

EVALUATION OF CHEMICAL COMPOSITION AND *IN VITRO* DRY AND ORGANIC MATTER DIGESTIBILITY OF SOME FORAGE PLANT SPECIES DERIVED FROM EGYPTIAN RANGELANDS

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ABSTRACT

The objective of the study was to evaluate wild forage plant species derived from Egyptian rangelands based on the determination of their chemical composition, *in vitro* digestibility and gross energy value. Twenty four wild palatable forage plants were identified and analyzed for crude fiber, ether extract, crude protein, nitrogen free extract, ash, neutral detergent fiber and acid detergent fiber, acid detergent lignin, cellulose, hemi-cellulose, silica, *in vitro* dry matter digestibility (IVDMD) and *In vitro* organic matter digestibility (IVOMD) and gross energy. The differences among species were significant in the same group of plants and among different groups (grasses, legumes and forbs). Significant differences were observed between grasses in all factors evaluated, except acid detergent lignin. In legumes the differences among species in values of acid detergent fiber, acid detergent lignin and cellulose were not significant. Contrary, all tested factors among forbs species were significantly different. IVDMD and IVOMD showed a significantly higher trend for grasses compared to legumes and forbs. Contrary, forbs showed a higher trend in gross energy compared to grasses and legumes. It can concluded that the high protein, moderate fiber contents and high *in vitro* digestibility found in *Ammophila arenaria*, *Trigonella maritime* and *Vicia monantha*, *Vicia sativa*, *Ononis vaginalis*, *Atriplex nummularia* and *Lycium shawii* make them good quality wild forage plant species that can be used in ruminants' nutrition under Egyptian rangelands conditions.

Key words: Grasses, legumes, forbs, fiber, silica and energy.

INTRODUCTION

The issue of animal feed supply and its quality is aggravated in arid, semi-arid and tropical regions with scarce and erratic rainfall that limits the growth of herbaceous species and biomass yield (Boufennara *et al.*, 2012). Egyptian deserts covered by native palatable plants can provide continuous forage supply for domestic grazing animals and support the national economy. Although the natural plant cover of Egyptian deserts is quite low and scattered, the flora in the North West coast is relatively rich and diverse. The Western Mediterranean Coastal land is one of the richest phyto-geographical regions in Egypt because of its relatively high rainfall. The natural vegetation includes many annuals species, mostly herbs and a few grasses, perennial herbs, shrubs, sub-shrubs, and a few trees. These species represent 50% of the total flora of Egypt (UNESCO, 2003). Heneidy (2004) found that the most representative plant families in North West Coastal region of Egypt were *Asteraceae*, *Fabaceae*, *Poaceae* and *Chenopodiaceae*. Other families were represented by less than 2 % of total, *e.g.*, *Aizoaceae*, *Apiaceae*, *Brassicaceae*, *Caryophyllaceae*, *Liliaceae*, *Plantaginaceae*, *Ranunculaceae*, *Solanaceae*

and *Thymelaceae*. Täckholm (1974) and Boulos (1995) reported that *Fabaceae* and *Asteraceae* are the largest families in Egypt with the greatest number of plant species. Indigenous shrub and bush species have also a potential to (i) prevent desertification, (ii) mitigate the effects of droughts, (iii) allow soil fixation, (iv) enhance the restoration of the vegetation and the recuperation of rangelands. Hence, there is an increasing interest in the rational utilization of potential livestock feed resources such as browse species that are adapted to these environments (Robles *et al.*, 2008). Obviously, the plant species selected for the present study grow throughout the Western Mediterranean Coastal of Egypt and are commonly used as range plants and feed supplements for grazing animals in many regions. In spite of their limited nutritional value, these forage resources are indispensable as feeds for herbivores in such kind of when production systems which are based on grazing rangelands (Papanastasis *et al.*, 2008). Also, chemical composition of pasture vegetation is crucial, particularly in combination with *in vitro* digestibility, to evaluate the nutritive value of browse species that are not known previously (Laudadio *et al.*, 2009). Consequently, a detailed analysis of browse species is important to

74 identify the most proper shrub species for ruminants, in
75 terms of nutrient content and digestibility (Tufarelli *et al.*
76 2010). Boufennara *et al.* (2012) reported that all the
77 chemical, *in vitro* and *in situ* measurements are useful
78 tools in initial screening studies to rank the forage
79 according to their nutritive quality. Keeping in view these
80 facts, this study designed to evaluate the chemical
81 composition, *in vitro* digestibility and gross energy value
82 of some wild forage plant species derived from Egyptian
83 rangelands.

