

FOLIAGE APPLIED GIBBERELIC ACID INFLUENCES GROWTH, NUTRIENT CONTENTS AND QUALITY OF LETTUCES (*LACTUCA SATIVA* L.)

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ABSTRACT

Plant growth regulators (PGRs) have been considered as agrochemicals applied widely in agricultural production, wherein, gibberelic acid (GA₃) has gained an interest in using to enhance the growth and improve yield for various vegetables. In this work, lettuces were sprayed with GA₃ at different concentration of 0, 10, 30, 50 and 100 mg L⁻¹, respectively, at three dates after transplanting (the 7-day, the 14-day, and the 21-day). The impacts of GA₃ on the growth, nutrient elements, and the absorption of heavy metals in lettuces were investigated. The accumulation of GA₃ by lettuce plants were confirmed by liquid chromatography - tandem mass spectrometer system (LC-ESI-MS/MS). The results demonstrated, that low concentrations of GA₃ (10 and 30 mg L⁻¹) increased the plant height, number of leaves and fresh weight of lettuces, whereas the opposite results were reported in high concentration of GA₃ treatment (100 mg L⁻¹). In addition, GA₃ decreased nutrient contents of Fe and Mn, and heavy metals' absorption in lettuces. However, the higher concentrations of GA₃ treatment use, the more their accumulations were found in lettuce samples. To the best of our knowledge, this is the first investigation of the influences of GA₃ on nutrient elements and the absorption of heavy metals in lettuce. This is needed to further investigate the improvement of crop production efficiency with PGRs, and to minimize their harmful impact on human health and the ecological environment.

Keywords: GA₃, heavy metals, lettuce, nutrients

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INTRODUCTION

Lettuce (*Lactuca sativa* L.) is an economic value vegetable, which contains high content of vitamin A and several crucial minerals for the health of human namely calcium (Ca), iron (Fe), phosphorus (P), zinc (Zn), magnesium (Mg), manganese (Mn), and potassium (K) (Kim *et al.*, 2016). It is widely used in salad mixtures with other vegetables like carrot, cucumber, and tomato... or in sandwiches. Various properties of lettuces were reported to improve health benefits in the previous studies. Fernanze *et al.* (1997) and USDA (2004) indicated that β-carotene and lutein in lettuces contributing to reduce colorectal cancer, cataracts, heart disease, and stroke in human; whereas phenolic compounds and carotenoids were responsible for antioxidant scavenging and possess antioxidant capacity properties, respectively (López *et al.* 2014). In addition, α-linolenic and omega-3 polyunsaturated fatty acids were found as the major fatty acid in lettuces (Pereira *et al.*, 2001; Le *et al.*, 2008).

Among the growth of population in the world, there is an increase in food consumption, especially vegetables, which might reduce the risks of cancer and degenerative diseases (Lynch *et al.*, 2009; Soerjomataram *et al.*, 2010; Khan *et al.*, 2017). To meet consumers'

increasing demands for green vegetables, it is critical to raise the productivity in vegetables with easy to use, cost-effective, and environmentally friendly techniques (Alessandro *et al.*, 2019). There are many ways to reach these goals, for instance, genetic improvement, grafting, novel production systems, PGRs, and development promoting microorganism (Shah *et al.*, 2006; Maggio *et al.*, 2010; Khan and Samiullah, 2003).

PGRs have been considered as a new generation of agrochemicals applied in agricultural production and in horticulture to control the development processes of plant crops from germination to post-harvest preservation (Sunil *et al.*, 2015). Among these, gibberellic acid (GA₃) has gained an interest in using to enhance the growth and improve yield for various vegetables, such as bean (*Phaseolus vulgaris*), rapeseed-mustard, black cumin (*Nigella sativa* L.), tomato, etc. (Emongor, 2002; Khan and Samiullah, 2003; Shah *et al.*, 2006; Khan *et al.*, 2006). In addition, GA₃ has been also applied to increase the tolerance of vegetables to abiotic stresses, including drought, salinity, salt, etc. (Shah, 2007; Maggio *et al.*, 2010). Besides the effectiveness of GA₃ application in vegetable production, their influence on the nutrient contents, quality and safety of vegetable products to consumers need to be further studied.

In the present research, the influences of gibberellic acid on the growth, nutrient contents and quality of Lettuces (*Lactuca sativa* L.) were investigated. In addition, the absorption of heavy metals associated to GA₃ application was also studied, which might be the basic in exploring solutions for treatments of heavy metals in plant tissues.

MATERIALS AND METHODS

Chemicals: GA₃ was bought from East Asia Agricultural Limited company (Ho Chi Minh City, Vietnam) while certified standard of GA₃ ($\geq 99\%$) and nutrient elements were bought from OlChemim Ltd. (Czech Republic). HPLC grade Methanol (MeOH; 99.80%) and formic acid (FA; $\geq 98\%$) were obtained from Sigma–Aldrich (Singapore). Deionized (DI) water was produced by using a Millipore Milli-q Integral 3 Water Purification System (France).

