-
circRNA282	chr3:4570759 4894600(+)	intergenic region	n/a	-
circRNA283	chrPltd:1334 6030(-)	intergenic region	n/a	-
circRNA284	NW_014573403.1:3476693 3480114(+)	exon	100786845	LOC100786845
circRNA285	chr6:47141085 47141651(+)	exon	100786541	LOC100786541
circRNA286	chrPltd:6798 32218(-)	intergenic region	n/a	-
circRNA287	chr13:28041005 28047156(+)	intron	102661733	LOC102661733
circRNA288	chr19:48928138 48968214(-)	intergenic region	n/a	-
circRNA289	chr12:4360128 4387377(-)	intergenic region	n/a	-
circRNA290	chr4:23760483 23761143(+)	exon	100812174	LOC100812174
circRNA291	chr17:40827810 40869109(+)	intergenic region	n/a	-
circRNA292	chr13:10447778 10470877(-)	intergenic region	n/a	-
circRNA293	chr20:37122805 37131872(+)	intergenic region	n/a	-
circRNA294	chr16:34412557 34682363(-)	intergenic region	n/a	-
circRNA295	chr8:11501656 11507851(+)	intergenic region	n/a	-
circRNA296	chr13:30007848 30023925(+)	intergenic region	n/a	-
circRNA297	chr8:30765388 30766580(-)	exon	100820096	LOC100820096
circRNA298	chr5:3981095 4011431(+)	intergenic region	n/a	-
circRNA299	chr6:45102685 45254175(+)	intergenic region	n/a	-
circRNA300	chr16:923887 931704(+)	intergenic region	n/a	-
circRNA301	chr6:11318309 11325740(-)	intergenic region	n/a	-
circRNA302	chr19:36659723 36664203(+)	exon	100793739	LOC100793739
circRNA303	chr1:46803408 46803829(-)	intron	106794272	LOC106794272
circRNA304	chrPltd:5776 6042(-)	exon	3989271	rbcL
circRNA305	chr4:137984 138510(-)	exon	100788288	LOC100788288
circRNA306	chr8:37078609 37092568(+)	intergenic region	n/a	-
circRNA307	chr12:39185836 39186051(+)	intergenic region	n/a	-
circRNA308	chrPltd:282 5526(-)	intergenic region	n/a	-
circRNA309	chr1:56072739 56076395(-)	exon	100799306	LOC100799306
circRNA310	chr8:7420347 7440559(-)	intergenic region	n/a	-
circRNA311	chr20:46162027 46163376(+)	exon	100818409	LOC100818409
circRNA312	chr7:5818456 5888529(+)	intergenic region	n/a	-
circRNA313	chr6:10302651 10308900(+)	exon	100814187	LOC100814187
circRNA314	chr3:28917416 28918014(+)	exon	100786632	LOC100786632
circRNA315	chrPltd:377 5347(-)	intergenic region	n/a	-
circRNA316	chr6:40400733 40400774(-)	exon	100799589	LOC100799589
circRNA317	chr4:8324414 8325968(-)	exon	100800444	LOC100800444
circRNA318	NW_014573484.1:24753 33868(-)	intergenic region	n/a	-
circRNA319	chr16:18633081 18640675(-)	exon	100804783	LOC100804783
circRNA320	chr11:2399523 2408669(-)	intergenic region	n/a	-
circRNA321	chr8:8532001 8535752(+)	intergenic region	n/a	-
circRNA322	chr12:3933041 3939585(+)	intergenic region	n/a	-
circRNA323	chr14:1314153 1334576(-)	intergenic region	n/a	-
circRNA324	chr13:10446662 10468981(-)	intergenic region	n/a	-
circRNA325	chr3:1158932 1210925(-)	intergenic region	n/a	-
circRNA326	chr3:42262802 42267433(+)	exon	100789841	LOC100789841
circRNA327	chr2:11464189 11584195(-)	intergenic region	n/a	-
circRNA328	chr10:45843564 45857438(+)	intergenic region	n/a	-



circRNA329	chr4:36385564 36390772(+)	exon	100802776	LOC100802776
circRNA330	chr4:7317376 7327424(+)	intergenic region	n/a	-
circRNA331	chr6:1103449 1109238(-)	intron	100810431	LOC100810431
circRNA332	chr8:3719682 3720386(+)	intergenic region	n/a	-
circRNA333	chr10:48018585 48019731(-)	exon	100527205	LOC100527205
circRNA334	chr6:11227827 11233143(-)	intergenic region	n/a	-
circRNA335	chr7:49936 51389(+)	exon	100809207	LOC100809207
circRNA336	chr3:39375739 39377161(-)	exon	100813574	LOC100813574
circRNA337	chr6:4486683 4496806(-)	intron	100785124	LOC100785124
circRNA338	chrPltd:1334 6179(-)	intergenic region	n/a	-
circRNA339	chrPltd:1049 5769(-)	intergenic region	n/a	-
circRNA340	chr8:7188226 7188902(+)	exon	100792580	LOC100792580
circRNA341	chr1:40785846 40826530(+)	intergenic region	n/a	-
circRNA342	chr20:1006131 1021407(+)	intergenic region	n/a	-
circRNA343	chr15:915450 936881(+)	intergenic region	n/a	-
circRNA344	chr8:16365101 16376243(-)	intergenic region	n/a	-
circRNA345	chr8:75121 76650(-)	exon	102666240	LOC102666240
circRNA346	chr8:47796962 47797769(-)	exon	100819381	LOC100819381
circRNA347	chr6:49814463 49828200(+)	intergenic region	n/a	-
circRNA348	chr4:36567203 36567727(-)	exon	100806823	LOC100806823
circRNA349	chr10:51486078 51486491(-)	exon	100796688	LOC100796688
circRNA350	chr3:28917416 28917704(+)	exon	100786632	LOC100786632
circRNA351	chr20:12826184 12829486(-)	exon	100810922	LOC100810922
circRNA352	chr14:8966193 8988940(-)	intergenic region	n/a	-
circRNA353	chr8:42021557 42025348(+)	exon	100797527	LOC100797527