84 MATERIALS AND METHODS

85 The experiments were conducted at the
86 Experimental Laboratories of the Natural Resources
87 Department, Institute of African Research and Studies,
88 Cairo University, Animal Production Department,
89 Faculty of Agriculture, Cairo University, Egypt and
90 Department of Animal Nutrition and Feed Management,
91 Poznan University of Life Sciences, Poznan, Poland
92 during period 2015-2017.

93 **Plant materials:** Ten samples of twenty-four wild
94 palatable forage plants were collected from different sites
95 of Egyptian Rangelands in Mediterranean coast region
96 (Marsa Matrouh area) and shoot parts of annual plants
97 were clipped and new sprouts of perennials were cut
98 during spring seasons (Table 1). Plant species were firstly
99 identified at the field, collected and prepared as
100 herbarium sheets after being pressed. All collected and
101 preserved plants were taken to the Cairo University
102 herbarium for accurate identification. Floristic
103 identifications were performed according to Täckholm
104 (1974) and the scientific names of species were updated
105 by Boulos (1999, 2000, 2002 and 2005).

106 **Chemical composition of feeds:** Samples of feeds were
107 analyzed according to A.O.A.C. (2007, Table 1) for dry
108 matter (method no. 934.01) and ash (method no. 942.05).
109 Crude protein was determined by Kjehl-Foss Automatic
110 16210 analyzer (method no. 976.05), crude fat by Soxtec
111 System HT analyzer (method no. 2003.05), and crude
112 fiber by Tecator Fibertec System I (method no. 978.10).
113 Nitrogen-free-extractives (NFE %) was calculated using
114 the difference method by subtracting the sum of crude
115 protein, crude fiber, ether extract and total ash from 100.
116
$$\text{NFE \%} = 100 - (\text{CP \%} + \text{CF \%} + \text{EE \%} + \text{TA \%})$$

117 **Batch culture fermentation:** The rumen inoculum was
118 obtained 3 h after the morning feeding from three rumen
119 cannulated Polish Holstein-Friesian dairy cows (mean
120 body weight 600 ± 23 kg) fed the diet (kg/day) containing
121 alfalfa silage, 46.0; meadow hay, 1.80; corn meal, 0.90;
122 dry brewer's grains, 0.60; protein concentrate (35% crude
123 protein), 1.50; wheat bran, 0.60; and commercial
124 concentrate (19% crude protein), 5.50. Ruminal content

was squeezed through four layers of cheesecloth into a
Schott Duran® bottle (SCHOTT North America, Inc.
Corporate Office, Elmsford, NY 10523, USA) with an O₂
- free headspace and immediately transported to the
laboratory in a water bath preheated to $39 \pm 0.5^\circ\text{C}$. Each
substrate (wild palatable forage plants) was grounded
through a 0.15–0.4 mm screen, bulked and stored in
sealed plastic containers. The ground substrates (400 ± 1
mg) were weighted into each individual batch culture
fermentation bottle (100 mL). The batch culture
fermentation was carried out according to Szumacher-
Strabel *et al.* (2004). Briefly, rumen fluid was diluted
with a buffer (mg/L; K₂HPO₄ 292, KH₂PO₄ 240,
(NH₄)₂SO₄ 480, NaCl 480, MgSO₄·7H₂O 100,
CaCl₂·2H₂O 64, Na₂CO₃ 4, and cysteine HCl 600) in ratio
2:3. Then aliquots of 40 mL were transferred into
incubation bottles. The bottles were filled with CO₂ and
then closed with a rubber stopper and aluminum-sealed.

***In vitro* dry and organic matter digestibility:** After
incubation, the bottles content was transferred to the
previously weighed crucible. The residues of incubation
was washed with 50 ml distilled water and dried at 105°C
for 3 days. *In vitro* dry matter digestibility (IVDMD) was
calculated after 24-h incubation using the following
equation:

$$\text{IVDMD (\%)} = [(\text{initial DM input} - (\text{Residue} - \text{Blank})) / \text{initial DM input}] \times 100$$

Then the samples of palatable forage plants,
residue and blank samples were ashed at 450 °C and
weighed. IVOMD (*In vitro* organic matter digestibility)
was calculated according to the following formulas:
$$\text{IVOMD (\%)} = [(\text{initial OM input} - (\text{Residue} - \text{Blank})) / \text{initial OM input}] \times 100$$

Energy calculation: Gross Energy (GE) of each
palatable forage plant was estimated according to DLG
(1982) equation:

$$\text{GE (MJ/kg DM)} = 0.0242\text{CP} + 0.0366\text{EE} + 0.0209\text{CF} + 0.0170\text{NFE (g)}$$

Statistical analysis: The layout of the experiment was
arranged in a randomized complete blocks design, with 3
blocks (replicates), and the resulted data were subjected
to statistical analysis, employing F-test for significance at
 $P = 0.05$ and computing of LSD values to separate means
in different statistical groups according to the described
method by Little and Hills (1978).