Instrumentation: The inductive coupled plasma-mass spectrometry (Agilent 7900 ICP-MS, Santa) was used to analyze the contents of heavy metals and nutrients in soil and plant samples. The optimized operating conditions of ICP-MS are Nebulizer gas flow rates: 0.88 L/min; Auxiliary gas flow: 1.2 L/min; Plasma gas flow: 16 L/min; ICP RF power: 1200 W; Lens voltage: 7.25 V; Analyzer vacuum: 5.0×10^{-7} . In addition, the instrument was located in a temperature-control laboratory ($20 \pm 0.5^\circ\text{C}$), which ensure to stabilize for the optimization and analysis procedures.

Experimental design: Lettuce (*Lactuca sativa* L.) seeds were purchased from the Field Crops Research Institute (FCRI), Vietnam. Experimental lettuce seeds were sowed in moist soil for germination. After 7 days, similar seedlings of lettuces were selected and transferred to the pots (60 cm \times 40 cm \times 45 cm, length \times width \times height) containing fertilizers, with nine plants per box. Lettuces were sprayed by GA₃ at different concentration of 0, 10, 30, 50, and 100 mg L⁻¹, respectively, at three dates after transplanting (the 7-day, the 14-day, and the 21-day). GA₃ was dissolved in tap-water. All experiments were repeated in three replications and carried out from February to March, 2019 in the greenhouse of Vietnam Academy of Science and Technology with the temperature of $25 \pm 0.5^\circ\text{C}$; and the air humidity of $70 \pm 0.5\%$.

Determination of metal concentrations in the experimental soil: The soil samples (1.5 g) were prepared in a well-labelled 100 mL polytetrafluoroethylene (PTFE) Teflon bombs and diluted with 6 mL (98%) HNO₃, 3 mL (35%) HCl and 0.25 mL (30%) H₂O₂ solutions. The digestion procedure was done in 30 min using the milestone ETHOS 900 microwave labstation (INSTR: MLS-1200 MEGA) (Tiimub and

Darley, 2015; Adama *et al.*, 2016). The samples were then centrifuged at 4000 rpm for 10 min, then filtered through 0.22 μm pore size PTFE membranes and diluted with deionized water before injection into the ICP-MS system to determine metal concentrations such as: Fe, Mn, Sn, Pb, Cd, Se, Hg and Cr (Rui *et al.*, 2008a and 2008b). All experiments were repeated in three replications.

Measurement of plant height, leaf number, and fresh weight of lettuces: The height of plants and leaf number of lettuces were determined at the first day and the days after transplanting (DAT) includes the seventh, the fourteenth, the twenty-first and the thirty-five days, respectively, and before GA₃ spraying. The height of lettuces was measured from the growing point to cotyledon node while leaf number was calculated by counting leaf by leaf. After treating with GA₃ for 35 days, lettuce plants were harvested (removal of roots), and then were immediately weighted for fresh weight.

Measurement of nutrient contents in lettuces: The shoots were dried at 65°C for 24-36 hour until constant weight by a fan-forced oven, and then were ground to a fine powder using a high-speed pulverizer to determine nutrient contents. About 30 mg of lettuces were treated by 6 mL HNO₃ (98%) for 24h. Then 3 mL H₂O₂ was added into each sample before digesting at 180°C for 4-5 h until 1 mL solution remained by electrical plate. The residue was diluted with deionized water to 10 mL, nutrient contents were determined using inductively coupled plasma mass spectrometer (The Agilent 7900 ICP-MS).

Determination of gibberellic acid GA₃ accumulated in lettuces: For determination of GA₃ concentration in lettuces, samples were collected at the dates, including the beginning transplanting, the date 7, the date 14th, the date 21th, and the date 35th (harvest day). Approximately 300 mg of lettuce samples (fresh and fine powder) were treated by 3,000 μL of 2-propanol/water/concentrated HCl (2:1:0.002, v/v/v) solution, then shaken at 4°C for 30 min. Afterwards, 6,000 μL of dichloromethane was added and agitated for another 30 min at 4°C before centrifugation at 4,000 rpm for 10 min. There were two phases forming after centrifugation, and lower layer was concentrated by the nitrogen flow for 1 hour, and then dissolved in 1 mL MeOH. The samples were then filtered through 0.22 μm pore size PTFE membranes before injection and analyzed by the LC-ESI-MS/MS system (Qingfeng *et al.*, 2014).

Data analysis: The one-way analysis of variance (ANOVA) was applied to analyze the data using SPSS 22.0 software. The means are averaged from three replicates and the standard derivations with a confidence interval of 95% ($p < 0.05$) in all cases.

RESULTS

The mean concentrations of metals in the experimental soil: The mean concentrations of metals (Fe, Mn, Sn, Pb, Cd, Se, Hg and Cr) in the experimental soil were shown in Table 1. Fe was the highest among the metals with a mean value of 63.2736 (mg/kg) recorded in the study soil. The second position belonged to the mean concentration of Mn, at 25.2431 (mg/kg). The

concentrations of Pb and Cr elements ranged in the third and fourth positions, at 2.4314 (mg/kg) and 2.1674 (mg/kg), respectively. Particularly, Sn concentration was recorded in 1.0565 (mg/kg) whereas the mean values of Cd, Se and Hg concentrations in the experimental soil were 0.2277 (mg/kg); 0.0956 (mg/kg) and 0.0897 (mg/kg), respectively. This indicated that there was a difference in metal contents in the soil using for experiments.