circRNA354	chr18:56252463 56269397(-)	intergenic region	n/a	-
circRNA355	chr3:44497299 44500474(-)	intergenic region	n/a	-
circRNA356	chr18:53367914 53372053(-)	intergenic region	n/a	-
circRNA357	chr1:40771890 40773726(-)	intron	100798947	LOC100798947
circRNA358	chr6:14637715 14640541(+)	exon	100809371	LOC100809371
circRNA359	chr11:571108 574697(+)	exon	100811424	LOC100811424
circRNA360	chr15:46896807 46974363(-)	intergenic region	n/a	-
circRNA361	chr15:47836267 47843921(-)	intergenic region	n/a	-
circRNA362	chr2:11464363 11552345(-)	intergenic region	n/a	-
circRNA363	chr2:4502992 4513141(-)	intergenic region	n/a	-
circRNA364	chr19:35364051 35365400(+)	exon	100819645	LOC100819645
circRNA365	chr11:813724 815308(+)	exon	100787912	LOC100787912
circRNA366	chr17:35897816 36081885(-)	intergenic region	n/a	-
circRNA367	chrPltd:6079 6579(-)	exon	3989271	rbcL
circRNA368	chrPltd:102870 103328(+)	intron	3989347	trnA-UGC
circRNA369	chr6:45102182 45254042(+)	intergenic region	n/a	-
circRNA370	chr5:35142714 35143144(+)	exon	100810237	LOC100810237
circRNA371	chr16:4790284 4799283(+)	intergenic region	n/a	-
circRNA372	chr11:2577854 2579263(-)	exon	100779024	LOC100779024
circRNA373	NW_014573484.1:34721 63753(-)	intergenic region	n/a	-
circRNA374	chr1:14066764 14067633(-)	exon	100797182	LOC100797182
circRNA375	chr12:30691510 30693925(+)	exon	100782437	LOC100782437
circRNA376	chr8:18737081 18738917(-)	exon	100792410	LOC100792410
circRNA377	chr14:8138504 8177289(-)	intergenic region	n/a	-
circRNA378	chr2:48058096 48059921(+)	exon	100781655	LOC100781655
circRNA379	chr6:4487693 4497730(-)	intron	100785124	LOC100785124
circRNA380	chr10:39936497 39937548(+)	exon	100794911	LOC100794911
circRNA381	chr8:33257202 33258020(+)	exon	100786931	LOC100786931
circRNA382	chr12:4470195 4490394(+)	intergenic region	n/a	-
circRNA383	chr1:51532362 51538884(-)	intergenic region	n/a	-
circRNA384	chr6:23193531 23197354(-)	exon	100789027	LOC100789027

circRNA385	chr20:2814577 2815634(+)	exon	100814882	LOC100814882
circRNA386	chr8:37880531 37882399(+)	exon	100796816	LOC100796816
circRNA387	chrPltd:1181 6611(-)	intergenic region	n/a	-
circRNA388	chr12:1287231 1293233(+)	intergenic region	n/a	-
circRNA389	chr18:55772619 55773937(-)	exon	100793178	LOC100793178
circRNA390	chr10:30293099 30310511(+)	exon	100798991	LOC100798991
circRNA391	chr17:3800722 3802514(+)	exon	100801645	LOC100801645
circRNA392	chr3:41848632 41849015(+)	exon	100784188	LOC100784188
circRNA393	chr6:2636196 2643514(+)	intergenic region	n/a	-
circRNA394	chr7:17920647 17933805(-)	intergenic region	n/a	-
circRNA395	chrPltd:58664 58709(+)	intergenic region	n/a	-
circRNA396	chr14:2860491 2876806(+)	intergenic region	n/a	-
circRNA397	chrPltd:6617 62647(-)	intergenic region	n/a	-
circRNA398	chr6:18324693 18398819(-)	intergenic region	n/a	-
circRNA399	chr20:384307 387477(+)	exon	100789712	LOC100789712
circRNA400	chr11:10012736 10013350(-)	exon	100789855	LOC100789855
circRNA401	chr3:18838854 18840058(-)	exon	100808588	LOC100808588
circRNA402	chr12:1215614 1220684(-)	intergenic region	n/a	-
circRNA403	chr3:10814925 10816669(-)	exon	100784706	LOC100784706
circRNA404	chr1:16836073 16837051(+)	exon	100796646	LOC100796646
circRNA405	chr18:50041099 50043084(+)	exon	100807514	LOC100807514
circRNA406	chr19:834176 839119(+)	intergenic region	n/a	-
circRNA407	chr8:43752918 43761102(-)	intergenic region	n/a	-
circRNA408	chr20:37121678 37130681(+)	intergenic region	n/a	-
circRNA409	chr8:14047911 14048116(+)	exon	100777674	LOC100777674
circRNA410	chr9:43216289 43217581(+)	exon	100800190	LOC100800190
circRNA411	chr18:57756398 57756679(-)	exon	100818553	LOC100818553
circRNA412	chr3:2832111 2896774(+)	intergenic region	n/a	-
circRNA413	chr20:47802450 47802973(-)	exon	100306581	LOC100306581
circRNA414	chr15:33417004 33427556(+)	exon	100812785	LOC100812785
circRNA415	chr2:1938628 1939750(-)	exon	100812482	LOC100812482
circRNA416	chr11:6225224 6228417(+)	exon	100819979	LOC100819979
circRNA417	chr9:5778704 5782165(+)	exon	100820121	LOC100820121
circRNA418	chr4:2553357 2555253(-)	exon	100796383	LOC100796383
circRNA419	chr20:46343586 46346482(+)	exon	100777257	LOC100777257
circRNA420	chr9:3677492 3684587(-)	intergenic region	n/a	-
circRNA421	chr3:38374249 38375242(+)	exon	100795648	LOC100795648
circRNA422	chr1:1094199 1136368(-)	intergenic region	n/a	-
circRNA423	chr5:35568721 35578613(-)	intergenic region	n/a	-
circRNA424	chr12:32986247 32987034(-)	exon	100819481	LOC100819481