RESULTS

Chemical composition of feeds: Crude fiber (CF %) content in grasses was higher than legumes and forbs (Table 2). Among grasses, CF increased from 19.17% (*Ammophila arenaria*), 26.03% (*Lophochloa cristata*), to 38.30% (*Lygeum spartum*). Among legumes, the CF ranged from 15.30% in *Astragalus homosusto* to 30.10% in *Lygos raetam*. Among forbs CF increased from

178 11.69% (*Atriplex nummularia*), 13.61% (*Tamarix*
 179 *nilotica*), to 28.74% (*Gymnocarpos decandrum*). 234
 180 The ether extract (EE %) contents of grasse235
 181 varied between 1.74% (*Hordeum marinum*) and 5.62%236
 182 (*Phalaris minor*). In contrast, among legumes, ethe237
 183 extract contents varied between 1.18% (*Vicia sativa*) and238
 184 4.49% (*Trigonella maritime*). While, among forbs ethe239
 185 extract content of analyzed plants varied between 1.17%240
 186 (*Deverra trotuosa*) and 3.95% (*Tamarix nilotica*). 241
 187 The highest percentages of crude protein (CP)242
 188 attained in *Vicia sativa*, *Ononis vaginalis* and *Vicia*
 189 *monantha* were 16.70, 13.98 and 12.72% respectively244
 190 among legumes (Table 2). In grasses, the CP content245
 191 attained in *Ammophila arenaria* (14.00 %) and246
 192 *Phragmites australis* (13.09 %). While, among forbs the247
 193 highest percentages of CP attained in *Gymnocarpo*
 194 *decandrum* (18.47%) and *Tamarix nilotica* (13.93%). 249
 195 The values of nitrogen free extract (NFE %) i250
 196 forbs increased from 49.51% (*Tamarix nilotica*), 56.54%251
 197 (*Atriplex halimus*) to 63.10% (*Atriplex nummularia*) and252
 198 ranged from 50.78% in *Astragalus homosus* to 51.59% i253
 199 *Vicia monantha* as legumes. Among grasses, NFE254
 200 increased from 43.33% (*Hordeum marinum*), 44.11%255
 201 (*Phalaris minor*) to 44.75% (*Aegilops kotschy*) as show256
 202 in Table 2. 257
 203 Total ash (TA %) among legumes increase258
 204 from 9.70% (*Lygos raetam*), 12.21% (*Vicia sativa*) to
 205 22.05% (*Trigonella maritime*) and 23.24% (*Ononis*
 206 *vaginalis*). In grasses, the TA ranged from 11.48% in
 207 *Phragmites australis* to 21.22% in *Lophochloa cristata*,
 208 Forbs total ash ranged from 12.01% in (*Deverra*
 209 *trotuosa*), to 18.99% (*Tamarix nilotica*). 264
 210 The neutral detergent fiber (NDF), acid
 211 detergent fiber (ADF), acid detergent lignin (ADL)
 212 cellulose, hemi-cellulose and silica contents of some wild
 213 plants are summarized in Table (3). Among grasses, the
 214 highest and lowest values of NDF contents were 66.70
 215 g/kg (*Lygeum spartum*) and 29.56 g/kg (*Ammophila*
 216 *arenaria*). In legumes, the highest and lowest values of
 217 NDF contents were 50.61 g/kg and 33.67 g/kg in
 218 *Trigonella maritime* and *Astragalus homosus*,
 219 respectively. In forbs, the highest and lowest values of
 220 NDF contents were 63.95 g/kg (*Deverrat rotuosa*) and
 221 22.34 g/kg (*Atriplex nummularia*). 276
 222 Acid detergent fiber contents of grasses varied
 223 from 41.23 g/kg (*Lygeum spartum*) to 25.47 g/kg
 224 (*Ammophila arenaria*). In contrast, among legumes, ADF
 225 varied between 15.29 g/kg for *Astragalus homosus* and
 226 36.77 g/kg for *Lygos raetam*. While, among forbs ADF
 227 content of analyzed plants varied between 16.77 g/kg for
 228 *Atriplex nummularia* and 45.80 g/kg for *Deverra*
 229 *trotuosa*. 284
 230 Among the tested grasses, legumes and forbs
 231 the highest ADL contents (17.24, 16.98 and 14.31 g/kg)
 232 were recorded in *Lolium rigidum*, *Hordeum marinum* and

Deverra trotuosa, respectively. In contrast, the lowest
 values of ADL contents (2.67, 2.67 and 3.08 g/kg) were
 recorded in *Phragmites australis*, *Lophochloa cristata*
 and *Atriplex halimus*, respectively.