Table 1. Metal concentrations in the experimental soil.

	Fe	Mn	Sn	Pb	Cd	Se	Hg	Cr
Concentrations (mg/kg)	63.2736	25.2431	1.0565	2.4314	0.2277	0.0956	0.0897	2.1674

The effect of GA₃ on the growth of *Lactuca sativa* L.: Figure 1 illustrates the influence of GA₃ on the growth of lettuces via plant height and the number of leaves during the 35 days of experiment. It is interesting to note that the plant height was obviously lower in plants treated with 100 (mg L⁻¹) during the experimental process in comparison with those of the controls and the lower concentrations of GA₃. No significant difference was found in the plant heights of lettuces between the control and the treatments in 10, 30 and 50 (mg L⁻¹) GA₃ at 7 DAT. Nevertheless, the effects of GA₃ concentrations on the plant height were clearly determined from the 14 DAT. It is obviously from the Figure 1A, the plant heights of lettuces exposed to 10, 30 and 50 (mg L⁻¹) GA₃ concentrations were higher than those of the control and plants exposed with 100 (mg L⁻¹) GA₃ at 14 DAT and 21 DAT. Particularly, there was no significant different between the lettuces treated with 10, 30 and 50 (mg L⁻¹) GA₃ at 14 DAT as at 21 DAT. After 35 days of experiments, the plant heights were 30.1 cm, 29.5 cm, and 27.5 cm in the plants treated with the concentrations of 30, 10, and 50 (mg L⁻¹) GA₃, respectively, which were taller than those of the controls (25.2 cm) while the heights of lettuces in the treatment of 100 mg L⁻¹ GA₃ were 14.6 cm, were lower than those of the controls.

It is clear from Figure 1B, there were no differences in the leaf numbers of lettuces between the GA₃ treatments and the control until the 14 DAT. The influences of GA₃ exposures on the leaf number were significant at 21 DAT, wherein, the numbers of leaves of plants treated with 10, 30, and 50 (mg L⁻¹) GA₃ were higher than the 100 (mg L⁻¹) GA₃ treatment and the control. Nevertheless, no considerable differences were found in the leaf number between the other three GA₃ exposures. The highest numbers of leaves were 9.28 (leaves/plant) in the plants exposed to 30 (mg L⁻¹) GA₃, followed by 8.76 leaves/plant and 8.53 leaves/plant in the treatments of 10 and 50 (mg L⁻¹), respectively, which were significantly higher than the leaf numbers of the

controls (7.67 leaves/plant) and the 100 (mg L⁻¹) GA₃ treatment. This demonstrate that low concentrations of GA₃ promote an increase in the leaf number of lettuces, whereas high concentrations of GA₃ constrain the development of lettuce leaves.

The noticeable differences were reported in the fresh weights of *Lactuca sativa* L. between the control and GA₃ treatments over 35 experimental days. As can be seen from the Figure 2, the highest fresh weight of lettuces was 30.08 (g/plant) at the 30 (mg L⁻¹) GA₃ treatment, which was 37.67% higher than the fresh weight of the controls (18.75 g). The following were 29.54 g/plant and 21.50 g/plant in plants treated with 10 and 50 (mg L⁻¹) GA₃, respectively, which were 36.53% and 12.79% higher than the lettuces without exposure of GA₃, respectively. Particularly, a figure of 14.58 g/plant was obtained in the fresh weight of lettuces in the treatment of 100 mg L⁻¹ GA₃, which were 22.24% lower than those of the controls. The results illustrated that different concentrations of GA₃ showed the different influences on the fresh weight of lettuces, wherein, the 30 mg L⁻¹ GA₃ is considered as the most suitable concentration for the growth of lettuces.

The effect of GA₃ on the nutrient contents and heavy metals in *Lactuca sativa* L.: Table 1 shows the influence of GA₃ application on nutrients (Fe and Mn) and heavy metals (Sn, Pb, Cd, Se, Hg, and Cr) in lettuces after 35 days of experiments. There was a noticeable difference in the nutrient contents of lettuces between control and GA₃ treatments. Obviously, the contents of Fe in lettuces treated with 10, 30, 50, and 100 (mg L⁻¹) of GA₃ concentrations were fluctuated in the scope of 0.0014-0.0090 (mg kg⁻¹), were significantly lower than those of the controls (33.1813 mg kg⁻¹). Similarly, Mn contents in plants sprayed by GA₃ concentrations (10-100 mg L⁻¹) were changed between 0.0001 and 0.0025 (mg kg⁻¹), which were smaller than the lettuces without GA₃ treatments (13.9890 mg kg⁻¹). It is interesting to note that, the contents of nutrients such as Fe and Mn were

decreased with the increase of GA₃ concentrations from 10 to 100 (mg L⁻¹).