circRNA425	chr11:574273 574697(+)	exon	100811424	LOC100811424
circRNA426	chr8:44901395 44911975(-)	intergenic region	n/a	-
circRNA427	NW_014573484.1:34349 63189(-)	intergenic region	n/a	-
circRNA428	chr3:9574537 9786039(-)	intergenic region	n/a	-
circRNA429	chrPltd:1334 6819(-)	intergenic region	n/a	-
circRNA430	chr16:924014 932357(+)	intergenic region	n/a	-
circRNA431	chr15:45648205 45649041(-)	exon	100800878	LOC100800878
circRNA432	chr1:40768871 40865437(+)	intergenic region	n/a	-
circRNA433	chr2:38755328 38755758(+)	exon	100780041	LOC100780041
circRNA434	chr17:8728097 8729452(-)	exon	100802507	LOC100802507
circRNA435	chr1:55134092 55137396(-)	exon	100780539	LOC100780539
circRNA436	chr16:33923291 33926590(-)	intergenic region	n/a	-
circRNA437	chr8:46678777 46681744(-)	intergenic region	n/a	-
circRNA438	chr17:4494181 4495631(-)	exon	102665860	LOC102665860
circRNA439	chr17:24453911 24469413(+)	exon	100775929	LOC100775929
circRNA440	chr8:8532014 8535752(+)	intergenic region	n/a	-

circRNA441	chr16:20941272 20941776(+)	exon	100818307	LOC100818307
circRNA442	chr10:2371824 2372135(+)	exon	100808741	LOC100808741
circRNA443	chrPltd:542 6911(-)	intergenic region	n/a	-
circRNA444	chr19:7190036 7254196(+)	intergenic region	n/a	-
circRNA445	chr1:14666006 14667229(+)	exon	100790337	LOC100790337
circRNA446	chr20:14775471 14777079(-)	exon	100813099	LOC100813099
circRNA447	chr8:3719672 3720386(+)	intergenic region	n/a	-
circRNA448	chr17:8386898 8387265(+)	intergenic region	n/a	-
circRNA449	chr8:41285540 41286106(+)	exon	100788193	LOC100788193
circRNA450	chr9:30315905 30320737(-)	intergenic region	n/a	-
circRNA451	chr10:20673681 20675114(-)	exon	100798812	LOC100798812
circRNA452	chr20:2814582 2815634(+)	exon	100814882	LOC100814882
circRNA453	chrPltd:917 6111(-)	intergenic region	n/a	-
circRNA454	chr18:3487168 3490245(+)	exon	100813040	LOC100813040
circRNA455	chr2:11523527 11585788(-)	intergenic region	n/a	-
circRNA456	chr19:45710381 45730939(+)	intergenic region	n/a	-
circRNA457	chr16:35007029 35078566(+)	intergenic region	n/a	-
circRNA458	chr19:7331679 7333066(-)	exon	102668567	LOC102668567
circRNA459	chr18:9120644 9145234(+)	intergenic region	n/a	-
circRNA460	chr9:45043768 45064896(-)	intergenic region	n/a	-
circRNA461	chr6:47133711 47146242(+)	intergenic region	n/a	-
circRNA462	chr1:40765144 40863424(+)	intergenic region	n/a	-
circRNA463	chr17:27563226 27564727(-)	exon	100802522	LOC100802522
circRNA464	chr18:53641714 53679942(+)	intergenic region	n/a	-
circRNA465	chrPltd:1164 6911(-)	intergenic region	n/a	-
circRNA466	chr10:49533428 49537563(+)	exon	100812875	LOC100812875
circRNA467	chr4:8651533 8653804(+)	exon	100801328	LOC100801328
circRNA468	chr1:40765144 40806696(+)	intergenic region	n/a	-
circRNA469	chr13:6820332 6841031(+)	exon	100781560	LOC100781560
circRNA470	chr16:4791276 4800308(+)	intergenic region	n/a	-
circRNA471	chr14:4501144 4513332(+)	intergenic region	n/a	-
circRNA472	chr17:15105982 15124542(-)	intergenic region	n/a	-
circRNA473	chrPltd:9820 10336(+)	intergenic region	n/a	-
circRNA474	chr13:30369914 30439244(-)	intergenic region	n/a	-
circRNA475	chr8:15967756 16006670(+)	intergenic region	n/a	-
circRNA476	chr20:34006730 34030841(+)	intergenic region	n/a	-
circRNA477	chr7:5598591 5642775(-)	exon	100780862	LOC100780862
circRNA478	chr13:14433658 14450955(+)	intergenic region	n/a	-
circRNA479	chrPltd:542 5567(-)	intergenic region	n/a	-
circRNA480	chrPltd:6007 6723(-)	exon	3989271	rbcL
circRNA481	NW_014573484.1:16062 64105(-)	intergenic region	n/a	-
circRNA482	chr2:42885381 42885672(+)	intron	100791567	LOC100791567
circRNA483	chr7:6618814 6642147(+)	intergenic region	n/a	-
circRNA484	chr20:2506759 2550951(-)	intergenic region	n/a	-
circRNA485	chr16:2984274 3010428(+)	intergenic region	n/a	-
circRNA486	chr6:49818526 49828200(+)	intergenic region	n/a	-
circRNA487	chr7:1332233 1344285(-)	intergenic region	n/a	-
circRNA488	chrPltd:959 6423(-)	intergenic region	n/a	-
circRNA489	chr11:841379 842960(+)	exon	100799181	LOC100799181
circRNA490	chrPltd:1046 6150(-)	intergenic region	n/a	-
circRNA491	chr9:41106857 41109855(+)	exon	100782181	LOC100782181
circRNA492	chr17:35521047 35522019(+)	exon	102668239	LOC102668239
circRNA493	chr7:1338146 1349989(-)	intergenic region	n/a	-
circRNA494	chr10:5027202 5028970(+)	exon	100817671	LOC100817671
circRNA495	chr8:42087512 42088359(-)	intron	102664025	LOC102664025
circRNA496	chr6:29627338 29630373(+)	exon	100792534	LOC100792534