Commonly, grasses had more cellulose than
 legumes and forbs. Within grasses, cellulose content of
 tested plants varied between 37.03 g/kg (*Lygeum*
spartum) and 9.39 g/kg (*Lolium rigidum*). In legumes,
 cellulose content varied between 4.39 g/kg (*Astragalus*
homosus) and 27.77 g/kg (*Lygos raetam*) whilst among
 forbs cellulose contents of tested plants varied between
 2.75 g/kg for *Atriplex nummularia* and 34.75 g/kg for
Gymnocarpos decandrum.

Generally, grasses had more hemi-cellulose than
 legumes and forbs (Table 3). The highest values of hemi-
 cellulose (35.26, 31.50 and 30.88 g/kg) found in *Lolium*
rigidum, *Aeluropus lagopoides* and *Phragmites australis*,
 respectively whilst the lowest values of hemi-cellulose
 (4.10, 4.49 and 4.69 g/kg) were found in *Ammophila*
arenaria, *Gymnocarpos decandrum* and *Tamarix*
nilotica, respectively. In silica contents varied from 7.42
 g/kg in *Aeluropus lagopoides* to 10.04 g/kg in *Lolium*
rigidum. Among legumes, it ranged from 13.37 g/kg in
Trigonella maritime to 2.95 g/kg in *Lotus polyphyllus*.
 Among forbs, silica content ranged from 0.21 g/kg in
Atriplex nummularia to 5.29 g/kg in *Deverra trotuosa*.

In vitro digestibility and gross energy of feeds: Overall
 results showed significant differences among different
 groups (Table 4). IVDMD and IVOMD showed a higher
 trend for the group of grasses compared to legumes and
 forbs. Contrary, forbs showed a higher trend in gross
 energy compared to grasses and legumes.

The highest values of IVDMD were 44.39,
 42.91, 40.48 and 40.37% in *Ammophila arenaria*,
Hordeum marinum, *Trigonella maritime* and *Vicia*
monantha, respectively. In contrast, the lowest values of
 IVDMD were 22.68, 23.51, 27.47 and 27.77% in *Deverra*
trotuosa, *Gymnocarpos decandrum*, *Lygo sraetam* and
Panicum coloratum, respectively.

IVOMD percentages of analyzed plants varied
 between 37.19% for *Ammophila arenaria* and 61.24% for
Lygeum spartum among grasses. Among legumes
 IVOMD varied between 34.07% (*Trigonella maritime*)
 and 60.57% (*Lygos raetam*) while, among forbs IVOMD
 of tested plants varied between 64.11% (*Deverra*
trotuosa) and 33.66% (*Atriplex halimus*).

Gross energy (Table 4) among forbs fluctuated
 between 1.76 MJ/kg DM (*Deverra trotuosa*) to 1.61
 MJ/kg DM (*Tamarix nilotica*). Among grasses, it varied
 from 1.78 MJ/kg DM (*Phragmites australis*) to 1.59
 MJ/kg DM (*Lophochloa cristata*). Likewise, among
 legumes, GE of tested legumes was ranged from 1.56
 MJ/kg DM in *Astragalus homosus* to 1.74 in MJ/kg DM in
Vicia sativa.

288 **Table 1. Botanical composition and life duration of tested forage plants.**