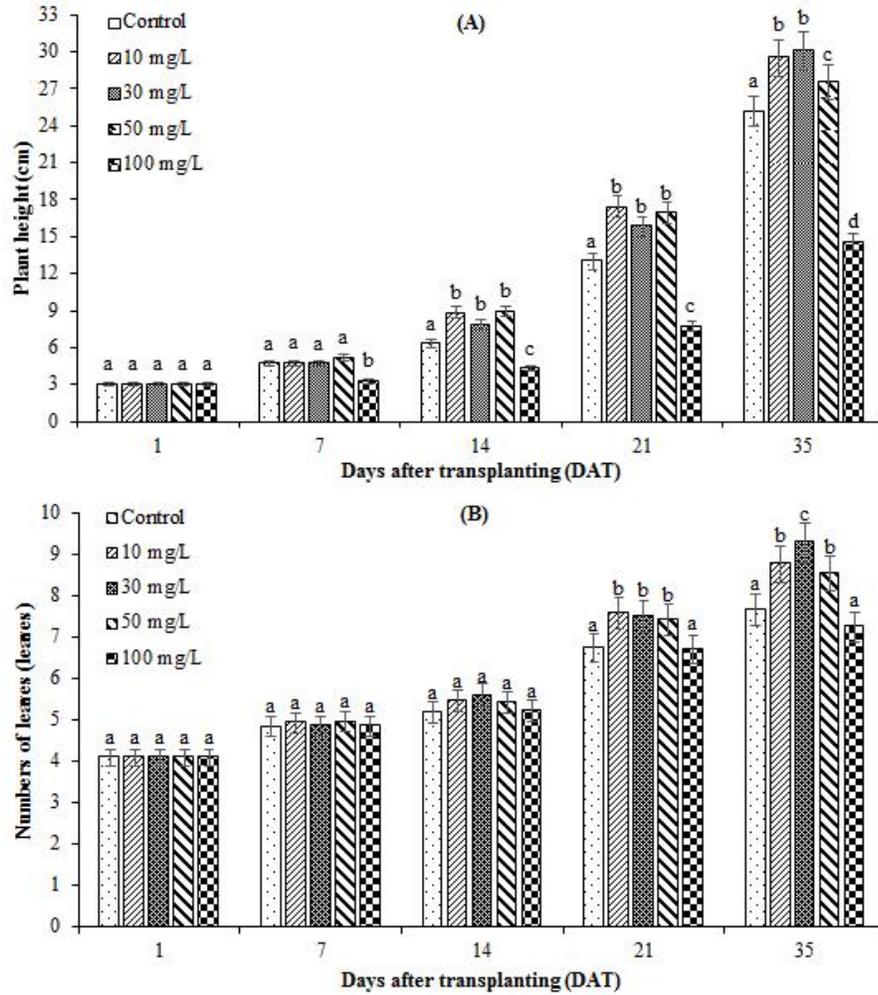


Figure 1 – The effect of GA₃ on the height (A) and leaf numbers (B) of *Lactuca sativa* L. (Different letters on the same DAT show significant differences at p < 0.05 level between GA₃ treatments and control)

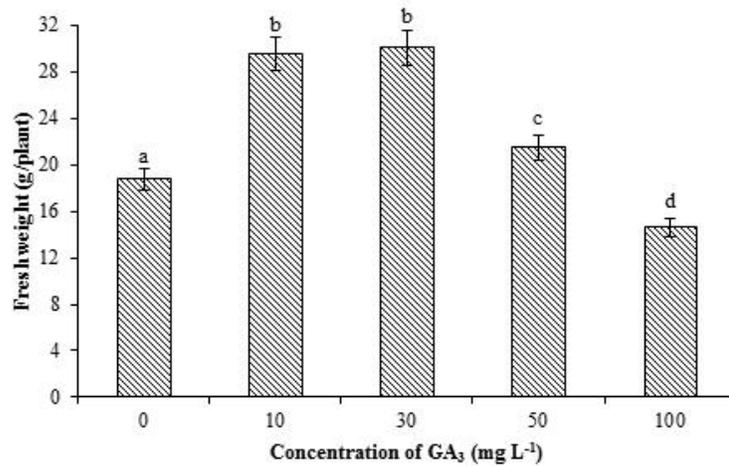


Figure 2 – Effects of GA₃ on the fresh weight of *Lactuca sativa* L. (Different small letters indicate significant differences at p < 0.05 level between GA₃ treatments and control)

Table 1 – The effect of GA₃ application on the contents of nutrients and heavy metals in *Lactuca sativa* L.