circRNA497	chr1:11290327 11291065(+)	exon	100785017	LOC100785017
circRNA498	chr3:2879639 2950454(+)	intergenic region	n/a	-
circRNA499	chr2:24765266 24765892(-)	exon	100807496	LOC100807496
circRNA500	NW_014573484.1:18217 25651(-)	intron	100499660	LOC100499660
circRNA501	chrPltd:1164 5496(-)	intergenic region	n/a	-
circRNA502	chr18:44967265 44971082(+)	exon	100775230	LOC100775230
circRNA503	chr14:28564345 28566845(+)	exon	100779093	LOC100779093
circRNA504	chr18:47025919 47057559(+)	intergenic region	n/a	-
circRNA505	chrPltd:537 919(-)	exon	3989259	psbA
circRNA506	chr16:31124428 31159292(-)	intergenic region	n/a	-
circRNA507	chr11:12533077 12534023(-)	exon	100800977	LOC100800977
circRNA508	chr2:11470224 11618890(-)	intergenic region	n/a	-
circRNA509	chr4:6449418 6452039(+)	exon	100790916	LOC100790916
circRNA510	chr10:51439654 51441948(+)	exon	100795105	LOC100795105
circRNA511	chr7:5616468 5737802(-)	intron	100780862	LOC100780862
circRNA512	chr10:30293099 30314447(+)	exon	100798991	LOC100798991
circRNA513	chr18:12237604 12587876(+)	intergenic region	n/a	-
circRNA514	chr4:45441851 45442249(-)	exon	100794615	LOC100794615
circRNA515	NW_014573484.1:27849 35247(-)	intergenic region	n/a	-
circRNA516	chrPltd:13750 14143(-)	intron	3989280	trnL-UAA
circRNA517	chr6:39232956 39234731(+)	exon	100806517	LOC100806517
circRNA518	chr1:40786124 40883182(+)	intergenic region	n/a	-
circRNA519	chr1:3456828 3487889(+)	intergenic region	n/a	-
circRNA520	chr9:37589871 37594714(-)	intergenic region	n/a	-
circRNA521	chr3:2832111 2879746(+)	intergenic region	n/a	-
circRNA522	chr2:1807162 1808637(-)	exon	100808200	LOC100808200
circRNA523	chr2:11527971 11618890(-)	intergenic region	n/a	-
circRNA524	chr8:16365515 16377618(-)	intergenic region	n/a	-
circRNA525	chr20:45575598 45576203(-)	intergenic region	n/a	-
circRNA526	chr13:42278004 42279321(-)	exon	100803144	LOC100803144
circRNA527	chr11:8483683 8497403(-)	intergenic region	n/a	-
circRNA528	chr10:45052893 45053786(-)	exon	100815875	LOC100815875
circRNA529	chr13:14936060 14948575(-)	intergenic region	n/a	-
circRNA530	chr20:24827235 24830481(+)	exon	100815591	LOC100815591
circRNA531	chr15:11462686 11478125(-)	intergenic region	n/a	-
circRNA532	chrPltd:671 5398(-)	intergenic region	n/a	-
circRNA533	chrPltd:1049 5721(-)	intergenic region	n/a	-
circRNA534	chr15:919396 942380(+)	intergenic region	n/a	-
circRNA535	chr18:51497586 51620823(+)	intergenic region	n/a	-
circRNA536	chr2:11590205 11618890(-)	intergenic region	n/a	-
circRNA537	chrPltd:731 6721(-)	intergenic region	n/a	-
circRNA538	chr9:43219537 43223182(+)	exon	100800190	LOC100800190
circRNA539	chr16:26320094 26321368(-)	exon	100775901	LOC100775901
circRNA540	chr14:21796188 21797950(-)	exon	100779257	LOC100779257
circRNA541	chr9:45043492 45064608(-)	intergenic region	n/a	-
circRNA542	chr3:9528007 9786039(-)	intergenic region	n/a	-
circRNA543	chr6:704808 709514(+)	exon	100785855	LOC100785855
circRNA544	chr11:12539404 12540944(-)	exon	100800977	LOC100800977
circRNA545	chrPltd:6617 13742(-)	intergenic region	n/a	-
circRNA546	chr16:35881396 35886401(+)	intergenic region	n/a	-
circRNA547	chr4:44929057 44931900(+)	exon	100788458	LOC100788458
circRNA548	chr3:465865 484729(-)	intron	100788960	LOC100788960
circRNA549	chr8:3341690 3341916(-)	exon	100814574	LOC100814574
circRNA550	chr19:34074859 34076290(-)	exon	100806108	LOC100806108
circRNA551	chr20:2503798 2548226(-)	intergenic region	n/a	-
circRNA552	chr19:38372706 38378483(-)	intergenic region	n/a	-

circRNA553	chr1:53591944 53601538(-)	exon	100796650	LOC100796650
circRNA554	chr2:42885773 42886064(+)	intron	100791567	LOC100791567
circRNA555	chr8:45415924 45430992(-)	intergenic region	n/a	-
circRNA556	chr6:45214792 45238902(+)	intergenic region	n/a	-
circRNA557	chr12:39184138 39184692(+)	intergenic region	n/a	-
circRNA558	chr5:3977468 4003127(+)	intergenic region	n/a	-
circRNA559	chrPltd:1334 5452(-)	intergenic region	n/a	-
circRNA560	chr10:1958039 1968140(-)	intergenic region	n/a	-
circRNA561	chr1:3461948 3504862(+)	intergenic region	n/a	-
circRNA562	chr19:38966003 38966542(-)	exon	100814485	LOC100814485
circRNA563	chr7:4931363 4934848(-)	exon	100808136	LOC100808136
circRNA564	chr9:187830 195693(-)	exon	100787334	LOC100787334
circRNA565	chr3:9489221 9786039(-)	intergenic region	n/a	-
circRNA566	chr8:42671873 42675652(-)	exon	100782497	LOC100782497
circRNA567	chr9:28752908 28756519(-)	exon	100791541	LOC100791541
circRNA568	chr4:1272169 1272550(-)	exon	100500173	LOC100500173