Family Name	Scientific Name	Life Duration
	<i>Aegilops kotschy</i>	Annual
	<i>Aeluropus lagopoides</i>	Perennial
	<i>Ammophila arenaria</i>	Perennial
	<i>Hordeum marinum</i>	Annual
Poaceae	<i>Lolium rigidum</i>	Annual
(Gramineae)	<i>Lophochloa cristata</i>	Annual
	<i>Lygeum spartum</i>	Perennial
	<i>Panicum coloratum</i>	Perennial
	<i>Phalaris minor</i>	Annual
	<i>Phragmites australis</i>	Perennial
	<i>Astraglus homosus</i>	Annual
	<i>Lotus polyphyllus</i>	Perennial
	<i>Lygos raetam</i>	Perennial
Fabaceae	<i>Ononis vaginalis</i>	Perennial
(Leguminosae)	<i>Trifolium tomentosum</i>	Annual
	<i>Trigonella maritima</i>	Annual
	<i>Vicia monantha</i>	Annual
	<i>Vicia sativa</i>	Annual
Asteraceae	<i>Anacyclus alexandrinus</i>	Perennial
Chenopodiaceae	<i>Atriplex halimus</i>	Perennial
Chenopodiaceae	<i>Atriplex nummularia</i>	Perennial
Apiaceae	<i>Deverra trotuosa</i>	Perennial
Caryophyllaceae	<i>Gymnocarpos decandrum</i>	Perennial
Solanaceae	<i>Lycium shawii</i>	Perennial
Tamaricaceae	<i>Tamarix nilotica</i>	Perennial

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291 **Table 2. Chemical composition of tested feeds (on DM basis).**

Plant Species	Chemical Composition %					
	CF	EE	CP	NFE	Ash	OM
Grasses						
<i>Aegilops kotschy</i>	29.97	2.50	8.95	44.75	13.83	86.17
<i>Aeluropus lagopoides</i>	28.42	3.53	9.97	40.34	17.74	82.26
<i>Ammophila arenaria</i>	19.17	5.22	14.00	41.69	19.92	80.08
<i>Hordeum marinum</i>	29.65	1.74	7.27	43.33	18.01	81.99
<i>Lolium rigidum</i>	30.25	2.39	10.82	39.96	16.58	83.42
<i>Lophochloa cristata</i>	26.03	3.20	12.63	36.92	21.22	78.78
<i>Lygeum spartum</i>	38.30	1.92	7.64	41.22	10.92	89.08
<i>Panicum coloratum</i>	34.80	3.24	9.05	37.88	15.03	84.97
<i>Phalaris minor</i>	29.07	5.62	5.89	44.11	15.32	84.68
<i>Phragmites australis</i>	32.82	2.57	13.09	40.05	11.48	88.52
Mean	29.85	3.19	9.93	41.03	16.01	84.00
LSD _{0.05}	1.95	2.13	0.37	3.35	1.42	1.42
Legumes						
<i>Astraglus homosus</i>	15.30	2.52	11.84	50.78	19.57	80.43
<i>Lotus polyphyllus</i>	26.56	2.76	10.79	42.95	16.95	83.05
<i>Lygo sraetam</i>	30.10	2.75	11.79	45.66	9.70	90.30
<i>Ononis vaginalis</i>	22.47	4.03	13.98	36.27	23.24	76.76
<i>Trigonella maritima</i>	26.72	4.49	11.25	35.49	22.05	77.95
<i>Vicia monantha</i>	20.34	2.26	12.72	51.59	13.09	86.91
<i>Vicia sativa</i>	25.72	1.18	16.70	44.20	12.21	87.79
Mean	23.89	2.86	12.72	43.85	16.69	83.31
LSD _{0.05}	0.28	2.63	0.11	2.88	0.73	0.73

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Forbs						
<i>Anacyclus alexandrinus</i>	26.61	2.79	8.99	45.19	16.41	83.59
<i>Atriplex halimus</i>	13.97	2.48	13.03	56.54	13.97	86.03
<i>Atriplex nummularia</i>	11.69	3.29	9.00	63.10	12.92	87.08
<i>Deverra trotuosa</i>	35.02	1.17	13.90	37.91	12.01	87.99
<i>Gymnocarpos decandrum</i>	28.74	3.50	18.47	33.34	15.94	84.06
<i>Lycium shawii</i>	26.21	3.85	15.34	39.36	15.23	84.77
<i>Tamarix nilotica</i>	13.61	3.95	13.93	49.51	18.99	81.01
Mean	22.26	3.00	13.24	46.42	15.07	84.93
LSD _{0.05}	1.47	1.23	0.11	4.58	4.21	4.21
Collective-Mean	25.90	3.04	11.71	43.42	15.93	84.07
Collective-LSD _{0.05}	1.46	1.89	0.27	3.77	2.66	2.66

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Table 3. Fiber fractions and silica content of tested feeds (on DM basis).