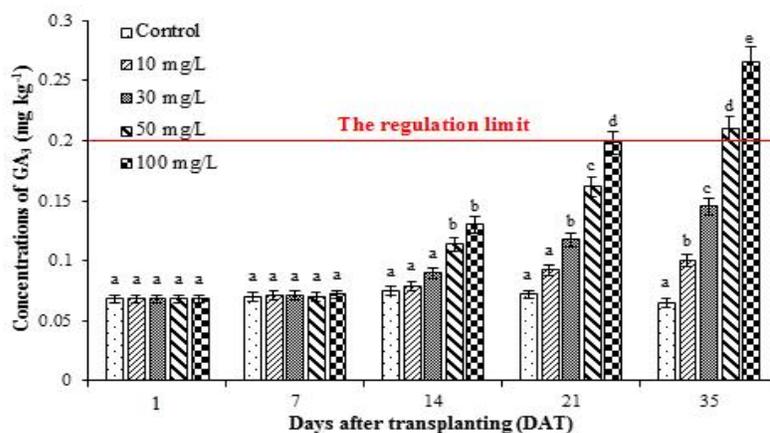
GA ₃ (mg L ⁻¹)	Fe (mg kg ⁻¹)	Mn (mg kg ⁻¹)	Sn (mg kg ⁻¹)	Pb (mg kg ⁻¹)	Cd (mg kg ⁻¹)	Se (mg kg ⁻¹)	Hg (mg kg ⁻¹)	Cr (mg kg ⁻¹)
Control	33.1813 ^a	13.9890 ^a	0.2729 ^a	1.6489 ^a	0.0812 ^a	0.0125 ^a	0.0130 ^a	0.5189 ^a
10	0.0090 ^b	0.0025 ^b	0.0045 ^b	0.0045 ^b	0.0005 ^b	0.0025 ^b	0.0060 ^b	0.0208 ^b
30	0.0080 ^b	0.0015 ^b	0.0020 ^b	0.0005 ^c	0.0002 ^b	0.0020 ^b	-	0.0194 ^b
50	0.0069 ^b	0.0008 ^b	0.0015 ^b	-	-	0.0010 ^b	-	0.0169 ^b
100	0.0014 ^c	0.0001 ^b	0.0001 ^b	-	-	0.0001 ^b	-	0.0108 ^b

(Different small letters on the same column present the significant difference at $p < 0.05$ level between GA₃ treatments and control; “-”: none detection)

As can be seen from the Table 1, the considerable difference was reported on the contents of heavy metals namely Sn, Pb, Cd, Se, Hg, and Cr in lettuces (*Lactuca sativa* L.) exposed to GA₃ treatments in comparison with those of the controls. The results showed that Sn, Se, and Cr contents in lettuces sprayed with GA₃ treatments were decreased among with the increase in concentrations of GA₃ application, and were remarkable lower than those of the controls. Se contents in lettuces exposed to GA₃ were changed between 0.0001 and 0.0025 (mg kg⁻¹), were 80-99% lower than those of the controls (0.0125 mg kg⁻¹). The Cr contents in lettuces at the GA₃ treatments fluctuated from 0.0108-0.0280 (mg kg⁻¹), was 96-98% lower than the plants without GA₃ (0.5189 mg kg⁻¹). Sn contents were found at 0.0001-0.0045 mg kg⁻¹, were 99% lower than those of the control. It is interesting to note that, Pb and Cd contents in lettuces were decreased by 10 and 30 (mg L⁻¹) of GA₃ treatments, however, these heavy metals were not found in lettuces treated with 50 and 100 (mg L⁻¹) of GA₃. Particularly, the contents of Hg were found at 0.0060 mg kg⁻¹ in the 10 (mg L⁻¹) of GA₃ treatment which were 53.9% lower than those of the controls (0.0130 mg kg⁻¹), but were without appears in plants treated with higher concentrations of GA₃ (30, 50 and 100 mg L⁻¹). These results illustrate that the decrease in the content of heavy metals in lettuces exposed with GA₃ in comparison with those of the control.

The bio-accumulation of GA₃ in *Lactuca sativa* L.:

Figure 3 reveals the GA₃ contents in lettuces after 35 days exposed to plant growth regulator (GA₃). No significant differences were reported in the concentrations of GA₃ in lettuces between the control and GA₃ treatments at the first transplanting and 7 DAT. As can be seen from the Figure 3, GA₃ contents in lettuces were increased among with the time of experiment from the 14 DAT to the 35 DAT. The contents of GA₃ were found at 0.079; 0.090; 0.114, and 0.130 (mg kg⁻¹) in the lettuces treated with 10, 30, 50, and 100 (mg L⁻¹) GA₃, respectively, which were higher than those of the controls (0.075 mg kg⁻¹) at the 14 DAT. Obviously, GA₃ contents were increased to 21.7%; 38.5%; 55.56%, 63.6% in plants exposed to GA₃ treatments including concentrations such as 10, 30, 50, and 100 (mg L⁻¹) in comparison with those of the controls at the 21 DAT. Particularly, after 35 days of experiments, the highest content of GA₃ was 0.265 (mg kg⁻¹) in lettuces exposed with 100 (mg L⁻¹) GA₃; following, was 0.210 (mg kg⁻¹) upon 50 (mg L⁻¹) GA₃ treatment; then, were 0.145 and 0.100 (mg kg⁻¹) in the exposures of 30 and 10 (mg L⁻¹) GA₃, respectively; and, the last was 0.065 (mg kg⁻¹) in the control. These data illustrated that the higher concentrations of GA₃ treatment use, the more their accumulations in lettuce samples.