circRNA569	chr4:49076905 49081918(+)	exon	100801705	LOC100801705
circRNA570	chr9:30316147 30321342(-)	intergenic region	n/a	-
circRNA571	chr15:21056604 21057039(-)	exon	100807099	LOC100807099
circRNA572	chr14:34295690 34296306(+)	exon	100810450	LOC100810450
circRNA573	chr17:23824263 23833586(-)	exon	100818869	LOC100818869
circRNA574	chr18:51251234 51269583(+)	intergenic region	n/a	-
circRNA575	chr12:26068859 26068918(-)	exon	100796730	LOC100796730
circRNA576	chr3:39353007 39362821(+)	intergenic region	n/a	-
circRNA577	chr5:4118913 4132521(-)	intergenic region	n/a	-
circRNA578	chr11:8927977 8931149(-)	intergenic region	n/a	-
circRNA579	chr2:11494849 11587639(-)	intergenic region	n/a	-
circRNA580	chr10:15340377 15341056(-)	exon	100796678	LOC100796678
circRNA581	chr20:385741 391272(+)	exon	100789712	LOC100789712
circRNA582	chr20:32240050 32241606(+)	exon	100784393	LOC100784393
circRNA583	chr1:51378408 51379239(-)	exon	100808533	LOC100808533
circRNA584	chr2:24200228 24203368(-)	exon	100804313	LOC100804313
circRNA585	chr20:129203 131337(-)	exon	100781736	LOC100781736
circRNA586	chr19:37044897 37080087(+)	intergenic region	n/a	-
circRNA587	chrPltd:1049 5347(-)	intergenic region	n/a	-
circRNA588	chr8:23358371 23361893(+)	exon	100800879	LOC100800879
circRNA589	chrPltd:5246 6900(-)	intergenic region	n/a	-
circRNA590	chr14:1313759 1334359(-)	intergenic region	n/a	-
circRNA591	chrPltd:1181 6603(-)	intergenic region	n/a	-
circRNA592	chr15:4322950 4324170(+)	exon	100776255	LOC100776255
circRNA593	chr5:27718317 27718897(+)	exon	100819679	LOC100819679
circRNA594	chr13:14936060 14948150(-)	intergenic region	n/a	-
circRNA595	chr19:44906123 44918231(+)	intergenic region	n/a	-
circRNA596	chr7:42473036 42473578(+)	intergenic region	n/a	-
circRNA597	chr12:31887941 31888453(+)	exon	100801333	LOC100801333
circRNA598	chr18:17167950 17168533(+)	exon	102661320	LOC102661320
circRNA599	chr14:4760468 4761097(+)	exon	100795722	LOC100795722
circRNA600	chr18:47058227 47089306(+)	intergenic region	n/a	-
circRNA601	chr2:30936356 30936846(-)	exon	100804634	LOC100804634
circRNA602	chr14:1389083 1394515(+)	exon	100785869	LOC100785869
circRNA603	chr5:4117012 4132020(-)	intergenic region	n/a	-
circRNA604	chr7:1862042 1944358(-)	intergenic region	n/a	-
circRNA605	chr1:3461321 3504695(+)	intergenic region	n/a	-
circRNA606	chr16:33437015 33492350(+)	intergenic region	n/a	-
circRNA607	chr19:39072996 39073308(+)	exon	100809299	LOC100809299
circRNA608	chr11:18326812 18439336(-)	intergenic region	n/a	-

circRNA609	chr5:4116687 4131112(-)	intergenic region	n/a	-
circRNA610	chr15:20902156 20908672(-)	exon	100804432	LOC100804432
circRNA611	chr9:45042379 45063446(-)	intergenic region	n/a	-
circRNA612	chrMT:42344 42620(+)	intergenic region	n/a	-
circRNA613	chr9:45042640 45063769(-)	intergenic region	n/a	-
circRNA614	chr1:3493493 3504862(+)	intergenic region	n/a	-
circRNA615	chrPltd:102420 103021(+)	intergenic region	n/a	-
circRNA616	chr9:41496971 41497611(+)	exon	100775562	LOC100775562
circRNA617	chr20:372302 385614(+)	exon	100789712	LOC100789712
circRNA618	chr2:11525980 11616531(-)	intergenic region	n/a	-
circRNA619	chr14:28661848 28668769(+)	exon	100807603	LOC100807603
circRNA620	chr18:53635793 53672379(+)	intergenic region	n/a	-
circRNA621	chr18:53880118 53883466(-)	exon	100818890	LOC100818890
circRNA622	chr16:36808057 36812010(+)	exon	100800352	LOC100800352
circRNA623	chr18:41246638 41296638(-)	intergenic region	n/a	-
circRNA624	chr9:46987019 46987225(+)	exon	100776652	LOC100776652
circRNA625	chr12:39172032 39202220(-)	intergenic region	n/a	-
circRNA626	chr11:33084644 33104103(+)	intergenic region	n/a	-
circRNA627	chr17:23824263 23825603(-)	exon	100818869	LOC100818869
circRNA628	chr1:18850487 18852875(+)	exon	100799827	LOC100799827
circRNA629	chr7:5879136 5883809(+)	exon	100783381	LOC100783381
circRNA630	chr16:4790967 4799454(+)	intergenic region	n/a	-
circRNA631	chrPltd:10079 10213(+)	intron	3989275	trnV-UAC
circRNA632	chr16:32565209 32582792(+)	intergenic region	n/a	-
circRNA633	chrPltd:992 1349(-)	exon	3989259	psbA
circRNA634	chr20:34671296 34671744(-)	exon	100787418	LOC100787418
circRNA635	chr9:49013639 49014125(-)	intergenic region	n/a	-
circRNA636	chr2:12773477 12798698(+)	intergenic region	n/a	-
circRNA637	chr4:45004008 45006078(+)	exon	100788458	LOC100788458
circRNA638	chr1:49275216 49277073(+)	exon	100782701	LOC100782701
circRNA639	chr20:47359638 47359992(-)	intron	100804890	LOC100804890