Plant Species	Fiber Fractions (g/kg)					Silica (g/kg)
	NDF	ADF	ADL	Cellulose	Hemi-cellulose	
Grasses						
<i>Aegilops kotschy</i>	59.81	31.70	4.14	27.57	28.11	4.44
<i>Aeluropus lagopoides</i>	64.64	33.14	4.72	28.42	31.50	7.42
<i>Ammophila arenaria</i>	29.56	25.47	5.19	20.28	4.10	3.44
<i>Hordeum marinum</i>	61.13	32.64	16.98	15.66	28.48	7.23
<i>Lolium rigidum</i>	61.89	26.63	17.24	9.39	35.26	10.04
<i>Lophochloa cristata</i>	52.70	30.22	2.67	27.56	22.48	4.61
<i>Lygeum spartum</i>	66.70	41.23	4.20	37.03	25.47	3.69
<i>Panicum coloratum</i>	64.16	40.70	4.61	36.09	23.46	2.70
<i>Phalaris minor</i>	53.07	32.13	5.07	27.06	20.94	1.82
<i>Phragmites australis</i>	65.95	35.07	2.67	32.40	30.88	4.29
Mean	57.96	32.89	6.75	26.15	25.07	4.97
LSD _{0.05}	1.54	0.63	NS	0.63	1.79	1.45
Legumes						
<i>Astragalus homosus</i>	33.67	15.29	10.90	4.39	18.38	9.01
<i>Lotus polyphyllus</i>	40.55	34.12	9.45	24.67	6.43	2.95
<i>Lygosraetam</i>	50.52	36.77	9.00	27.77	13.75	4.07
<i>Ononisvaginalis</i>	35.90	25.43	3.65	21.78	10.47	3.87
<i>Trigonella maritima</i>	50.61	26.83	8.83	18.00	23.78	13.73
<i>Vicia monantha</i>	40.16	25.22	4.90	20.33	14.94	4.22
<i>Vicia sativa</i>	42.66	28.64	6.45	22.18	14.02	3.89
Mean	42.01	27.47	7.60	19.87	14.54	5.96
LSD _{0.05}	2.91	NS	NS	NS	2.91	0.64
Forbs						
<i>Anacyclus alexandrinus</i>	42.22	34.52	7.62	26.90	7.70	2.90
<i>Atriplex halimus</i>	31.20	22.03	3.08	18.96	9.17	2.48
<i>Atriplex nummularia</i>	22.34	16.77	14.02	2.75	5.57	0.21
<i>Deverra trotuosa</i>	63.95	45.80	14.31	31.49	18.15	5.29
<i>Gymnocarpos decandrum</i>	49.00	44.52	9.77	34.75	4.49	1.40
<i>Lycium shawii</i>	31.43	23.67	6.85	16.82	7.76	0.78
<i>Tamarix nilotica</i>	23.32	18.63	6.37	12.25	4.69	0.79
Mean	37.64	29.42	8.86	20.56	8.22	1.98
LSD _{0.05}	3.72	0.61	0.40	0.67	3.65	1.02
Collective-Mean	47.38	30.30	7.61	22.69	17.08	4.39
Collective-LSD _{0.05}	1.20	0.62	0.20	0.63	2.50	1.05

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298 **Table 4. *In vitro* dry, organic matters digestibility and gross energy of tested feeds.**
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Plant Species	<i>In vitro</i> Digestibility %		Gross Energy (MJ/kg DM)
	DMD	OMD	
Grasses			
<i>Aegilops kotschyi</i>	36.72	55.04	1.70
<i>Aeluropus lagopoides</i>	30.75	53.96	1.65
<i>Ammophila arenaria</i>	44.39	37.19	1.64
<i>Hordeum marinum</i>	42.91	46.00	1.60
<i>Lolium rigidum</i>	35.58	50.65	1.66
<i>Lophochloa cristata</i>	35.88	47.48	1.59
<i>Lygeum spartum</i>	28.40	61.24	1.76
<i>Panicum coloratum</i>	27.77	60.08	1.71
<i>Phalaris minor</i>	37.02	50.09	1.71
<i>Phragmites australis</i>	34.85	52.85	1.78
Mean	35.43	51.46	1.68
LSD _{0.05}	10.82	11.97	0.04
Legumes			
<i>Astragalus homosus</i>	34.23	35.97	1.56
<i>Lotus polyphyllus</i>	30.32	51.60	1.65
<i>Lygos raetam</i>	27.47	60.57	1.79
<i>Ononis vaginalis</i>	39.12	37.95	1.57
<i>Trigonella maritima</i>	40.48	34.07	1.60
<i>Vicia monantha</i>	40.37	45.42	1.69
<i>Vicia sativa</i>	35.40	50.52	1.74
Mean	35.34	45.16	1.66
LSD _{0.05}	10.31	10.40	0.05
Forbs			
<i>Anacyclus alexandrinus</i>	30.09	44.90	1.64
<i>Atriplex halimus</i>	34.08	33.66	1.66
<i>Atriplex nummularia</i>	31.80	37.96	1.66
<i>Deverra trotuosa</i>	22.68	64.11	1.76
<i>Gymnocarpos decandrum</i>	23.51	61.64	1.74
<i>Lycium shawii</i>	34.94	44.24	1.73
<i>Tamarix nilotica</i>	33.65	40.43	1.61
Mean	30.11	46.71	1.69
LSD _{0.05}	10.72	9.74	0.07
Collective-Mean	33.85	48.23	1.68
Collective-LSD _{0.05}	10.42	11.18	0.05