**Figure 3 – The remained content of GA₃ accumulating in *Lactuca sativa* L.**

(Different small letters on the same DAT show significant differences at $p < 0.05$ level between GA₃ treatments and control)

DISCUSSION

The application of GA₃ on lettuces (*Lactuca sativa* L.) were reported in the previous studies. Georgeo *et al.* (2014) indicated that, lettuces were increased in plant heights, leaf numbers, and the fresh and dry weights under GA₃ treatments. According to Taslima (2015), the tallest plant heights of lettuces were 18.16 cm found in the treatment of 50 mg L⁻¹ GA₃ whereas the shortest of lettuces were reported in those of the control (15.57 cm). However, the highest yield of lettuces was 29.05 tons/ha in plants treated with 25 mg GA₃ while those of the control was only 16.27 tons/ha. In the other study, Alessandro *et al.* (2019) evaluated the effects of GA₃ on lettuces via the growth and yield parameters. Different concentrations of 0, 10⁻⁸, 10⁻⁶, and 10⁻⁴ (M) GA₃ were added into the floating system's mineral solution and were tested for 22 days. The lettuces were stimulated in the growth and were enhanced in the yield upon the exposures of 10⁻⁸ and 10⁻⁶ M GA₃ concentrations, especially, the highest leaf number per lettuce was obtained by treated to 10⁻⁶ (M) GA₃. Our data are in agreement with these studies that gibberellic acid (GA₃) was obviously impacted on the growth of lettuces, wherein, low concentrations of GA₃ was positive affected on the plant height, leaf number and biomass of lettuces. This is the basic to choose the suitable concentration of GA₃ applying to increase the productivity and efficiency in other vegetables and agricultural crop production.

In addition, several reports were studied the effects of GA₃ on the nutrients as well as the accumulation of heavy metals in agricultural plants. Akman (2009) indicated that the highest contents of Fe and Mn in wheat were found in the treatment without GA₃, whereas GA₃ application increased Fe and Mn contents in barley compared to the control. In the other study, Mukhtar (2008) found that 100 (mg L⁻¹) of GA₃ decreased Fe and Mn contents in *H. sabdariffa* in comparison with those of the controls. However, according to Hassan *et al.*, (1979), GA₃ was reported as the plant growth regulator enhancing to uptake of nutrients as Fe and Mn in corn plants. These data indicated that Fe and Mn contents in different plants were different impacted by GA₃ application, especially, the decrease of Fe and Mn contents in lettuces in comparison with those of the controls, which need further study to limit the impacts on the quality of vegetables.

The accumulation of Pb and Cd were found in lettuces (*Lactuca sativa* L.) watered with the wastewater from the Quetta city (Abdul *et al.*, 2011). According to Mamdouh and Osam (2018), Pb and Cd were uptaken and accumulated by lettuces grown on a metal-contaminated soil. Chang *et al.* (2013) reported lettuces uptake and accumulate heavy metals, namely Cd, Hg, Pb, and Cr in leaves from the soils collected in the Pearl river Delta, South China. These data indicate that lettuces

could uptake and accumulate heavy metals in their leaves, which associated with potential health risks to human. However, our study results showed that Sn, Pb, Cd, Hg, Se, and Cr were presented in low concentrations in lettuces exposed to GA₃ compared with those of the plants without GA₃ treatment while the same properties of soil present in Table 1 using for experiments. It might the application of GA₃ decrease the uptake of heavy metals in lettuces, which is needed to further investigate to limit the potential risks impact on consumers and ecological environment as well as to study and application of GA₃ in limiting heavy metals' absorption in other vegetables and agricultural crops.

The residues of GA₃ in fruits and vegetables might be the potential risks influence to consumers' health (Gehan *et al.*, 2015). According to the Maximum Residue Limits (MRLs) of the EU, the safety limit of gibberellin residues in vegetables was 0.20 mg kg⁻¹ (EU, 2010). In addition, several countries, such as Japan and U.S.A have regulated that a content of 0.20 mg kg⁻¹ was considered as the maximum residue limit of gibberellins in vegetables (Yuan and Xu, 2002). This illustrated that the residues of GA₃ in lettuces treated with high concentrations of GA₃ (50 and 100 mg L⁻¹) were higher than the regulations of the EU, U.S.A and Japan. On the other hand, some researchers indicated that the application of agrochemicals, especially, plant growth regulators might be lead to environmental pollution, serious affect to human health and animals (Celik and Kara, 1997; Sørensen and Danielsen, 2006; Ismail *et al.*, 2007; Erin *et al.*, 2008). This suggests that high concentrations of GA₃ (50 and 100 mg L⁻¹) were applied in vegetable production cause the high ratios of GA₃ contents accumulate on vegetables, which necessary to further study for improving the efficiency of vegetable production using plant growth regulator (GA₃) as well as minimize their impacts on human health and the ecological environment.