circRNA640	chr15:1655721 1689193(+)	intergenic region	n/a	-
circRNA641	chr9:5181613 5181889(-)	exon	100809968	LOC100809968
circRNA642	chr6:4486042 4494546(-)	intron	100785124	LOC100785124
circRNA643	chr17:37182620 37200573(-)	intergenic region	n/a	-
circRNA644	chr11:30287670 30317704(-)	intergenic region	n/a	-
circRNA645	chr11:10052071 10063439(-)	intergenic region	n/a	-
circRNA646	chr14:8138838 8177736(-)	intergenic region	n/a	-
circRNA647	chr3:2880809 2899803(+)	intergenic region	n/a	-
circRNA648	chr9:65301 82486(+)	intergenic region	n/a	-
circRNA649	chr12:18579993 18959633(-)	intergenic region	n/a	-
circRNA650	chr17:15106671 15125224(-)	intergenic region	n/a	-
circRNA651	chr19:37222664 37225402(-)	exon	100785430	LOC100785430
circRNA652	chrPltd:18994 19115(+)	exon	3989266	psaA
circRNA653	NW_014573484.1:15243 24253(-)	intron	100499660	LOC100499660
circRNA654	chr19:48927123 48967266(-)	intergenic region	n/a	-
circRNA655	chr8:37880518 37882399(+)	intron	100796816	LOC100796816
circRNA656	chr6:42502018 42700635(+)	intergenic region	n/a	-
circRNA657	chr3:5475546 5946000(+)	intergenic region	n/a	-
circRNA658	chr12:5347411 5349201(+)	exon	100787759	LOC100787759
circRNA659	chr12:791788 805322(+)	intergenic region	n/a	-
circRNA660	chr11:16525588 16660259(+)	intergenic region	n/a	-
circRNA661	chr3:2833514 2898802(+)	intergenic region	n/a	-
circRNA662	chr6:45215558 45239558(+)	intergenic region	n/a	-
circRNA663	chrPltd:1334 52726(-)	intergenic region	n/a	-
circRNA664	chr3:2886488 2954409(+)	intergenic region	n/a	-

circRNA665	chr16:3423143 3424288(+)	exon	100819741	LOC100819741
circRNA666	chr9:42911177 42911692(-)	exon	100790691	LOC100790691
circRNA667	chr2:5914840 5917782(+)	exon	100776137	LOC100776137
circRNA668	chr11:11730051 11731824(+)	exon	100798497	LOC100798497
circRNA669	chr3:27920667 27921314(-)	exon	100813936	LOC100813936
circRNA670	chr11:5595343 5596154(-)	exon	100795117	LOC100795117
circRNA671	chr4:52054899 52056237(+)	exon	100775631	LOC100775631
circRNA672	chr14:9303047 9389492(-)	intergenic region	n/a	-
circRNA673	chr20:46232035 46233159(-)	exon	100816122	LOC100816122
circRNA674	chr13:36343547 36350053(+)	exon	100816142	LOC100816142
circRNA675	chrMT:47104 47375(+)	intergenic region	n/a	-
circRNA676	chr6:15350288 15350698(-)	exon	100778359	LOC100778359
circRNA677	NW_014573484.1:19046 28545(-)	intron	100499660	LOC100499660
circRNA678	NW_014573484.1:116 19533(-)	intergenic region	n/a	-
circRNA679	chr5:91365 93631(-)	exon	100798000	LOC100798000
circRNA680	chr2:11589139 11617353(-)	intergenic region	n/a	-
circRNA681	chrPltd:9820 10340(+)	intergenic region	n/a	-
circRNA682	chr2:13814031 13817247(+)	exon	100797559	LOC100797559
circRNA683	chrPltd:1334 6423(-)	intergenic region	n/a	-
circRNA684	chr4:41688385 41689222(+)	exon	100798329	LOC100798329
circRNA685	chr11:463972 464617(+)	exon	100808747	LOC100808747
circRNA686	chr15:46456111 46462169(+)	intergenic region	n/a	-
circRNA687	chr5:15714461 15715073(+)	exon	100787957	LOC100787957
circRNA688	chr6:11892438 11893851(-)	exon	100791330	LOC100791330
circRNA689	chr7:5307300 5313427(+)	intergenic region	n/a	-
circRNA690	chr7:10472719 10484541(-)	intergenic region	n/a	-
circRNA691	chr20:34018305 34034475(+)	intergenic region	n/a	-
circRNA692	chr20:25333849 25336387(+)	exon	100337655	LOC100337655
circRNA693	chr6:6457880 6461165(+)	intergenic region	n/a	-
circRNA694	chr20:45829100 45830084(+)	intergenic region	n/a	-
circRNA695	chr4:48019423 48020758(+)	exon	100810372	LOC100810372
circRNA696	chr19:1879350 1893556(+)	intergenic region	n/a	-
circRNA697	chr7:19647953 19648541(+)	exon	100807255	LOC100807255
circRNA698	chr8:44684764 44705621(+)	intergenic region	n/a	-
circRNA699	chr15:915233 936673(+)	intergenic region	n/a	-
circRNA700	chr13:22495827 22497063(+)	exon	100788136	LOC100788136
circRNA701	chr1:40763642 40803484(+)	intergenic region	n/a	-
circRNA702	chr8:44292005 44296495(+)	intergenic region	n/a	-
circRNA703	chr17:12179480 12181447(-)	exon	100780029	LOC100780029
circRNA704	chr5:682855 683335(+)	exon	100500448	LOC100500448
circRNA705	chr13:15500721 15501426(+)	exon	100792021	LOC100792021
circRNA706	chrPltd:1334 6190(-)	intergenic region	n/a	-
circRNA707	chr15:4750651 4752561(+)	exon	100793450	LOC100793450
circRNA708	chr1:40764103 40805360(+)	intergenic region	n/a	-
circRNA709	chr20:27308877 27395942(-)	intergenic region	n/a	-
circRNA710	chr1:40766818 40864749(+)	intergenic region	n/a	-
circRNA711	chr15:15637009 15654719(+)	intergenic region	n/a	-