300
301 **DISCUSSION**
302 **Chemical composition:** The chemical composition is the
303 first step to evaluate the nutritive value for any tested
304 feed. The present investigation primarily indicated the
305 potential to use the tested plants as a source of nitrogen
306 and carbohydrate in ruminant diets. In our study, crude
307 fiber and ether extract contents increased in the species
308 that contained less protein and decreased in the species
309 with high protein content. Similar findings reported in
310 earlier studies (Abbas *et al.* 2008; El-Morsy, 2002;
311 Ibrahim, 1995). It is reported that due to the increase in
312 temperature, the highest percentage of crude fiber was
313 recorded during summer followed by spring, autumn and
314 winter in descending order (Ibrahim, 1995). Hussain and
315 Durrani (2009) also reported that mature plants usually
316 contained higher CF content than in young plants.
317 Protein is an important nutrient supplied by
318 forages and plant species belonging to *Fabaceae* family
319 that generally produce higher forage quality than other
320 species because legumes usually have high crude protein
321 and favor higher intake than others. Our results agreed
322 with those of Hussain and Durrani (2009) who showed
323 that shrubs generally contained slightly higher CP than
324 grasses. However, Ammar *et al.* (2004) observed that
325 leguminous forages and trees have been used as a basic
326 feed animal supply in many regions of the world mainly
327 because of their high protein contents throughout the
328 year. Holechek *et al.* (1995) found that leaves of forbs
329 and shrubs are generally higher in protein than leaves of
330 grass. Reiad *et al.* (1996) indicated that it is more likely

331 that perennial plants are often relatively lower in CP. **O**385
 332 **the other hand**, El-Shesheny (2007) reported that ther**386**
 333 was insignificant effect of plant growth seasons on crud**387**
 334 protein and also nitrogen-free extract consisting o**388**
 335 carbohydrates, sugars, starches at the North West coast**389**
 336 region of Egypt. In this respect, Holechek *et al.* (1995**390**
 337 indicated that NFE is one of the most importan**391**
 338 components of forage because it represents the basi**392**
 339 source of energy for grazing animals. Baumi (2003**393**
 340 reported that the increase or decrease of any of the **394**
 341 mentioned components (carbohydrate or crude protein **395**
 342 contents) affect the quality of the foliage of the plant **396**
 343 species. Literature explains that there is negativ**397**
 344 relationship between total ash and organic matter conten**398**
 345 Hussain and Durrani (2009) reported that the increase o**399**
 346 decrease of ash content with advancing age by differen**400**
 347 plants might be due to variation in soil and other habit**401**
 348 features that need to be explored. However, El-Kerdawy **402**
 349 (1992) in the North West coastal region of Egypt claim**403**
 350 that mineral concentrations of range plants on dry matter **404**
 351 basis were greatly affected by plant species and stage o**405**
 352 growth but not by plant location except for phosphorus **406**
 353 and sodium in case of perennials. Therefore, the high**407**
 354 percentages of ash mean the lowest percentages of OM **408**
 355 Generally, legumes had more ash percentages or les**409**
 356 organic matter percentages than grasses and forbs. **410**