Conclusions: In summary, GA₃ had significant effect on the growth, nutrient elements, and the absorption of heavy metals in lettuce (*Lactuca sativa* L.). Low concentrations of GA₃ (10 and 30 mg L⁻¹) increased the plant height, leaf number and fresh weight of lettuces whereas the opposite results were reported in high concentration of GA₃ treatment (100 mg L⁻¹). We found that nutrient contents (Fe and Mn) in lettuces treated with GA₃ were significant decreased in comparison with those of the controls. In addition, the treatment of GA₃ reduced the absorption of heavy metals (Sn, Pb, Cd, Se, Hg, and Cr) in lettuces, which may limit the potential risk impacts on human health. However, the higher concentrations of GA₃ treatment were used, the more of its accumulation was found in lettuce samples. High contents of GA₃ were found in lettuces treated with 100 and 50 (mg L⁻¹) GA₃ (0.265 and 0.210 mg kg⁻¹, respectively), which were

higher than the regulations of the EU, U.S.A and Japan, and might be affected on the health of consumers. To the best of our knowledge, this is the first investigation on the influences of plant growth regulator (GA₃) on nutrient contents and heavy metals' absorption, and the accumulation of GA₃ applying in high concentrations in lettuces. However, further studies are needed for improving the efficiency of vegetable production using plant growth regulator as well as minimize their impacts on consumers' health and the ecological environment.

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REFERENCES

- Abdul, K.K.A., A.B. Zahoor and A.K. Safdar (2011). Accumulation of heavy metals by lettuces (*Lactuca sativa* L.) irrigated with different levels of wastewater of Quetta city. *Pakistan J. Bot.* 43(6): 2953-2960.
- Adama, M., R. Esena, B. Fosu-Mensah and D. Yirenya-Tawiah (2016). Heavy metal contamination of soils around a hospital waste incinerator bottom ash dumps site. *J. Environ. Public Health.* Article ID 8926453. doi:10.1155/2016/8926453.
- Akman, Z. (2009). Effects of plant growth regulators on nutrient contents of young wheat and barley plants under saline conditions. *J. Anim. Vet. Adv.* 8(10): 2018-2021.
- Alessandro, M., M. Alessandra, S. Leo and V. Filippo (2019). Effect of Gibberellic Acid on Growth, Yield, and Quality of Leaf Lettuce and Rocket Grown in a Floating System. *Agronomy* 9: 382. doi:10.3390/agronomy9070382.
- Çelik, A. and M. Kara (1997). The effects of plant growth regulators on activity of eight serum enzymes in vitro. *J. Environ. Sci. Health. A – Environ. Sci. Eng. Toxicology.* 32(6): 1755-1761.
- Chang, C.Y., H.Y. Yu, J.J. Chen, B.F. Li, H.H. Zhang and P.C. Liu (2013). Accumulation of heavy metals in leaf vegetables from agricultural soils and associated potential health risks in the Pear river Delta, South China. *Environ. Monit. Assess.* 186(3).
- Emongor, V.E. (2002). Effect of benzyladenine and gibberellins on growth, yield and yield components of common bean (*Phaseolus vulgaris*). *UNISWA Res. J. Agri. Sci. Technol.* 6: 65-72.
- Erin, N., B. Afacan, Y. Ersoy, F. Ercan and M.K. Balci (2008). Gibberellic acid, a plant growth regulator, increases mast cell recruitment and alters Substance P levels. *Toxicology* 254(1, 2): 75-81.
- EU Pesticides Database (2010). <http://ec.europa.eu/sanco/pesticides/public/index.cfm?event=substance.selection&ch=1>
- Fernandez, E., C. LaVecchia, B. Davanzo, E. Negri and S. Franceschi (1997). Risk factors for colorectal cancer in subjects with family history of the disease. *Br. J. Cancer* 75(9): 1381-1384.
- Gehan, A.H., S.A. Shreen and E.M. Samira (2015). Evaluation of synthetic plant growth regulators residues in fruits and vegetables and health risk assessment in Giza, Egypt. *J. Soil. Sci. Agri. Eng., Mansoura University* 6(9): 1075-1089.
- George, T., A.P. Spyridon and M.K. Ebrahim (2014). Effect of GA₃ and nitrogen on yield and marketability of lettuce (*Lactuca sativa* L.). *Aust. J. Crop. Sci.* 8(1): 127-132.
- Hassan, H.M., N.F. Kheir, Y.H. El-Shaby and A.A. Ibrahim (1979). The accumulations some minerals in corn plants as affected by GA₃ and micronutrient. *Ann. Agri. Sci. Moshtohor. J.* 6: 159-166.
- Ismail, C., T. Yasin and I. Ismail (2007). Evaluation of toxicity of abscisic acid and gibberellic acid in rats: 50 days drinking water study. *J. Enzyme Inhib. Med. Chem.* 22(2): 219-226.
- Khan, I., C.N. Tango, S. Miskeen, B.H. Lee and D.H. Oh (2017). Hurdle technology: A novel approach for enhanced food quality and safety: A review. *Food Control* 73: 1426-1444.
- Khan, M.M.A., C. Gautam, F. Mohammad, M.H. Siddiqui, M. Naeem and M.N. Khan (2006). Effect of gibberellic acid spray on performance of tomato. *Turk. J. Biol.* 30: 11-16.
- Khan, N.A. and M. Samiullah (2003). Comparative effect of modes of gibberellic acid application on photosynthetic biomass distribution and productivity of rapeseed-mustard. *Physiol. Mol. Biol. Plants.* 9: 141-145.
- Kim, M.J., Y. Moon, J.C. Tou, B. Mou and N.L. Waterland (2016). Nutritional value, bioactive compounds and health benefits of lettuce (*Lactuca sativa* L.). *J. Food Compost. Anal.* 49: 19-34.
- Le, G.M., B. Schraauwers, I. Larrieu and J.J. Bessoule (2008). Development of a biomarker for metal bioavailability: The lettuce fatty acid composition. *Environ. Toxicol. Chem.* 27(5): 1147-1151.
- López, A., G. Javier, J. Fenoll, P. Hellín and P. Flores (2014). Chemical composition and antioxidant capacity of lettuce: Comparative study of regular-sized (Romaine) and baby-sized (Little