circRNA712	chr15:11466928 11481396(-)	intergenic region	n/a	-
circRNA713	chr4:7316309 7326365(+)	intergenic region	n/a	-
circRNA714	chrPltd:1049 6642(-)	intergenic region	n/a	-
circRNA715	chr8:16370065 16382792(-)	intergenic region	n/a	-
circRNA716	chrPltd:1049 6582(-)	intergenic region	n/a	-
circRNA717	chrPltd:6376 6801(-)	intergenic region	n/a	-
circRNA718	chr3:39759591 39760220(+)	exon	100799358	LOC100799358
circRNA719	chr2:11527161 11618108(-)	intergenic region	n/a	-
circRNA720	chr17:39769754 39770767(+)	exon	100814610	LOC100814610

circRNA721	chr11:405214 413342(+)	intergenic region	n/a	-
circRNA722	chr12:14916220 14953212(-)	intergenic region	n/a	-
circRNA723	chr7:5883734 5883809(+)	exon	100783381	LOC100783381
circRNA724	chr1:40769208 40870116(+)	intergenic region	n/a	-
circRNA725	chr19:7331679 7333071(-)	exon	102668567	LOC102668567
circRNA726	chr12:16512715 16737230(+)	intergenic region	n/a	-
circRNA727	chr6:22933744 22939330(-)	exon	100783185	LOC100783185
circRNA728	chr9:45275349 45279479(-)	exon	100809429	LOC100809429
circRNA729	chrPltd:1046 6022(-)	intergenic region	n/a	-
circRNA730	chr13:42278598 42280386(-)	exon	100803144	LOC100803144
circRNA731	chrPltd:965 5831(-)	intergenic region	n/a	-
circRNA732	chr7:6616976 6639868(+)	intergenic region	n/a	-
circRNA733	chr17:34578204 34579654(-)	exon	100816565	LOC100816565
circRNA734	chr7:15549263 15551752(+)	exon	100803526	LOC100803526
circRNA735	chr7:5883746 5883809(+)	exon	100783381	LOC100783381
circRNA736	chr5:17918842 17919263(+)	exon	100791478	LOC100791478
circRNA737	chr1:40826151 40870281(+)	intergenic region	n/a	-
circRNA738	chr2:24200228 24211971(-)	exon	100804313	LOC100804313
circRNA739	chr11:10342035 10342821(-)	exon	100790051	LOC100790051
circRNA740	chr18:7034744 7036702(-)	exon	100809649	LOC100809649
circRNA741	chr20:35683676 35684970(+)	exon	100777969	LOC100777969
circRNA742	chrPltd:1334 5979(-)	intergenic region	n/a	-
circRNA743	chr11:15566414 15575817(-)	intergenic region	n/a	-
circRNA744	chr13:14412571 14430796(+)	intergenic region	n/a	-
circRNA745	chr17:32901153 32910593(-)	exon	100788741	LOC100788741
circRNA746	chr11:28468445 28514562(+)	intergenic region	n/a	-
circRNA747	chr20:36115002 36143608(-)	intergenic region	n/a	-
circRNA748	chr9:4010101 4010606(-)	exon	100779134	LOC100779134
circRNA749	chr3:45174287 45175859(+)	exon	100816236	LOC100816236
circRNA750	chr14:13655490 13724529(-)	intergenic region	n/a	-
circRNA751	chr12:35934159 35947677(-)	intron	100807714	LOC100807714
circRNA752	chr3:5026677 5051006(+)	intergenic region	n/a	-
circRNA753	chr12:3366413 3368360(+)	exon	100783343	LOC100783343
circRNA754	chr3:2889318 2956200(+)	intergenic region	n/a	-
circRNA755	chr16:2984739 3010585(+)	intergenic region	n/a	-
circRNA756	chr12:37054496 37056330(-)	exon	100804362	LOC100804362
circRNA757	chr7:5616308 5737485(-)	intron	100780862	LOC100780862
circRNA758	chr11:406172 414451(+)	intergenic region	n/a	-
circRNA759	chr6:44787166 44830669(-)	intergenic region	n/a	-
circRNA760	chr17:27301206 27304033(-)	exon	106796783	LOC106796783
circRNA761	chr13:4159199 4165789(-)	intergenic region	n/a	-
circRNA762	chr15:1649919 1683496(+)	intergenic region	n/a	-
circRNA763	chr9:48491035 48491617(+)	exon	100800714	LOC100800714
circRNA764	chr12:2142564 2144715(-)	exon	100306616	LOC100306616
circRNA765	chr17:35870205 36081663(-)	intergenic region	n/a	-
circRNA766	chr7:5568052 5642775(-)	intergenic region	n/a	-
circRNA767	chr15:15637208 15656311(+)	intergenic region	n/a	-
circRNA768	chr16:31136928 31164051(-)	intergenic region	n/a	-
circRNA769	NW_014573403.1:447609 452435(-)	intergenic region	n/a	-
circRNA770	chr12:39171046 39201052(-)	intergenic region	n/a	-
circRNA771	chr8:34225120 34225496(+)	exon	100780532	LOC100780532
circRNA772	chr17:30723779 30726287(-)	exon	100788049	LOC100788049
circRNA773	chr8:11323834 11328246(-)	exon	100803721	LOC100803721
circRNA774	chr4:52380522 52380592(+)	exon	100786150	LOC100786150
circRNA775	chr6:4019007 4020713(-)	exon	100788141	LOC100788141
circRNA776	chr18:43345766 43346768(+)	exon	100792819	LOC100792819



circRNA777	chr8:44020129 44038088(+)	intergenic region	n/a	-
circRNA778	chr15:12789840 12790338(+)	intron	100784626	LOC100784626
circRNA779	chr8:12661447 12667988(-)	exon	100788022	LOC100788022
circRNA780	chr1:65205 66320(-)	exon	102661143	LOC102661143
circRNA781	chr15:10846721 10847648(+)	exon	100780698	LOC100780698
circRNA782	chr1:1092992 1135833(-)	intergenic region	n/a	-