357 **Fiber fractions:** Results of our study stated that legum**411**
 358 slightly varied ($P>0.05$) among species in values of aci**412**
 359 detergent fiber, acid detergent lignin and cellulose**413**
 360 Contrary, all test factors among forbs species wer**414**
 361 significantly different. The increased fiber content ma**415**
 362 be due to plant species, maturity and environmen**416**
 363 conditions. Similar results had been reported b**417**
 364 Boufennara *et al.* (2012) who found the high level o**418**
 365 fiber content (NDF, ADF and lignin) in some of the**419**
 366 forage species could be explained partly by the ecologica**420**
 367 conditions-high temperature and low precipitations. Thi**421**
 368 tended to increase the cell wall fraction and to decreas**422**
 369 the soluble contents of the plants. On the other hand**423**
 370 Sultan *et al.*, (2008) indicated that the structur**424**
 371 constituents (NDF, ADF, hemi-cellulose and lignin**425**
 372 increased in grasses from early bloom to maturity stag**426**
 373 These results are in agreement with those obtained b**427**
 374 Hussain and Durrani (2009) who indicated that NDF**428**
 375 concentration of grasses was higher than of shrub**429**
 376 However, Kramberger and Klemencic (2003) reporte**430**
 377 increase in NDF concentration with maturity of plants**431**
 378 Inline to our findings, Hussain and Durrani (2009**432**
 379 observed that ADF contents of grasses were significan**433**
 380 higher than of shrubs at pre-reproductive and post**434**
 381 reproductive stages. Boufennara *et al.* (2012) reporte**435**
 382 that monocots had higher NDF and ADF and lower ligni**436**
 383 contents than dicots. Cherney *et al.* (1993) reporte**437**
 384 increase in all fiber constituents with increasing maturity **438**

Furthermore, Brown *et al.* (1984) reported that the soil fertility could also influence grass lignin concentration.

The values of hemicelluloses recorded in our study agreed with the findings of Ghadaki *et al.* (1975) who indicated the cellulose content of grasses was higher than that of shrubs, fortes, and especially legumes. Similar results had been reported by Hussain and Durrani (2009) who found out those grasses generally had greater hemicellulose contents than shrubs.

***In vitro* digestibility and gross energy:** *In vitro* digestibility considered as the second step of feed evaluation. Variations in the results of IVDMD can be attributed to several factors, such as processing of samples, difference in chemical composition of feed, preparation of buffer solution or handling of equipment (Tufarelli *et al.*, 2010). Furthermore, The CP, fiber contents, DM degradability and IVOMD values are used as indicator to use as feed supplements for ruminant (Andualem *et al.*, 2016).

IVDMD showed a wide range of qualities in the 24 plant materials tested related to their different chemical compositions (Table 4). In this respect, Tufarelli *et al.* (2010) reported that IVDMD and IVOMD showed a higher trend for the forbs group compared to legumes and grasses. In addition, Foguekem *et al.* (2011) suggested that IVDMD is strongly influenced by the amount of fiber represented by the NDF, ADF and cellulose levels in the plant tissues. Furthermore, Badrzadeh *et al.* (2008) reported that crude protein, digestible dry matter in *Vicia narbonensis* L. var. *affinis* were higher than in the other species of vetches; this is due to low amount of acid and neutral detergent fiber. The results of our study were in line with those of Cherney *et al.* (1990) who observed a negative correlation of IVDMD with NDF, ADF and lignin. Sultan *et al.* (2008) and Van Soest (1978) reported poor relationship of NDF and digestibility. On the other hand, Foguekem *et al.* (2011) found that changes in the IVDMD are positively correlated with those of crude protein levels. Also, the feeds (indigenous fodder trees and shrubs in Ethiopia) were shown to decrease their crude protein and IVOMD from the wet to dry season while their cell wall constituents were shown to increase (Girma *et al.* 2015).

Generally, forbs contained slightly higher gross energy than grasses and legumes. These results are in agreement with those obtained by Hussain and Durrani (2009) who observed that shrubs contained slightly higher gross energy than the grasses. Contrary, Cook and Harris (1968) observed that grasses had higher gross energy than shrubs. Khachatur (2006) indicated that the wild growing legume forages were characterized by high nutritive and energy values.

Conclusions: The high crude protein and moderate fiber content, along with high IVDMD found in *Ammophila*

440 *arenaria*, *Trigonella maritima* and *Vicia monantha*, *Vicia* 492
 441 *sativa*, *Atriplex nummularia*, *Ononis vaginalis* and 493
 442 *Lycium shawii* suggested that these plants have a greater 494
 443 nutritive value than the highly fibrous and of moderate 495
 444 IVDMD species such as: *Lygeum spartum*, *Lygos raetana* 496
 445 *Panicum coloratum*, *Lolium rigidum*, *Gymnocarpus* 497
 446 *decandrum*, *Deverra trotuosa* and *Aeluropus lagopoides* 498
 447 Because of the relatively high fiber content the plant 499
 448 species of the greater nutritive value can be used as 500
 449 components of ruminants' diets. However further animal 501
 450 experiment is needed to validate this conclusion. 502

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