- Gem and Mini Romaine) types. *J. Food. Compost. Anal.* 33: 39-48.
- Lynch, M.F., R.V. Tauxe and C.W. Hedberg (2009). The growing burden of foodborne outbreaks due to contaminated fresh produce: Risks and opportunities. *Epidemiol. Infect.* 137: 307-315.
- Maggio, A., G. Barbieri, G. Raimondi and S. DePascale (2010). Contrasting effects of GA₃ treatments on tomato plants exposed to increasing salinity. *J. Plant. Growth. Regul.* 29: 63-72.
- Mamdouh, A.E. and E.N. Osama (2018). Heavy metals uptake and translocation by lettuce and spinach grown on a metal-contaminated soil. *J. Soil. Sci. Plant. Nutr.* 18(4): 1097-1107.
- Mukhtar, F.B. (2008). Effect of Some Plant Growth Regulators on the Growth and Nutritional Value of *Hibiscus sabdariffa* L. (Red sorrel). *Int. J. Pure. Appl. Sci.* 2(3): 70-75.
- Pereira, C., D. Li and A.J. Sinclair (2001). The alpha-linolenic acid content of green vegetables commonly available in Australia. *Int. J. Vitam. Nutr. Res.* 71(4): 223-228.
- Qingfeng, N., Z. Yu, Q. Minjie, Y. Fengxia and T. Yuanwen (2014). Simultaneous quantitative determination of major plant hormones in pear flowers and fruit by UPLC/ESI-MS/MS. *Anal. Methods* 6: 1766-1773.
- Rui, Y.K., L.C. Qu and X.P. Kong (2008a). Effects of soil use along Yellow River basin on the pollution of soil by heavy metals. *Spectrosc. Spect. anal.* 28(4): 934-936.
- Rui, Y.K., J.B. Shen and F.S. Zhang (2008b). Application of ICP-MS to Determination of Heavy Metal Content of Heavy Metals in Two Kinds of N Fertilizer. *Spectrosc. Spect. Anal.* 28(10): 2425-2427.
- Shah, S.H. (2007). Effects of salt stress on mustard as affected by gibberellic acid application. *Gen. Appl. Plant. Physiol.* 33: 97-106.
- Shah, S.H., I. Ahmad and M. Samiullah (2006). Effect of gibberellic acid spray on growth, nutrient uptake and yield attributes during various growth stages of black cumin (*Nigella sativa* L.). *Asian. J. Plant. Sci.* 5: 881-884.
- Soerjomataram, I., D. Oomen, V. Lemmens, A. Oenema, V. Benetou, A. Trichopoulou, J.W. Coebergh, J. Barendregt and E. de Vries (2010). Increased consumption of fruit and vegetables and future cancer incidence in selected European countries. *Eur. J. Cancer* 46: 2563-2580.
- Sørensen, M.T. and V. Danielsen (2006). Effects of the plant growth regulator, chlormequat, on mammalian fertility". *Int. J. Androl.* 29(1): 129-33.
- Sunil, P., J. Tarun, O.P. Singh, R. Neelesh, M. Rishikesh and P.K. Jain (2015). Plant growth regulators in vegetable production: an overview. *Plant. Arch.* 15(2): 619-626.
- Taslina, A. (2015). Effect of gibberellic acid and spacing on growth and yield of lettuce (*Lactuca sativa* L.L.). Master Thesis, Department of Horticulture, Sher-e-bangla agricultural university. Dhaka-1207.
- Tiimub, B.M. and Dartey (2015). Determination of selected heavy metals contamination in water from downstream of the Volta Lake at Manya Krobo district in eastern region of Ghana. *Int. Res. J. Public. Environ. Health.* 2(11), 167-173.
- USDA (2004). U.S. Department of Agriculture, Agricultural Research Service, National nutrient data base for standard Reference, Release 17. <http://www.nal.usda.gov/fnic/foodcomp/Data/SR17/sr17.html>.
- Yuan, L. and D.Q. Xu (2002). Stimulatory Effect of Exogenous GA₃ on Photosynthesis and the Level of Endogenous GA (1+3) in Soybean Leaf. *Acta. Photophysiol. Sin.* 28: 317-320.