circRNA783	chr8:33250697 33253207(+)	exon	100786931	LOC100786931
circRNA784	chrPltd:5419 5742(-)	exon	3989271	rbcL
circRNA785	chr1:53934829 53936075(+)	exon	100790851	LOC100790851
circRNA786	chr3:29505798 29510259(+)	exon	100500111	LOC100500111
circRNA787	chr4:44234918 44248694(-)	intergenic region	n/a	-
circRNA788	chr7:16581545 16586973(+)	intergenic region	n/a	-
circRNA789	chr4:20611036 20612674(-)	exon	100807902	LOC100807902
circRNA790	chr9:10445089 10448959(+)	exon	100780371	LOC100780371
circRNA791	chr16:31516054 31523831(-)	intergenic region	n/a	-
circRNA792	chr13:18691319 18691999(-)	exon	100777822	LOC100777822
circRNA793	chr12:39175298 39205563(-)	intergenic region	n/a	-
circRNA794	chr3:11451080 11454212(-)	exon	100819282	LOC100819282
circRNA795	chr7:41734490 41736319(-)	exon	100789933	LOC100789933
circRNA796	chr13:28350331 28351147(-)	intron	100819150	LOC100819150
circRNA797	chr13:30627515 30632709(+)	exon	100813299	LOC100813299
circRNA798	chr7:43007302 43007672(-)	exon	100301901	DXS1
circRNA799	chr10:42469222 42471814(+)	exon	100810184	LOC100810184
circRNA800	chr8:15991994 15996000(+)	exon	100776818	LOC100776818
circRNA801	chr18:24821791 24846120(-)	exon	100818895	LOC100818895
circRNA802	chr16:36345772 36387941(+)	intergenic region	n/a	-
circRNA803	chr20:35485340 35486636(-)	exon	100818587	LOC100818587
circRNA804	chrPltd:1049 1428(-)	intergenic region	n/a	-
circRNA805	chr11:15567014 15576696(-)	intergenic region	n/a	-
circRNA806	chr7:5337328 5344129(-)	intergenic region	n/a	-
circRNA807	chr6:47133484 47144602(+)	intergenic region	n/a	-
circRNA808	chr3:23079358 23080745(+)	exon	100780970	LOC100780970
circRNA809	chr16:7174054 7174914(+)	exon	100799113	LOC100799113
circRNA810	chr1:64055 64199(-)	exon	102661143	LOC102661143
circRNA811	chr15:49961001 49961874(+)	exon	100811882	LOC100811882
circRNA812	chr2:11463971 11545580(-)	intergenic region	n/a	-
circRNA813	chr10:46239930 46246023(-)	intergenic region	n/a	-
circRNA814	chr12:26067884 26068699(-)	exon	100796730	LOC100796730
circRNA815	chr18:51534325 51601919(+)	intergenic region	n/a	-
circRNA816	chr15:916064 938793(+)	intergenic region	n/a	-
circRNA817	chr9:24718305 24739833(+)	intergenic region	n/a	-
circRNA818	chr20:34666212 34666927(+)	exon	102669236	LOC102669236
circRNA819	chr8:11667794 11668295(+)	exon	100811720	LOC100811720
circRNA820	chr8:12191899 12192399(-)	intron	100780527	LOC100780527
circRNA821	chr7:15549263 15553458(+)	exon	100803526	LOC100803526
circRNA822	chr3:27908660 27921314(-)	exon	100813936	LOC100813936
circRNA823	chr8:20585673 20586101(+)	exon	100783218	LOC100783218
circRNA824	chr12:39176507 39206477(-)	intergenic region	n/a	-
circRNA825	chr12:4469844 4490071(+)	intergenic region	n/a	-
circRNA826	chr3:44856675 44860262(+)	exon	100814483	LOC100814483
circRNA827	chr1:27707 28264(-)	intergenic region	n/a	-
circRNA828	chr2:45645550 45645620(-)	exon	100783265	LOC100783265
circRNA829	chr13:18691319 18691996(-)	exon	100777822	LOC100777822
circRNA830	chr10:50445129 50445829(-)	exon	100794236	LOC100794236

Table S4: Significant GO enrichment analysis of differently expressed lncRNA targets.

		GO terms	Type	Q-value	lncRNAs Number
Cis-acting manner	Drought	nucleoplasm	cellular_component	5.60E-04	4
		transcription from RNA polymerase II promoter	biological_process	1.31E-03	4
		RNA binding	molecular_function	7.76E-03	6
		RNA processing	biological_process	2.68E-02	4
		structural constituent of ribosome	molecular_function	2.99E-02	4
	Saline-alkaline	transcription factor activity	molecular_function	9.55E-03	4
		signal transduction	biological_process	1.36E-02	4
		phosphate-containing compound metabolic process	biological_process	1.66E-02	4
		transcription, DNA-templated	biological_process	4.05E-03	7
		cellular response to stimulus	biological_process	1.33E-02	5
Trans-acting manner	Drought	ATPase activity	molecular_function	1.47E-03	4
		pyrophosphatase activity	molecular_function	1.29E-03	5
		acting on acid anhydrides	molecular_function	1.35E-03	5
		phosphate-containing compound metabolic process	biological_process	1.49E-03	5
		hydrolase activity, acting on acid anhydrides	molecular_function	5.98E-03	5
	Saline-alkaline	hydrolyzing O-glycosyl compounds	molecular_function	1.85E-04	4
		acting on glycosyl bonds	molecular_function	3.00E-04	4
		carbohydrate metabolic process	biological_process	2.47E-03	5
		cellular protein modification process	biological_process	8.46E-03	4
		regulation of transcription, DNA-templated	biological_process	1.17E-02	4